RESEARCH ARTICLE

Epilepsia

Genetic risk factor identification for common epilepsies guided by integrative omics data analysis

Correspondence

Herbert Schulz, Department of Microgravity and Translational Regenerative Medicine, Medical Faculty, University Hospital Magdeburg, Otto von Guericke University, 39106 Magdeburg, Germany.

Email: herbert.schulz@med.ovgu.de

Funding information

The German Research Foundation (DFG) and the Luxembourg Fond Nationale de Recherche (FNR) as a part of the research unit FOR-2715, Grant/Award Number: INTER/DFG/21/16394868 MechEPI2, Le1030/16-2, NO755/13-1, BE2078/5-1, BE2078/10-2, NO755/6-1 and SHU3585/1.1

Abstract

Objective: Genetic generalized epilepsies (GGEs) comprise the most common genetically determined epilepsy syndromes, following a complex mode of inheritance. Although many important common and rare genetic factors causing or contributing to these epilepsies have been identified in the past decades, many features of the genetic architecture are still insufficiently understood. This study integrates genome-wide association study (GWAS) data from the International League Against Epilepsy Consortium on Complex Epilepsies with transcriptome-wide association studies to identify genes whose genetically regulated expression levels are associated with epilepsy.

Methods: To achieve this, we used multiple computational approaches, including MAGMA, a tool for gene analysis of GWAS data, and its derivatives E-MAGMA and H-MAGMA, to improve gene mapping accuracy by utilizing tissue-specific expression and chromatin interaction data. Furthermore, we developed ME-MAGMA to incorporate methylation quantitative trait loci data, providing insights into epigenetic factors.

Epilepsia. 2025;00:1–15. wileyonlinelibrary.com/journal/epi

¹Department of Microgravity and Translational Regenerative Medicine, Medical Faculty, University Hospital Magdeburg, Otto von Guericke University, Magdeburg, Germany

²Clinic for Plastic, Aesthetic, and Hand Surgery, Medical Faculty, University Hospital Magdeburg, Otto von Guericke University, Magdeburg, Germany

³Department of Engineering, Brandenburg University of Applied Sciences, Brandenburg, Germany

⁴Cologne Center for Genomics, University of Cologne, Cologne, Germany

⁵University Hospital Cologne, Medical Faculty, University of Cologne, Cologne, Germany

⁶Luxembourg Center for Systems Biomedicine, University of Luxembourg, Esch-sur-Alzette, Luxembourg

⁷Department of Epileptology, Eberhard Karls University, Tübingen, Germany

⁸Institute for Cellular Neurosciences II, Medical Faculty, University of Bonn, Bonn, Germany

⁹Department of Biomedicine, Faculty of Health, Aarhus University, Aarhus, Denmark

^{*}List of consortium members is provided in the Supplementary Material.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2025} The Author(s). Epilepsia published by Wiley Periodicals LLC on behalf of International League Against Epilepsy.



Results: We identified a total of 897 false discovery rate-corrected (<.05) candidates. These include voltage-gated calcium channels, voltage-gated potassium channels, and other genes such as *NPRL2*, *CACNB2*, and *KCNT1* associated with epilepsy pathogenesis that act as key players in neuronal communication and signaling in the brain.

Significance: In this study, we propose new candidate genes to expand the dataset of potential epilepsy-causing genes. Further research on these genes may enhance our understanding of the complex regulatory mechanisms underlying GGE and other types of epilepsy, potentially revealing targets for therapeutic intervention.

KEYWORDS

epigenetics, epilepsy, genetic generalized epilepsy, GWAS, ILAE, MAGMA, transcriptomics

1 | INTRODUCTION

Epilepsy is a common neurological disorder that affects millions of people worldwide, characterized by recurring, unprovoked seizures. It can affect individuals of all ages, impacting their quality of life. Although its causes vary from brain injuries to infections or genetics, research is increasingly focusing on genetic factors to improve diagnosis, treatment, and prevention.¹

Epilepsy is a broad term that includes a variety of disorders, which are categorized based on seizure types, age at onset, developmental status, comorbid features, and underlying etiologies. One specific type is genetic generalized epilepsy (GGE), which refers to a group of epileptic syndromes with a strong indication of a hereditary predisposition. It is characterized by generalized seizures, meaning the seizures involve bilateral hemispheric networks from the onset. In contrast, focal epilepsy (FE) is defined by seizures that originate from a specific, localized region within one cerebral hemisphere. ^{2,3}

Genome-wide association studies (GWAS) are a widely used genomic approach to identifying genetic variants that contribute to disease risk³ and allow identification of specific genomic regions that increase the risk of developing a particular phenotype or disease, providing insights into a disorder's underlying genetic architecture, including epilepsy. The largest GWAS on epilepsy so far was conducted by the International League Against Epilepsy (ILAE) Consortium on Complex Epilepsies, which performed a large-scale transethnic meta-analysis by combining GWAS data from populations of European, Asian, and African genetic ancestry. 4,5 In this study, 26 genome-wide significant loci were identified, of which 19 were specific to GGE. These loci collectively implicate 29 putative causal genes, such as CACNA2D2, KCNIP2, SCN1A, and RBFOX1, underlying the genetic architecture of epilepsy.^{4,5}

Key points

- A new method called ME-MAGMA is designed to identify genes influenced by epigenetic regulators associated with epilepsy.
- A multilayered approach is used, utilizing positional, transcriptional, and epigenetic information to identify epilepsy-associated genes.
- The genes identified through chromatin interaction profiles from both the adult and fetal datasets suggest some possible roles in developmental effects on neurological pathways.

Despite the large size of this GWAS, many features of the genetic architecture of epilepsy are still insufficiently understood. One way forward is the identification of genes whose genetically regulated expression levels are significantly associated with complex human diseases or traits, 6 as is the primary objective of transcriptomewide association studies (TWAS). Here, we aim to integrate GWAS data on individuals of European genetic ancestry generated by the ILAE Consortium on Complex Epilepsies with gene expression data from TWAS to identify such associations for epilepsy. To achieve this, we employed MAGMA⁷ and several of its derivatives. This widely used computational tool implements a multiple linear principal components regression model to derive gene-level p-values using GWAS summary statistics as input. It is essential to distinguish gene identification in conventional GWAS from MAGMA-based analyses. Single nucleotide polymorphism (SNP)-level GWAS associate individual variants with disease risk, often assigning them to nearby genes without implying functional involvement. In contrast, MAGMA aggregates

association signals across all SNPs within or near a gene and can integrate functional annotations such as gene expression, methylation, and chromatin interactions. This provides a more biologically informed view by highlighting genes potentially involved through regulatory or transcriptional mechanisms.

Besides MAGMA, we considered three derivatives that

Besides MAGMA, we considered three derivatives that allow the integration multilayered data to enhance analytical power and improve the signal-to-noise ratio, mainly by combining multiple genetic annotations and capturing diverse mechanisms underlying genetic associations with complex traits. 8,9 We developed ME-MAGMA, a novel adaptation that assigns risk variants to genes based on methylation quantitative trait (meQTL) data. 10 This approach enables the exploration of epigenetic mechanisms, particularly DNA methylation, as a key player in gene regulation, influencing disease susceptibility. To incorporate tissue-specific gene regulation, we used E-MAGMA, an extension of MAGMA that integrates expression quantitative trait loci (eQTLs) information, mapping risk variants to putative genes based on their transcriptional activity and expression levels. 11 This approach improves the interpretation of GWAS results by linking genetic variation to gene expression changes relevant to epilepsy and related traits. To further investigate the developmental gene regulation, we utilized Hi-C-coupled MAGMA (H-MAGMA), 12 which leverages Hi-C chromatin interaction data to map genetic variants to their target genes. 12

Through the integration of GWAS data, eQTL, chromatin interaction profiles, and methylation data, our multiomics analytical framework aims to identify and prioritize candidate genes related to epilepsy based on vetted hypotheses.

2 MATERIALS AND METHODS

2.1 Ethics statement

The datasets used were generated based on the following ethics statement by the ILAE Consortium on Complex Epilepsies^{4,5}: "We have complied with all relevant ethical regulations. All study participants provided written informed consent for the use of their data in genetic studies of epilepsy. For minors, written informed consent was obtained from their parents or legal guardians. Local institutional review boards approved study protocols at each contributing site."

2.2 Data source

For this study, we used the third iteration of GWAS summary statistics, referred to as ILAE3, produced by the ILAE

Consortium on Complex Genetics. These summary statistics, available online at the Epilepsy Genetic Association Database website (https://www.epigad.org/), comprise 4.9 million SNPs tested for phenotypic association using mixed model meta-analysis in 52 538 controls and 29 944 individuals with epilepsy. In these summary statistics, epilepsy cases are classified into three major categories: FE, GGE, and unclassified epilepsy. The "all epilepsy" group comprises individuals from all three categories.⁵

Usually, the estimates based on reference data with similar ancestry to the data from which SNP p-values were computed yield higher accuracy results in MAGMA. Therefore, we used the previous version of the dataset, referred to as ILAE2, as a reference panel, provided by the ILAE Consortium on Complex Epilepsies in PLINK format (Table 1). This dataset included 38324 individuals of European ancestry, comprising 14130 epilepsy cases and 24194 controls. To ensure the data quality and reliability of these data, all ambiguous, nonautosomal variants and SNPs with a minor allele frequency of <.01 were excluded using PLINK 2.0. 14,15 After performing this quality control, the dataset comprised 5535915 variants. This dataset was used as a reference, because it is part of the ILAE3 dataset, whose summary statistics were used for the analysis.

2.3 Magma

MAGMA gene analysis is performed to quantify the degree of association each gene has with the phenotype. It employs an SNP-wise model that first analyses individual SNPs within a gene using GWAS summary statistics. It then combines the resulting SNP p-values into a gene- or region-level test statistic, calculated as the mean χ^2 statistic, where a gene-level p-value is derived using a known approximation of the sampling distribution. To determine which SNP belongs to each gene, information on the gene and variant positions is needed. This information is provided by the user in the form of an annotation file. MAGMA also allows users to define a window around genes, which includes variants that are outside the gene itself but may still be relevant, such as those located within the promoter region and other regulatory areas. Additionally, MAGMA requires reference data to account for linkage disequilibrium, which in our case is the ILAE2 dataset. We used conventional MAGMA⁷ to investigate the association between common epilepsies and genetic variants located within the gene body, that is, the region of a gene that is transcribed into RNA, encompassing both exons and introns. For this, we generated an annotation file using the hg19 reference from the 1000 Genomes Project, ¹⁶ resulting in a dataset comprising



TABLE 1 Data sources.

Dataset name	Sample size	Description	URL
ILAE3	Individuals: 82482 Cases: 29944 Controls: 52538 SNPs GGE: 4866066 All: 4880199 Focal: 4861642	Third iteration of the GWAS dataset generated by ILAE Consortium ⁵ Summary statistics used for MAGMA input	https://www. epigad.org/ gwas_index. html
ILAE2	Individuals: 38 324 Cases: 14 130 Controls: 24 194 SNPs: 5 535 915	Second iteration of the GWAS dataset generated by ILAE Consortium on Complex Epilepsies in PLINK format ⁴ and used as reference data to account for LD for MAGMA input in this study	
meQTL dataset	meQTLs: 409 372	Dataset generated by Schulz et al. ¹⁰ Used for meQTL and CpG association information	
Illumina BeadChip dataset	cis-CPG: 25165 Genes: 9967	Dataset generated by Price et al. ¹³ using Illumina Infinium HumanMethylation450 BeadChip array Used for CPG and gene association information in this study	https://www. ncbi.nlm. nih.gov/geo/ query/acc.cgi? acc=GPL16304

Abbreviations: GGE, genetic generalized epilepsy; GWAS, genome-wide association studies; ILAE, International League Against Epilepsy; LD, linkage disequilibrium; meQTL, methylation quantitative trait; SNP, single nucleotide polymorphism.

34434987 SNPs mapped to 18524 genes, to be used with MAGMA.

2.4 E-magma

To explore the association between eQTL and epilepsy, we employed the E-MAGMA framework (https://github.com/eskederks/eMAGMA-tutorial), a derivative of the MAGMA software designed to integrate cis-eQTL data with genetic association studies. E-MAGMA enables the investigation of how genetic variants influence gene expression and, consequently, impact susceptibility to disease. This framework provides annotation information for 14 brain-related tissues collected from GTEx (version 8). However, to maintain consistent analysis conditions, we focused specifically on eQTL data from hippocampal tissue. The annotation file for hippocampal tissue comprised 294761 SNPs and 3474 genes.

2.5 ME-MAGMA

meQTLs are genetic variants that influence the variation in DNA methylation patterns across the genome. To investigate the relationship between these meQTLs and genes, we developed the ME-MAGMA (https://github.com/ashwini-mushunuri/ME-MAGMA) framework to identify genetically driven DNA methylation events

potentially involved in epileptogenesis. As the genomic coordinates linking meQTL changes in DNA methylation at specific CpG sites were not readily available, we constructed a custom MAGMA-compatible annotation file by integrating data from two primary sources.

The first dataset, referred to hereafter as the meQTL dataset, was an unpublished, unpruned dataset generated in one of our previous studies. 10 It was derived from fresh-frozen hippocampal tissue samples from 110 unrelated individuals with temporal lobe epilepsy (TLE) and comprises 984176 cis-meQTL SNP–CpG site pairs. Associations were identified using a false discovery rate (FDR) threshold of 5% within a cis window of $\pm 500\,\mathrm{kb}$ between SNP genotypes and quantitative methylation rate (β -value) of CpGs or 3'-RNA expression levels, using a linear regression model implemented in Matrix eQTL. 10

Thesecond dataset was obtained from the Gene Expression Omnibus (GEO) under accession ID GPL16304 and belongs to the Illumina Infinium HumanMethylation450 BeadChip dataset version 1.0 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GPL16304). This dataset includes CpG site identifiers and their nearest associated gene symbols. To enrich this information with standardized gene identifiers and genomic coordinates, we retrieved Ensembl gene IDs, Entrez gene IDs, chromosome names, and gene start and end positions for each gene listed in the Illumina BeadChip dataset using BioMart (GRCh37 build and Ensembl Genes version 114). 16

MUSHUNURI ET AL. Epilepsia 15

This query yielded 11092 Ensembl gene IDs corresponding to 9967 unique gene symbols. These Ensembl IDs were mapped to their respective CpG site IDs in the Illumina BeadChip dataset using R v4.3.2 and tidyverse v2.0. ^{18,19} With the CpG IDs as a common key, we subsequently linked the Ensembl gene information to the meQTL SNPs from the meQTL dataset, thereby establishing associations between cis-meQTLs and genes via their shared CpG sites.

This integrative mapping resulted in a dataset comprising 25 165 unique cis-CpG sites and 409 372 associated meQTL SNPs, leading to a total of 1350 219 SNP-CpG-gene associations. The final dataset was formatted into a MAGMA-compliant annotation file for downstream gene-based analysis, allowing for a precise investigation of meQTL-gene relationships.

2.6 H-MAGMA

To capture the dynamic changes in chromatin interactions that occur during brain development and to gain insights into the genetic factors contributing to epilepsy, we utilized H-MAGMA. The H-MAGMA annotation files, which comprise chromatin interaction data, were created by assigning exonic and promoter SNPs to their corresponding target genes based on their genomic locations, using the Gencode v26 (GRCh37) gene model. This gene model encompasses a total of 58 219 genes, which include protein-coding genes, small noncoding RNA genes, and various types of pseudogenes (including processed, unprocessed, unitary, and polymorphic pseudogenes).

The annotation information is available on GitHub (https://github.com/thewonlab/H-MAGMA) for two developmental stages—adult and fetal brain—as well as for three cell types: induced pluripotent stem cell (iPSC)-derived neurons, iPSC-derived astrocytes, and cortical neurons. However, in this study, we only utilized datasets from adult and fetal brain tissues, referred to as H-MAGMA Adult and H-MAGMA Fetal, respectively. The annotation information for adult brain tissue contains 3 943 744 SNPs and 54 341 genes and pseudogenes, and the fetal brain tissue annotation includes 3 978 004 SNPs and 54 332 genes and pseudogenes.

2.7 | Gene-level analysis

We conducted a comprehensive gene-level analysis using various regulatory genomic features described above to identify candidate genes that are associated with epilepsy. To ensure consistency in the computational parameters, we utilized MAGMA's "snp-wise=mean" mode for all

methods. This approach averages the effects of all SNPs associated with a gene, providing a more balanced and representative measure of their collective influence.

The resulting gene-level associations were FDR-adjusted and filtered using an FDR threshold of \leq .05 to ensure statistical robustness and minimize the risk of false positives. This analysis was conducted separately using the ILAE3 summary statistics for focal epilepsy, genetic generalized epilepsy, and "all epilepsy" (Figure 1).

2.8 Gene prioritization

To identify key candidates within this extensive list, we used VarElect,²⁰ which is a phenotype-based variation prioritization tool. It prioritizes genes based on their potential association with specific phenotypes, using factors such as pathway interactions and gene expression patterns. This approach enabled us to identify several critical genes as promising candidates (Table 2) for further investigation. The keyword "epilepsy" was used to associate the phenotype with the genes.

3 RESULTS

3.1 Genetic generalized epilepsies

In this gene-level association study using MAGMA and its extensions, we analyzed a total of 51 866 genes and pseudogenes. Pseudogenes are genomic DNA sequences resembling mutated or truncated versions of functional genes. Emerging evidence suggests that many pseudogenes are transcribed at nonrandom levels, with some transcripts exhibiting biological functions, warranting their classification as a subclass of functional long noncoding RNAs.²¹

After applying a 5% FDR correction, we identified 897 genes associated with GGE, as shown in Table 2. Notably, 18 of these genes had previously been implicated in SNP-level GWAS conducted by the ILAE Consortium on Complex Genetics, indicating a strong overlap between our findings and established genetic associations. The overlapping genes include AP3D1, ARHGEF15, ATXN1, CACNA2D2, CDK5RAP3, FANCL, GABRA2, KCNIP2, KCNN2, PCDH7, RBFOX1, RMI1, SCN1A, SCN2A, SPOCK1, STX1B, TMPRSS15, and PNPO. Furthermore, some overlap was observed across all calculations, as illustrated in Figure 2.

In the H-MAGMA Adult and H-MAGMA Fetal dataset analyses, we identified 413 and 489 genes and pseudogenes, respectively, 316 of which are common to both studies (see Supplemental Data 1). Among our candidate genes controlled for 5% FDR in both H-MAGMA and

FIGURE 1 Flowchart describing various dataset usage at different stages of methodology. DLPFC, dorsolateral prefrontal cortex; eQTL, expression quantitative trait locus; FDR, false discovery rate; ILAE, International League Against Epilepsy; meQTL, methylation quantitative trait.

TABLE 2 Number of associated genes and pseudogenes following 5% Bonferroni and FDR correction.

	FDR-corrected value			Bonferroni- corrected value		
Method	GGE	FE	All	GGE	FE	All
ME-MAGMA	101	1	9	16		1
E-MAGMA	51	1	16	14	1	1
MAGMA	236	2	29	35	1	6
H-MAGMA adult	534	0	38	60		8
H-MAGMA fetal	489	0	47	59		7

Abbreviations: FDR, false discovery rate; FE, focal epilepsy; GGE, genetic generalized epilepsy.

MAGMA analyses, we found *GNAI2*, *GNAT1*, *LSMEM2*, *RASSF1*, *SEMA3F*, *SLC38A3*, and *IFRD2*, which are located at locus 3p21.31 (see Supplemental Data 1).

In the ME-MAGMA analysis, we identified *AP3D1*, *HYAL2*, *NPRL2*, *RASSF1*, and *NAB1* as the candidates with the lowest *p*-values (see Supplemental Data 1). Among these, *NPRL2*, *HYAL2*, and *RASSF1* are located at locus 3p21.31 (Figure 3A), suggesting the potential influence of meQTLs in linking this locus to GGE.

Additionally, in Figure 3A, genes appear clustered on chromosome 17, suggesting a potential methylation-related

effect specific to this chromosome. This is further supported by the identification of several genes, such as *PER1* (17p13.1), *SP6* (17q21.32), and *MYCBPAP* (17q21.33), in the ME-MAGMA analysis. Similarly, the E-MAGMA analysis revealed additional candidate genes on chromosome 17, including *TVP23B*, *TRIM16L*, and *PRPSAP2* at 17p11.2. We also identified genes common to both E-MAGMA and H-MAGMA on chromosome 17, such as *CDK5RAP3* at 17q21.32, *VAMP2* at 17p13.1, and *SPATA20* at 17q21.33.

3.2 | Focal epilepsies

For FEs, our analysis suggests three genes that had not been previously reported in the ILAE Consortium studies.^{4,5} These genes include *FANCD2P2* and *CUL4A*, identified through MAGMA, and *PROZ*, identified through ME-MAGMA and E-MAGMA (Supplemental Data 1).

3.3 | All epilepsies

Using the "all epilepsy" summary statistics, we observed associations with 97 genes (Supplemental Data 1). Among these, five genes had already been recognized in the original study. ^{4,5} These genes include *KCNIP2*, *RMI1*, *SCN1A*, and *FANCL*, identified through H-MAGMA Adult,

FIGURE 2 (A) Venn diagram illustrating the intersection of results obtained from H-MAGMA, E-MAGMA, and ME-MAGMA, highlighting both shared and unique findings. (B) Venn diagram showing the overlap between the results from MAGMA and its extensions. The overlap includes genes associated with genetic generalized epilepsy at the 5% false discovery rate level for ME-MAGMA (n = 58), E-MAGMA (n = 28), H-MAGMA Adult (n = 85), and H-MAGMA Fetal (n = 164), respectively. Only four genes—DPM2, VAMP2, MP1, and MRDC1—were common across four analyses, and only MP1 and MP2 were common across all five analyses, with MP2 being the only one previously reported in the original study.

H-MAGMA Fetal, and MAGMA, as well as *SPOCK1*, identified through H-MAGMA Adult, H-MAGMA Fetal, MAGMA, and ME-MAGMA. Additionally, *ARHGEF15*, identified through H-MAGMA Fetal and MAGMA, was also observed in our analysis of GGE.

4 DISCUSSION

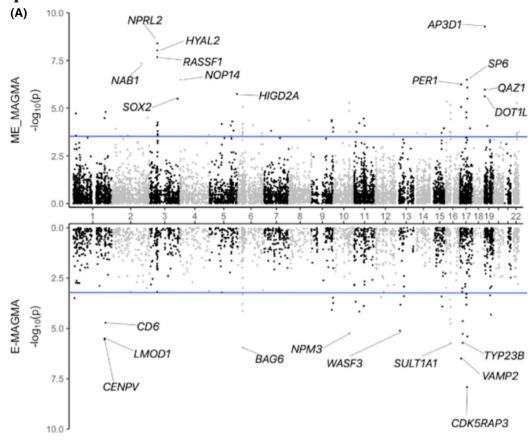
The etiology of epilepsy is complex, often involving multiple genetic variations that disrupt neural pathways and contribute to seizure susceptibility. Focusing on a single gene or pathway is not always sufficient to fully capture the genetic underpinnings of epilepsy. Accordingly, a comprehensive approach that considers the cumulative effects of multiple genetic variants is essential for understanding its genetic architecture.^{5,22} In this study, we present evidence-based hypotheses regarding the contribution of these variants to epilepsy risk. To investigate their impact, we used the MAGMA statistical framework and its derivatives, E-MAGMA, H-MAGMA, and ME-MAGMA, which integrate diverse datasets such as transcriptional profiles, chromatin interaction maps, and epigenetic marks, respectively, to identify genes influenced by these genetic variations. We analyzed chromatin interaction maps from both adult and fetal brains using H-MAGMA, following the findings of Sey et al., 12 which show that psychiatric and neurological disorders tend to

have greater heritability enrichment in the fetal brain than in the adult brain, especially for childhood onset disorders like attention-deficit/hyperactivity disorder and autism spectrum disorder, highlighting their neurodevelopmental origins. ¹² A similar integrative gene-level approach, using multiple variations of MAGMA, has been applied in a study of traumatic brain injury, ²³ which supports the validity of our multiomic approach to uncovering regulatory mechanisms underlying neurological disorders.

ME-MAGMA is a novel method for identifying epilepsy-associated meQTL SNPs, offering insights into the epigenetic mechanisms contributing to seizure susceptibility and supporting the development of targeted therapies using data from hippocampal tissues obtained from TLE patients. Using these data from annotation files in MAGMA is acceptable because these annotations are not used to determine case–control status. Instead, they help highlight genomic regions that are biologically active or relevant in the context of epilepsy.

When analyzing GWAS data for GGE, incorporating epigenomic marks derived from TLE patients can reveal whether GGE-associated genetic variants are enriched in regions of the genome that are active in epileptic brain tissue. This approach can provide valuable insights into the regulatory mechanisms that may be shared between TLE and GGE, or it may highlight distinct pathways involved in each epilepsy subtype. One gene of interest is *NPRL2*, which has already been associated with FE²⁴ but has not

15281167, 0, Downloaded from https://onlinelbtrary.wiley.com/doi/10.1111/epi.70021, Wiley Online Library on [30/11/2025]. See the Terms and Conditions (https://onlinelbrary.wiley.com/terms-and-conditions) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons License



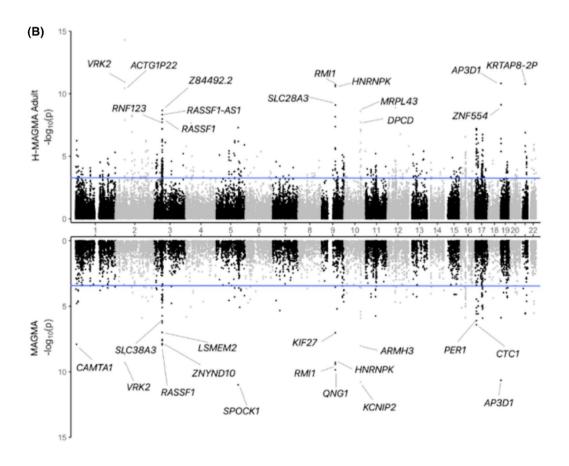


FIGURE 3 Miami plots illustrating *p*-values from gene-based analyses. In both panels, the blue lines mark the threshold corresponding to a 5% false discovery rate (FDR), and genes with the lowest FDR-corrected *p*-values are labeled. (A) ME-MAGMA and E-MAGMA analyses in hippocampal tissue (nominal *p*-value thresholds: 5.26×10^{-4} and 9.98×10^{-4} , respectively), with most top candidate genes clustered on chromosomes 3 and 17. (B) H-MAGMA Adult brain tissue and MAGMA analyses (nominal *p*-value thresholds: 5.75×10^{-4} and 7.19×10^{-4} , respectively), showing a notable clustering of top candidate genes on chromosome 3, suggesting a higher mutation rate that may contribute to epilepsy.

yet been linked to GGE. This suggests shared or distinct regulatory mechanisms of the same gene based on variations between the two types of epilepsy.

The value of integrating methylation data in epilepsy research is further supported by a study from Kobow et al., ²⁵ which used DNA methylomes and transcriptomes from focal cortical dysplasia (FCD) specimens and controls to distinguish major FCD subtypes from TLE and nonepileptic cases. ²⁵ These findings highlight the relevance of methylation in disease classification and mechanisms. Together, ME-MAGMA and similar epigenomic strategies underscore the potential of methylation-based analyses to deepen our understanding of different types of epilepsies.

4.1 | Multiple testing correction

Both the FDR and the Bonferroni correction are methods used to adjust for multiple testing when interpreting MAGMA results. FDR controls for the expected proportion of false positives among the significant results and is used to interpret gene sets. Bonferroni controls the probability of even one false positive. FDR is therefore the preferred method for our exploratory analysis, particularly for gene set overlap comparisons between different MAGMA derivatives (Figure 2). However, we have considered both correction methods in Table 2 in summary and Supplemental Data 1 in detail.

4.2 | Gene-based associations for genetic generalized epilepsies

4.2.1 | Voltage-gated calcium channels

Our MAGMA analysis found voltage-gated calcium channel (VGCC) genes, which are widely expressed in the central nervous system and regulate neuronal excitability and intracellular Ca^{2+} dynamics, to be associated with GGE. Calcium ions mediate key cellular processes such as excitation-transcription, excitation-contraction, and excitation-exocytosis coupling, especially in synaptic transmission. We found association between GGE and CACNA2D2 and CACNB2, which encode the $\alpha2\delta2$ and $\beta2$

VGCC subunits. *CACNA2D2* has been previously linked to GGE,⁵ and we reaffirm this connection in the current study. Additionally, *CACNA2D2* has been associated with monogenic forms of developmental and epileptic encephalopathy.^{27,28} On the other hand, *CACNB2* has not been associated with GGE specifically yet, but its upregulation is linked to TLE,²⁹ suggesting that calcium channel subunit dysfunction may contribute broadly to various epilepsy subtypes.

4.2.2 Voltage-gated potassium channels

Potassium channel activation plays a crucial role in repolarizing the cell membrane after excitation. Among the genes coding for voltage-gated potassium channels identified in this study, KCND3, KCNJ4, and KCNT1 were newly found to be associated with GGE, whereas KCNIP2 and KCNN2 have been described as potential candidates in prior ILAE consortium studies. 4,5 KCNT1, identified through H-MAGMA Adult, encodes the sodium-activated potassium channel subunit K_{Na}1.1. Gain-of-function variants in KCNT1 are linked to early onset epileptic encephalopathies, including early infantile developmental and epileptic encephalopathy (DEE), infantile epileptic spams syndrome, epilepsy of infancy with migrating focal seizures, and sleep-related hypermotor epilepsy. 30,31 KCND3, identified via MAGMA analysis, is associated with Dravetlike syndrome, DEE, intellectual disability, and cardiac arrhythmias. 32,33 KCNJ4, consistently identified across our GGE analyses, encodes the inwardly rectifying potassium channel subunit Kir2.3, which has been implicated in kainate-induced TLE through its upregulation.³⁴ Even though the monogenic syndromes associated with these genes are quite diverse, identification of these genes in our study suggests that common variants may contribute to the phenotype of GGE.

4.2.3 | Transmembrane protein-coding genes

Our GGE analysis identified several transmembrane protein-coding genes across different analytical frameworks. Notably, *TMEM107* was detected using both H-MAGMA and MAGMA, whereas *TMEM186* was identified



in H-MAGMA Fetal analyses. *TMEM115* and *TMPRSS15* emerged from MAGMA, and *TMEM177* was highlighted by E-MAGMA. Notably, *TMPRSS15* has previously been linked to GGE in an ILAE study.⁵

Mutations in *TMEM107* have been associated with ciliopathies such as Meckel syndrome and Joubert syndrome, conditions often accompanied by neurological abnormalities, including developmental delays and brain malformations.³⁵ Investigating this gene could provide novel insights into the role of chromatin interactions and their developmental effects on neurological pathways. *TMEM177* mutations are implicated in mitochondrial dysfunction, a key factor in various epilepsy subtypes, including infantile spasms, myoclonic epilepsy with ragged-red fibers, and TLE.^{36,37} Although *TMEM115* and *TMEM186* have not been directly linked to GGE in previous publications, the positioning of *TMEM115* at the 3p21.31 locus, known to harbor multiple epilepsy-related genes, suggests its potential relevance as a candidate gene for epilepsy.

4.2.4 | Zinc finger proteins

Zinc finger proteins (ZNFs) are involved in various cellular processes, including transcriptional regulation and DNA repair.³⁸ In this study, *ZBTB32*, *ZNF668*, and *ZNF629* were identified through E-MAGMA, H-MAGMA (Adult), and ME-MAGMA analyses, respectively.

ZBTB32 is linked to DEE 52,^{39,40} and deletions at its locus are associated with generalized epilepsy with febrile seizures plus (GEFS+) type 1. Mutations due to eQTLs in ZBTB32 can disrupt gene regulation, potentially contributing to GGE.

ZNF668 is crucial for DNA damage repair, with its deficiency leading to severe developmental issues and being linked to GEFS+ type 9.^{24,41} *ZNF629* encodes for a transcription factor regulating RNA polymerase II, with variants associated with intellectual developmental disorders and ⁴² juvenile myoclonic epilepsy. ⁴⁰ Epigenetic mutations in such factors can alter gene expression and contribute to seizure disorders.

These findings suggest a role for epigenetic and transcriptional dysregulation in the etiology of GGE and its subtypes.

4.2.5 Other noteworthy genes

Using the ME-MAGMA approach, we found other genes such as *NPRL2* and *SOX2*, that play crucial roles in the molecular pathways underlying seizure susceptibility and brain development. Loss-of-function mutations in *NPRL2* disrupt the GATOR1 complex, leading to hyperactivation

of the mTORC1 pathway, a significant contributor to seizure development. Additionally, this gene interacts with epigenetic regulators essential for chromatin modification and gene expression, underscoring the intricate relationship between signaling pathways and epigenetic mechanisms in neuronal function.

SRY-related HMG-box 2 (SOX2) is a transcription factor vital for hippocampal development, regulating neural stem cell proliferation. Epigenetic dysregulation of *SOX2* is linked to neurological disorders caused by impaired hippocampal development and synaptic plasticity, potentially contributing to epileptogenesis. 45,46

We also identified additional genes, such as *HYAL2*, *RASSF1*, and *NAB1*, that play significant roles in brain function. ^{47–49} Although *RASSF1* is not directly associated with GGE yet, its epigenetic signatures are linked to other epilepsy-causing genes such as *PTEN*, *BRAF*, *HRAS*, and *KRAS*, which have already been identified. ^{50,51} However, the relationship of *HYAL2* and *NAB1* with epilepsy remains poorly understood, warranting further investigation to elucidate their potential involvement in neural function.

EHMT1 encodes a histone methyltransferase that methylates H3K9. *EHMT1*-associated chromatin modification has been linked to neurodevelopmental disorders and intellectual disability. 41,52

Similarly, *UBTF*, a gene identified through all our analyses except E-MAGMA, has been directly associated with a severe neurodevelopmental disorder known as *UBTF* neuroregression syndrome, also referred to as childhood onset neurodegeneration with brain atrophy. Mutations in *UBTF* disrupt ribosome biogenesis, impairing protein synthesis and cellular function, which contributes to the development of neurodegenerative symptoms, including seizures.⁵³ These findings position *UBTF* as a potential candidate for epilepsy-related studies and highlight the possible role of eQTL in the disorder's pathogenesis.

4.2.6 | Loci 3p21.31 and 17q21.31–32

In this study, we identified multiple candidate genes located at the loci 3p21.31 and 17q21.31–32. Variations in these genes within the same region can lead to polygenic or oligogenic effects, meaning that although each gene may contribute just a small effect, collectively they can surpass a threshold that influences traits like epilepsy. Additionally, the genes that are situated close to one another may share enhancers or promoters, which is often found in coexpressed genes, allowing a single variant to impact the expression of multiple genes.

Deletions in the 3p21.31 locus have been associated with neurodevelopmental disorders, such as intellectual

disability, autism spectrum features, and epilepsy.⁵⁴ Moreover, the CACNA2D2 gene, which encodes a subunit of voltage-dependent calcium channels, has also been implicated in epilepsy through GWAS.⁵

Likewise, deletions in the 17q21.31 locus are linked to a genetic disorder called Koolen-de Vries syndrome. 55 The nearby 17q21.32 locus contains the PNPO gene, where recessive mutations can lead to severe pyridoxal phosphate deficiency, resulting in neonatal or infantile seizures, often showing as status epilepticus during fevers. This region also includes genes such as CBX1 and CDK5RAP3, which are involved in neuronal differentiation and migration, correlating with cortical microdysgenesis observed in juvenile myoclonic epilepsy and other forms of GGE.⁵⁶ Additionally, locus 17q21.32 has been reported as a potential pleiotropic locus for FE and GGE.57

Identification of multiple genes at these loci (Figure 3A) highlights the potential role of meQTLs and eQTL in modulating the relationship between the chromosomal regions 3p21.31 and 17q21.31-32 and GGE. Furthermore, the clustering of genes on chromosome 3, as depicted in the Miami plot from the H-MAGMA analysis (Figure 3B), suggests that these regions may influence neurodevelopmental processes that contribute to susceptibility to GGE.

4.3 Gene-based associations for all epilepsies

In addition to the genes identified for GGE, our analysis on the "all epilepsy" dataset identified genes like HNRNPK, MAGI2, PACS1, and TTC21B using MAGMA, E-MAGMA, and ME-MAGMA, respectively.

HNRNPK gene encodes heterogeneous nuclear ribonucleoprotein K, a multifunctional protein involved in various aspects of RNA metabolism. Mutations or haploinsufficiency in HNRNPK have been linked to a rare autosomal dominant neurodevelopmental disorder known as Au-Kline syndrome and autism spectrum disorder. 58,59

MAGI2 (membrane associated guanylate kinase, WW and PDZ domain containing 2) has been implicated in certain forms of epilepsy, particularly infantile epileptic spasms syndrome, a severe epilepsy syndrome occurring in the first year of life. Research has identified deletions in the MAGI2 gene on chromosome 7q11.23-q21.1 in patients with infantile epileptic spasms syndrome, suggesting a potential role in the disorder's pathogenesis. 60

Mutations in the PACS1 gene are associated with Schuurs-Hoeijmakers syndrome (PACS1 syndrome), a rare neurodevelopmental disorder characterized by intellectual disability, developmental delays, distinctive facial features, and seizures. 61 The identification of PACS1

through E-MAGMA suggests that epilepsy risk may be modulated by eQTLs influencing gene regulation.

Studies show that the cis-regulated expression level of TTC21B is associated with epilepsy, indicating that risk variants may confer epilepsy risk by regulating the expression of this gene. The identification of TTC21B in ME-MAGMA highlights the involvement of DNA methylation mechanisms.

The identification of pseudogenes ACTG1P22 and KRTAP8-2P in our H-MAGMA Adult and Fetal analysis suggests that these genes may provide valuable insights into the hereditary and developmental mechanisms underlying epilepsy.⁶²

4.4 Gene-based associations for focal epilepsies

In our analysis of a focal epilepsy dataset, we identified FANCD2P2, CUL4A, and PROZ as genes and pseudogenes of interest. FANCD2P2 is related to FANCD2, a key component of the Fanconi anemia pathway involved in DNA repair processes.

The CUL4A gene encodes a critical component of the cullin-RING ubiquitin ligase complex, which regulates protein degradation and cell cycle control. The PROZ gene encodes protein Z, a glycoprotein that plays a regulatory role in blood coagulation.

Although these genes are recognized for their essential functions in the human brain, mutation or dysregulation in these loci has not yet been directly linked to neurological disorders, including epilepsy. The identification of these genes in our study may serve as a foundation for future research into their potential roles in the mechanisms underlying neurological diseases. Moreover, incorporating disease-specific methylation data can enhance the discovery of more relevant genes associated with focal epilepsies.²⁵

CONCLUSIONS

In this study, we aimed to uncover new genes associated with epilepsy by integrating genetic variants through multiple layers of gene regulations including positional, transcriptional, and epigenetic. Rather than focusing solely on overlapping results across methods, we emphasized the unique regulatory insights each approach provides, as these can offer critical clues and directions for future research. To achieve this, we utilized a range of MAGMAbased methodologies, including MAGMA, E-MAGMA, H-MAGMA, and a novel approach developed in this study, ME-MAGMA. These methods allowed us to systematically

TABLE 3 List of key candidate genes selected based on their FDR, available literature, and VarElect score.

	Genes	VarElect score ^a	Disease-causing likelihood ^b	ME- MAGMA	E- MAGMA	H-MAGMA adult	H-MAGMA fetal	MAGMA
GGE	CACNB2	5.25	47.24					4.02×10^{-6}
	ZNF629	3.80	65.59	1.17×10^{-2}				
	ZNF668	1.61	66.64			1.64×10^{-2}		3.097×10^{-2}
	ZBTB32	1.39	54.25		2.57×10^{-2}			
	KCND3	18.70	88.13					1.07×10^{-2}
	KCNT1	105.48	77.33			3.82×10^{-2}		
	TMEM107	1.16	72.83			2.54×10^{-3}	2.79×10^{-3}	3.42×10^{-2}
	TMEM186	1.18	46.82			3.02×10^{-2}	3.25×10^{-2}	
	TMEM115	1.43	73.97					1.44×10^{-2}
	NPRL2	36.27	87.30	1.98×10^{-5}				
	SOX2	15.94	70.26	2.25×10^{-3}				
	RASSF1	2.12	26.16	5.21×10^{-5}		2.07×10^{-5}	2.62×10^{-5}	1.45×10^{-2}
	NAB1	3.68	81.39	9.08×10^{-5}		6.35×10^{-4}	1.98×10^{-4}	2.05×10^{-3}
	UBTF	13.95	91.65	1.07×10^{-2}		3.40×10^{-3}	1.42×10^{-3}	5.94×10^{-3}
	EHMT1	17.88	68.20					3.12×10^{-2}
All	HNRNPK	3.33	91.33			3.40×10^{-3}		3.00×10^{-3}
	MAGI2	15.56	80.25					1.30×10^{-2}
	PACS1	16.22	84.75		4.44×10^{-2}			
	TTC21B	18.35	29.84	4.05×10^{-2}				
Focal	PROZ	.11	42.49	5.75×10^{-2}	1.44×10^{-2}			
	CUL4A	.11	53.06					2.72×10^{-2}

Abbreviation: GGE, genetic generalized epilepsy.

analyze the genetic architecture of epilepsy, particularly GGE, by incorporating genomic, transcriptomic, and epigenomic information. Standard MAGMA provided gene-level associations from GWAS data. E-MAGMA and H-MAGMA extended this by integrating gene expression and chromatin interaction information, respectively, whereas ME-MAGMA introduced a new layer of analysis by incorporating meQTL data to capture epigenetic regulation. Together, these complementary approaches revealed candidate genes involved in seizure susceptibility, highlighting the complex interplay between genetic variants and different regulatory mechanisms (Table 3).

Here, we identified both known and novel genes associated with epilepsy. For each gene, we provide information regarding the specific types of genetic variation that may lead to the observed trait. Gaining deeper insights into these variations, and whether they impact on epigenetic mechanisms or developmental processes, can offer more targeted and biologically meaningful interpretations. For example, we observed overlapping genetic variants within several of these genes when mapped using both the H-MAGMA Adult and Fetal brain-specific annotation frameworks. This convergence suggests that these genes may exert regulatory or functional effects across multiple stages of neurodevelopment, potentially influencing susceptibility to epilepsy at different stages of life.

Our findings also highlight the critical role of ion channel dysfunction and transcriptional dysregulation in the development of epilepsy. We identified several key genes, such as *KCNT1*, *CACNB2*, *NPRL2*, and *SOX2*, that comprise common variants associated with epilepsy. In total, we uncovered a set of 879 genes and pseudogenes impacted by diverse genomic mechanisms, broadening the pool of potential therapeutic targets.

This research reinforces the critical role of epigenetic and transcriptional regulation in epilepsy, underscoring the necessity for multiomic approaches to unravel its complex pathogenesis. Although this study offers a set of robust, data-driven hypotheses, it also underscores the

^aVarElect score is an indication of the strength of the connection between the gene and the queried phenotype(s). The main purpose of the score is to enable ranking and prioritizing the list of queried genes by relevance to the queried phenotype(s). ²⁰

^bDisease-causing likelihood reflects the principle that a variant in a gene with high mutation intolerance is more likely to be disease-causing. Detailed description can be found in the referenced publication.²⁰

need for further functional validation and mechanistic exploration. Continued investigation into these genes, particularly in the context of different epilepsy subtypes and neurodevelopmental timelines, will be crucial for deepening our understanding of their roles in the epileptogenesis process.

AUTHOR CONTRIBUTIONS

Ashwini Mushunuri: Writing—original draft preparation; statistical analysis; methodology; visualization. Oluvomi Adesoji: Review of the original draft. Roland Krause: Data curation; review of the original draft. Patrick May: Review of the original draft. Holger Lerche: Review of the original draft. Albert Becker: Resources; review of the original draft. Daniela Grimm: Review of the original draft. International League Against Epilepsy Consortium on Complex **Epilepsies**: Resources; review of the original draft. Michael Nothnagel: Supervision; writing—review & editing. Herbert Schulz: Conceptualization; supervision; funding acquisition; methodology; writing—review & editing.

ACKNOWLEDGMENTS

This study was supported by the Research Unit FOR-2715 of the German Research Foundation (SHU 3585/1-1, NO 755/13-1, LE 1030/16-2, INTER/DFG/17/11583046, and INTER/DFG/21/16394868). The funding bodies had no role in the study design, data collection, analysis, interpretation, or manuscript preparation. Some of the data reported in this paper were collected as part of a project undertaken by the ILAE, and some of the authors are experts selected by the ILAE. However, the opinions expressed by the authors do not necessarily reflect the policy or position of the ILAE. Open Access funding enabled and organized by Projekt DEAL.

FUNDING INFORMATION

This research was funded by the German Research Foundation and the Luxembourg Fond Nationale de Recherche as a part of the research unit FOR-2715 (grants SHU3585/1.1, NO755/6-1, NO755/13-1, Le1030/16-2, INTER/DFG/21/16394868 MechEPI2, BE 2078/5-1, BE 2078/10-2).

CONFLICT OF INTEREST STATEMENT

The authors report no conflict of interest. We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

DATA AVAILABILITY STATEMENT

Research data are not shared.

ORCID

Ashwini Mushunuri https://orcid.

org/0009-0000-7756-5876

Oluyomi Adesoji D https://orcid.

org/0000-0002-8583-7170

Roland Krause https://orcid.org/0000-0001-9938-7126

Patrick May https://orcid.org/0000-0001-8698-3770

Albert Becker https://orcid.org/0000-0003-2661-3705

Daniela Grimm https://orcid.org/0000-0002-4991-3105

Michael Nothnagel https://orcid.

org/0000-0001-8305-7114

Herbert Schulz https://orcid.org/0000-0002-0226-964X

REFERENCES

- 1. Thijs RD, Surges R, O'Brien TJ, Sander JW. Epilepsy in adults. Lancet. 2019;393(16):689-701.
- 2. Scheffer IE, Berkovic S, Capovilla G, Connolly MB, French J, Guilhoto L, et al. ILAE classification of the epilepsies: position paper of the ILAE Commission for Classification and Terminology Epilepsia. 2017;58:512-21.
- 3. Myers CT, Mefford HC. Advancing epilepsy genetics in the genomic era. Genome Med. 2015;7:91.
- International League Against Epilepsy Consortium on Complex Epilepsies. Genome-wide mega-analysis identifies 16 loci and highlights diverse biological mechanisms in the common epilepsies. Nat Commun. 2018;9:5269.
- International League Against Epilepsy Consortium on Complex Epilepsies. GWAS meta-analysis of over 29,000 people with epilepsy identifies 26 risk loci and subtype-specific genetic architecture. Nat Genet. 2023;55:1471-82.
- 6. Song M, Liu J, Yang Y, Lv L, Li W, Luo XJ. Genome-wide meta-analysis identifies two novel risk loci for epilepsy. Front Neurosci. 2021;15:722592.
- 7. de Leeuw CA, Mooij JM, Heskes T, Posthuma D. MAGMA: generalized gene-set analysis of GWAS data. PLoS Comput Biol. 2015;11:e1004219.
- 8. Lu Q, Powles RL, Wang Q, He BJ, Zhao H. Integrative tissuespecific functional annotations in the human genome provide novel insights on many complex traits and improve signal prioritization in genome wide association studies. PLoS Genet. 2016;12:e1005947.
- 9. Kichaev G, Bhatia G, Loh PR, Gazal S, Burch K, Freund MK, et al. Leveraging polygenic functional enrichment to improve GWAS power. Am J Hum Genet. 2019;104:65-75.
- Schulz H, Ruppert AK, Herms S, Wolf C, Mirza-Schreiber N, Stegle O, et al. Genome-wide mapping of genetic determinants influencing DNA methylation and gene expression in human hippocampus. Nat Commun. 2017;8:1511.
- 11. Gerring ZF, Mina-Vargas A, Gamazon ER, Derks EM. E-MAGMA: an eQTL-informed method to identify risk genes using genome-wide association study summary statistics. Bioinformatics. 2021;37:2245-9.
- 12. Sey NYA, Hu B, Mah W, Fauni H, McAfee JC, Rajarajan P, et al. A computational tool (H-MAGMA) for improved prediction of brain-disorder risk genes by incorporating brain chromatin interaction profiles. Nat Neurosci. 2020;23:583-93.

- Price ME, Cotton AM, Lam LL, Farre P, Emberly E, Brown CJ, et al. Additional annotation enhances potential for biologically-relevant analysis of the Illumina Infinium HumanMethylation450 BeadChip array. Epigenetics Chromatin. 2013;6:4.
- 14. Purcell, Shaun CC. Available from: www.cog-genomics.org/plink/2.0/.
- 15. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience. 2015;4:7.
- Genomes Project C, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, et al. A global reference for human genetic variation. Nature. 2015;526:68–74.
- 17. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Ser B Stat Methodol. 1995;57:289–300.
- 18. R Core Team Rao. R: A language and environment for statistical computing. 2013.
- Wickham H, Averick M. Welcome to the tidyverse. J Open Source Softw. 2019;4:1686.
- Stelzer G, Plaschkes I, Oz-Levi D, Alkelai A, Olender T, Zimmerman S, et al. VarElect: the phenotype-based variation prioritizer of the GeneCards suite. BMC Genomics. 2016;17(Suppl 2):444.
- Salmena L. Pseudogenes: four decades of discovery. Methods Mol Biol. 2021;2324:3–18.
- Koko M, Krause R, Sander T, Bobbili DR, Nothnagel M, May P, et al. Distinct gene-set burden patterns underlie common generalized and focal epilepsies. EBioMedicine. 2021;72:103588.
- 23. Prapiadou S, Mayerhofer E, Georgakis MK, Kals M, Radmanesh F, Izzy S, et al. Exploring synaptic pathways in traumatic brain injury: a Cross-phenotype genomics approach. J Neurotrauma. 2025;42:131–42.
- Alsaif HS, Al Ali H, Faqeih E, Ramadan SM, Barth M, Colin E, et al. ZNF668 deficiency causes a recognizable disorder of DNA damage repair. Hum Genet. 2021;140:1395–401.
- Kobow K, Ziemann M, Kaipananickal H, Khurana I, Muhlebner A, Feucht M, et al. Genomic DNA methylation distinguishes subtypes of human focal cortical dysplasia. Epilepsia. 2019;60:1091–103.
- Lauerer RJ, Lerche H. Voltage-gated calcium channels in genetic epilepsies. J Neurochem. 2024;168:3853–71.
- Pippucci T, Parmeggiani A, Palombo F, Maresca A, Angius A, Crisponi L, et al. A novel null homozygous mutation confirms CACNA2D2 as a gene mutated in epileptic encephalopathy. PLoS One. 2013;8:e82154.
- Punetha J, Karaca E, Gezdirici A, Lamont RE, Pehlivan D, Marafi D, et al. Biallelic CACNA2D2 variants in epileptic encephalopathy and cerebellar atrophy. Ann Clin Transl Neurol. 2019;6:1395–406.
- Kjaer C, Palasca O, Barzaghi G, Bak LK, Durhuus RKJ, Jakobsen E, et al. Differential expression of the beta3 subunit of voltage-gated Ca(2+) channel in mesial temporal lobe epilepsy. Mol Neurobiol. 2023;60:5755–69.
- 30. Tinuper P, Bisulli F, Cross JH, Hesdorffer D, Kahane P, Nobili L, et al. Definition and diagnostic criteria of sleep-related hypermotor epilepsy. Neurology. 2016;86(10):1834–42.
- 31. McTague A, Nair U, Malhotra S, Meyer E, Trump N, Gazina EV, et al. Clinical and molecular characterization of KCNT1-related severe early-onset epilepsy. Neurology. 2018;90:e55–e66.

- 32. Wang J, Wen Y, Zhang Q, Yu S, Chen Y, Wu X, et al. Gene mutational analysis in a cohort of Chinese children with unexplained epilepsy: identification of a new KCND3 phenotype and novel genes causing Dravet syndrome. Seizure. 2019;66:26–30.
- 33. Takayama K, Ohno S, Ding WG, Ashihara T, Fukumoto D, Wada Y, et al. A de novo gain-of-function KCND3 mutation in early repolarization syndrome. Heart Rhythm. 2019;16:1698–706.
- 34. Akyuz E, Koklu B, Uner A, Angelopoulou E, Paudel YN. Envisioning the role of inwardly rectifying potassium (Kir) channel in epilepsy. J Neurosci Res. 2022;100:413–43.
- 35. Lambacher NJ, Bruel AL, van Dam TJ, Szymanska K, Slaats GG, Kuhns S, et al. TMEM107 recruits ciliopathy proteins to subdomains of the ciliary transition zone and causes Joubert syndrome. Nat Cell Biol. 2016;18:122–31.
- 36. Su Y, Cao N, Zhang D, Wang M. The effect of ferroptosis-related mitochondrial dysfunction in the development of temporal lobe epilepsy. Ageing Res Rev. 2024;96:102248.
- 37. Lorenzi I, Oeljeklaus S, Aich A, Ronsor C, Callegari S, Dudek J, et al. The mitochondrial TMEM177 associates with COX20 during COX2 biogenesis. Biochim Biophys Acta Mol Cell Res. 2018;1865:323–33.
- 38. Cassandri M, Smirnov A, Novelli F, Pitolli C, Agostini M, Malewicz M, et al. Zinc-finger proteins in health and disease. Cell Death Discov. 2017;3:17071.
- Akter H, Rahman MM, Sarker S, Basiruzzaman M, Islam MM, Rahaman MA, et al. Construction of copy number variation landscape and characterization of associated genes in a Bangladeshi cohort of neurodevelopmental disorders. Front Genet. 2023:14:955631.
- Stelzer G, Rosen N, Plaschkes I, Zimmerman S, Twik M, Fishilevich S, et al. The GeneCards suite: from gene data mining to disease genome sequence analyses. Curr Protoc Bioinformatics. 2016;54:1.30.31–1.30.33.
- 41. Zhang MW, Liang XY, Wang J, Gao LD, Liao HJ, He YH, et al. Epilepsy-associated genes: an update. Seizure. 2024;116:4–13.
- 42. Larivière S. Multiscale Connectomics of Temporal Lobe Epilepsy. Montreal: McGill University (Canada); 2022.
- 43. Hui JB, Silva JCH, Pelaez MC, Sevigny M, Venkatasubramani JP, Plumereau Q, et al. NPRL2 inhibition of mTORC1 controls Sodium Channel expression and brain amino acid homeostasis. eNeuro. 2022;9:ENEURO.0317-21.2022.
- 44. Laribee RN. Transcriptional and epigenetic regulation by the mechanistic target of rapamycin complex 1 pathway. J Mol Biol. 2018;430(7):4874–90.
- 45. Mercurio S, Serra L, Pagin M, Nicolis SK. Deconstructing Sox2 function in brain development and disease cells. 2022;11.
- 46. Sikorska M, Sandhu JK, Deb-Rinker P, Jezierski A, Leblanc J, Charlebois C, et al. Epigenetic modifications of SOX2 enhancers, SRR1 and SRR2, correlate with in vitro neural differentiation. J Neurosci Res. 2008;86:1680–93.
- 47. Peters A, Sherman LS. Diverse roles for Hyaluronan and Hyaluronan receptors in the developing and adult nervous system. Int J Mol Sci. 2020;21:5988.
- 48. Levallet G, Creveuil C, Bekaert L, Peres E, Planchard G, Lecot-Cotigny S, et al. Promoter hypermethylation of genes encoding for RASSF/hippo pathway members reveals specific alteration pattern in diffuse gliomas. J Mol Diagn. 2019;21:695–704.
- Le N, Nagarajan R, Wang JY, Svaren J, LaPash C, Araki T, et al. Nab proteins are essential for peripheral nervous system myelination. Nat Neurosci. 2005;8:932–40.

- 50. Li Q, Xu Z, Qin J, Yang Z. Epilepsy and developmental delay in pediatric patients with PTEN variants and a literature review. Pediatr Neurol. 2025;163:35-44.
- 51. Sran S, Bedrosian TA. RAS pathway: the new frontier of brain mosaicism in epilepsy. Neurobiol Dis. 2023;180:106074.
- 52. Kleefstra T, Kramer JM, Neveling K, Willemsen MH, Koemans TS, Vissers LE, et al. Disruption of an EHMT1-associated chromatin-modification module causes intellectual disability. Am J Hum Genet. 2012;91(13):73-82.
- 53. Braden AA, Xiao J, Hori R, Brown C, Khan MM. An overview of UBTF Neuroregression syndrome. Brain Sci. 2024;14:179.
- 54. Lovrecic L, Bertok S, Zerjav Tansek M. A new case of an extremely rare 3p21.31 interstitial deletion. Mol Syndromol. 2016;7:93-8.
- 55. Myers KA, Mandelstam SA, Ramantani G, Rushing EJ, de Vries BB, Koolen DA, et al. The epileptology of Koolen-de Vries syndrome: electro-clinico-radiologic findings in 31 patients. Epilepsia. 2017;58:1085-94.
- 56. Consortium E, Consortium EM, Steffens M, Leu C, Ruppert AK, Zara F, et al. Genome-wide association analysis of genetic generalized epilepsies implicates susceptibility loci at 1q43, 2p16.1, 2g22.3 and 17g21.32. Hum Mol Genet. 2012;21:5359-72.
- 57. Adesoji OM, Schulz H, May P, Krause R, Lerche H, Nothnagel M. Benchmarking of univariate pleiotropy detection methods applied to epilepsy. Hum Mutat. 2022;43:1314-32.
- 58. Au PYB, McNiven V, Phillips L, Innes AM, Kline AD. Au-Kline Syndrome. In: Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Amemiya A, editors. GeneReviews((R)). Seattle (WA): University of Washington; 1993.
- 59. Gillentine MA, Wang T, Hoekzema K, Rosenfeld J, Liu P, Guo H, et al. Rare deleterious mutations of HNRNP genes

- result in shared neurodevelopmental disorders. Genome Med. 2021;13(63). https://doi.org/10.1186/s13073-021-00870-6
- Marshall CR, Young EJ, Pani AM, Freckmann ML, Lacassie Y, Howald C, et al. Infantile spasms is associated with deletion of the MAGI2 gene on chromosome 7q11.23-q21.11. Am J Hum Genet. 2008;83:106-11.
- 61. Van Nuland A, Reddy T, Quassem F, Vassalli JD, Berg AT. PACS1-neurodevelopmental disorder: clinical features and trial readiness. Orphanet J Rare Dis. 2021;16:386.
- 62. Khurana E, Lam HY, Cheng C, Carriero N, Cayting P, Gerstein MB. Segmental duplications in the human genome reveal details of pseudogene formation. Nucleic Acids Res. 2010;38:6997-7007.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Mushunuri A, Adesoji O, Krause R, May P, Lerche H, Becker A, et al. Genetic risk factor identification for common epilepsies guided by integrative omics data analysis. Epilepsia. 2025;00:1-15. https://doi. org/10.1111/epi.70021