UniFuncNet: a flexible network annotation framework

Pedro Queirós ^{(0,1,*}, Oskar Hickl^{(0,2} Susana Martínez Arbas^{(0,1} Paul Wilmes^{(0,1,3}, Patrick May^(0,2)

1 Systems Ecology, Luxembourg Centre for Systems Biomedicine, University of Luxembourg, 4367, Luxembourg,

2 Bioinformatics Core, Luxembourg Centre for Systems Biomedicine, University of Luxembourg, 4367, Luxembourg

3 Department of Life Sciences and Medicine, Faculty of Science, Technology and Medicine, University of Luxembourg, Esch-sur-Alzette, Luxembourg

* Correspondence: pdqueiros@gmail.com

Abstract

Summary: Functional annotation is an integral part in the analysis of organisms, as well as of multi-species communities. A common way to integrate such information is using biological networks. However, current data integration network tools are heavily dependent on a single source of information, which might strongly limit the amount of relevant data contained within the network. Here we present UniFuncNet, a network annotation framework that dynamically integrates data from multiple biological databases, thereby enabling data collection from various sources based on user preference. This results in a flexible and comprehensive data retrieval framework for network based analyses of omics data. Importantly, UniFuncNet's data integration methodology allows for the output of a non-redundant composite network and associated metadata. In addition, a workflow exporting UniFuncNet's output to the graph database management system Neo4j was implemented, which allows for efficient querying and analysis.

Availability: Source code is available at https://github.com/PedroMTQ/UniFuncNet.

¹ Introduction

There exists an unprecedented amount of biomolecular data available thanks to the advances in, among others, sequencing, mass spectrometry and bioinformatics techniques. This allows for the study of function across several biological levels at high resolution, from single organisms to the combined functional potential of microbial communities. This wealth of information is difficult to access and use in a straightforward and scalable manner, e.g., due to the lack of a universal data repository and the use of a multitude of data formats and annotations.

⁸ Networks are frequently used for large-scale omics data analyses as these are versatile tools that can be ⁹ used to model complex biological systems [1]. The identification and mapping of functional entities (e.g., ¹⁰ proteins) to networks are central tasks performed during large-scale studies of new species or microbial ¹¹ communities. For example, networks have been used to study ecological interactions such as metabolic ¹² cross-feeding, synergism, and antagonism [2], to detect correlations in metabolic networks [3], to identify ¹³ keystone functions and genes [4].

Given the available functional annotations linked to omics data, a common modelling approach, among others [5], is to use genome-scale metabolic models (GSMM) to integrate all, or part, of the metabolic and transport reaction network(s) within an organism or community [6, 7]. Such networks are usually derived by mapping functional annotations to the corresponding reactions and pathways [8, 9], and can be used for the ¹⁸ *in silico* simulation of metabolism.

¹⁹ Several methodologies [10] and tools [11] are now available for the automated generation and semi-curation of

 $_{20}$ GSMMs. Many methods are able to automatically and accurately reconstruct well-known parts of metabolism,

²¹ which, due being shared by many taxa [12], have been more extensively studied [13]. While the apparent

22 conservation in function based on homology is advantageous when modelling well-studied metabolism, the

resulting GSMMs are often very general and redundant, which may not capture the peculiarities of individual

²⁴ organisms. Modelling species-specific metabolic pathways is important, e.g., for understanding microbial ²⁵ interactions [14], but challenging, since annotations are often incomplete [15, 16]. Here, knowledge integration

²⁶ from multiple databases (e.g., MIBig [17], KEGG [18], and MetaCyc [19]) may help.

Even though some resources provide frameworks for mapping functional entities (e.g., KEGG [18] and 27 MetaCyc [19]), combining them into a more comprehensive resource at a case-by-case basis is laborious, 28 since this integration requires extensive cross-linking, and often manual review/curation. One additional 29 complication is the use of different ontologies [20], which leads to the necessity of cross-linking ontology systems 30 with varying structures and resolutions (e.g., KEGG orthologs and gene ontologies [21, 22]). Additionally, 31 while some databases are structured and provide access through the use of application programming interfaces 32 (API) (e.g., KEGG), relational or non-relational or other standardized formats (e.g., json and xml), others 33 provide data in semi-unstructured formats, thereby requiring the implementation of more specialized data 34 processing methodologies (e.g., text mining [23]). 35 In essence, the diversity and quantity of biological databases, constitute some of the major challenges 36 in the integration of such data. These, and other technical challenges, make such resources inaccessible to 37 researchers without a computational background. 38 The challenge of integrating knowledge from multiple sources in an automated manner in the context of 39 network analysis was tackled through the development of the presented network annotation framework -40 (Uni)fied (Func)tional (Net)work (UniFuncNet). UniFuncNet automates the highly time-consuming process of 41

⁴¹ (complete (run) there (run) work (computed interact). Computed automates the nighty time-constituing process of
 ⁴² searching multiple databases, extracting and integrating data into a composite output. Biological databases
 ⁴³ commonly contain multiple entry types (e.g., compounds or genes), therefore, UniFuncNet's implementation
 ⁴⁴ reflects the general structure of such databases; for this purpose, we modelled four different entity types:

genes, reactions, proteins, and compounds. In turn, these data models can then be linked as a network, and
used for storing and exporting information in machine and human-readable formats. Combining data models
with multiple data collection methodologies results in a flexible yet robust data retrieval framework. In turn,
this allows researchers to fine-tune UniFuncNet to their specific routine data integration tasks, starting from

⁴⁹ simple use cases such as collecting ChEBI identifiers (IDs) for a list of compound names and finding reactions ⁵⁰ for certain protein IDs to linking compounds to organisms, or expanding GSMMs. To showcase how the user ⁵¹ can include UniFuncNet in their analysis, the last two previously mentioned use cases have been implemented ⁵² as separate example workflows; while these are simple wrappers around UniFuncNet and other tools, they ⁵³ may serve as a template for future, and potentially more complex, workflows.

⁵⁴ UniFuncNet aims to provide a straightforward, versatile, and accessible data collection and network ⁵⁵ annotation framework. UniFuncNet will prove useful across multiple domains of bioinformatics, especially at ⁵⁶ a moment in time where large-scale data integration is seen as fundamental rather than optional. In order to ⁵⁷ provide an easily and efficiently queryable database, we implemented an API that exports UniFuncNet's data ⁵⁸ to Neo4j.

⁵⁹ Materials and methods

60 Implementation

⁶¹ UniFuncNet was implemented in Python (v3.9) and currently collects data from KEGG [18], MetaCyc [19], ⁶² Rhea [24], ChEBI [25], HMDB [26], UniProt [27] and Pubchem [28], cross-linking the information between ⁶³ these databases. For web data collection, UniFuncNet uses the Python package "requests" (v2.25.1), which ⁶⁴ queries each database and collects the respective response (usually HTML or json). To parse the HTML

responses the "beautiful soup" [29] package is used (v4.10.0). For some of the databases, i.e., MetaCyc [19],

Rhea [24] and ChEBI [25] the database flat files are first downloaded, parsed and stored locally in a SQLite 66

(v3.36.0) database. In order to use the MetaCyc database, the user must obtain a license (academic licenses 67

are freely available) from MetaCyc (which we recommend since it's a highly curated and comprehensive 68 resource). For web data collection, UniFuncNet makes use of API calls to retrieve information (if possible).

69 However, whenever necessary, data is collected by querying the database's website and parsing the query result 70

(i.e., web scraping). Each query result (web or local data) is parsed according to the step of the workflow 71

and database being queried (with database-specific scrapers), and standardized according to UniFuncNet's 72

framework. This data parsing allows for the retrieval of annotations (IDs and synonyms) as well as any 73

connections between database entries. 74

To avoid overloading the respective web servers, UniFuncNet works in a strictly sequential manner and 75

additionally enforces a time-out for requests to the same server (10 seconds in-between queries by default). 76

Additionally, in order to avoid repeating web queries, UniFuncNet saves past web queries in memory and 77

retrieves the necessary entity whenever a query is repeated. This sequential methodology has the additional 78 benefit of not creating redundant entities which may lead to downstream issues with redundancy and output

79

network connectivity. 80

The results shown in this paper were collected from multiple sources, MetaCyc version 25.1 was used; the 81 Rhea and ChEBI data corresponded to the flat files uploaded on the 17th of November, 2021; and all data

82 extracted from the multiple websites was collected on the 24th of January, 2022. The version of UniFuncNet 83

used in this paper is v1.02. 84

Input and output 85

UniFuncNet takes as input a tab-separated file, containing a list of IDs (e.g., "P02769"), ID types (source of 86

the IDs, e.g., "uniprot"), entity types (e.g., "protein"), and search modes (e.g., "pg", for "protein-to-gene"). 87

UniFuncNet outputs one tab-separated file per entity type, i.e., genes, proteins, reactions, and compounds, 88

listing all the searched entities along with any associated metadata (e.g., database IDs) and all the associations 89 between each entity. Additionally, it outputs a file in simple interaction format (SIF), which allows for 90

integration into network frameworks, such as Cytoscape [30] or Neo4j. 91

For a detailed description of input format requirements and all outputs, as well as a usage guide refer to 92 UniFuncNet's documentation at https://github.com/PedroMTQ/UniFuncNet. 93

Workflows methodology 94

We implemented two example workflows to showcase potential applications of UniFuncNet. The first workflow 95 relates to the expansion of GSMMs using UniFuncNet, and the second to the mapping of compounds to 96 organisms. An example use case is provided for each of these workflows. In these use cases, UniFuncNet 97 collected information from the databases KEGG, MetaCyc, Rhea, and ChEBI. The code used for the 98 generation of results is available at https://gitlab.lcsb.uni.lu/pedro.queiros/benchmark_unifuncnet. ٩q After installation and download of the required tools and data, the workflows are fully automated (e.g., 100 automatically launching tools and doing the necessary data processing). Mantis [31] v1.3 was run for the 101 functional annotation, using the KOfam [32], Pfam [33] and MetaCyc [19] reference databases (the MetaCyc 102 database was generated with the code in https://github.com/PedroMTQ/refdb_generator). 103

Workflow I - Expansion of GSMMs 104

This workflow (Figure 1.A) receives as input multiple protein fasta files, i.e., proteomes, and outputs an 105 expanded network per sample in SIF format. The proteomes are passed to Mantis [31] while GSMMs are 106 created with CarveMe [34]. The enzyme commission numbers (ECs) and MetaCyc protein IDs absent in 107 the CarveMe GSMMs are exported from the Mantis' functional annotations into a UniFuncNet-formatted 108 input file (using the "prc" search mode). In this manner UniFuncNet can be used to collect data on the 109 additional ECs and MetaCyc protein IDs and connect them to the original GSMMs. In order to exclude 110 unspecific interactions, edges connecting to common cofactors were removed (this list of cofactors has been 111 manually curated but can be edited and is available at https://github.com/PedroMTQ/UniFuncNet/tree/ 112 main/Resources/cpd_to_ignore.tsv). 113

The workflow was implemented with CarveMe [34] v1.5. Additional information is available at https: 114 //github.com/PedroMTQ/UniFuncNet/tree/main/Workflows/GSMM_Expansion. It is crucial to note that 115

this workflow is merely an example use case, therefore any output files created should be thoroughly curated. 116

As an example application (henceforth referred to as "use case I"), we used the following five organisms' SwissProt[27] reference proteomes: UP000001031 [35] for Akkermansia muciniphila, UP000025221 [36] for Bradyrhizobium japonicum, UP000018291 [37] for Microthrix parvicella, UP000002528 [38] for Pelagibacter whime and UP000000586 [20] for Streptococcus programming

¹²⁰ *ubique*, and UP000000586 [39] for *Streptococcus pneumoniae*.

¹²¹ To evaluate the functional redundancy of the baseline and expanded networks, a presence/absence encoding

¹²² of each network's ECs was applied, followed by a cosine distance calculation using the NLTK package (v3.5), where equal encoded vectors have a score of 1 and completely different a score of 0. This calculation is

where equal encoded vectors have a score of 1 and completely different a score of 0. This calculation is henceforth referred to as the "ECs functional redundancy".

¹²⁵ Workflow II - Omics cross-linking.

The second workflow (Figure 1.B) attempts to link compounds to specific organisms by searching for information on compounds and linking them to functionally annotated organisms. The input are multiple species proteomes and information (i.e., IDs) on the compounds of interest. The output is a network connecting each compound to all proteomes, and hence, to all organisms. Additional information is available at https: //github.com/PedroMTQ/UniFuncNet/tree/main/Workflows/Compounds_to_Organisms_Mapping.

As an example application of this workflow (henceforth referred to as "use case II"), we applied it to the metabolomics dataset MTBLS497 from Metabolights [40]. In the respective experimental study [41] four organisms, *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*, were cultured, sampled and analysed to link them with 13 compounds of interest. The following proteomes from UniProt were used: *E. coli* UP000001410 [42], *K. pneumoniae* UP000000265 [43], *P. aeruginosa* the proteome UP000002438 [44], and *S. aureus* UP000008816 [45].

137 **Results**

138 UniFuncNet

UniFuncNet is a network annotation framework that collects user-defined data from multiple biological 139 databases, e.g., KEGG orthology IDs (Figure 2). The user input determines which information is collected by 140 UniFuncNet. UniFuncNet retrieves data from the respective biological databases and parses it; if applicable, 141 it then branches out and gathers any additional data associated with the originally retrieved data. This is 142 repeated iteratively until all sources of information are exhausted. When retrieving information for compounds, 143 UniFuncNet can retrieve data based on synonyms, and not only IDs, as it may facilitate the integration 144 of data where only synonyms are available. However, the reliability of synonyms-based data retrieval is 145 inferior to IDs due to its ambiguity [46]. UniFuncNet is available as a conda package, and it's respective 146 documentation is available at https://github.com/PedroMTQ/UniFuncNet. 147

148 Data models

In order to standardize the representation of the multiple types of data within the UniFuncNet framework, we 149 implemented multiple data models, each one representing an entity type, i.e., compounds, reactions, proteins, 150 and genes. In general, entities are associated with IDs from multiple databases and other entity-specific 151 data (e.g., compounds may have an associated chemical formula). The respective data models allow for a 152 standardized in-memory integration, storage, and manipulation of data. For example, the reaction data 153 models are especially helpful for integrating the same reaction from multiple databases; since some reaction 154 database entries do not provide cross-linking, it may be necessary to match reaction entities through their 155 stoichiometry and the compound entities they are associated with (i.e., reactants and products). If the 156 stoichiometry and the reactants and products compound entities are the same, the reactions can be considered 157 the same and merged into the same data model, thus avoiding redundancy. Additionally, these entities can 158 be connected to other entities (e.g., a gene can be connected to a protein), and can thus be exported as a 159 network. Entities are connected within the network following the search mode and databases used (Figure 3). 160

161 Search modes

¹⁶² UniFuncNet's data models represent the four main entity types ("g" = gene, "p" = protein, "r" = reaction, "c" ¹⁶³ = compound, see above). These data models are then organized to be retrieved according to the underlying bioRxiv preprint doi: https://doi.org/10.1101/2022.03.15.484380; this version posted March 17, 2022. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC 4.0 International license.



Figure 1. Use cases workflows. A Workflow I: UniFuncNet is used to aid in the expansion of a previously generated GSMM. First, a draft GSMM (grey network) and functional annotations (dashed box with grey and yellow nodes) are extracted from the input proteome (top dashed box with grey nodes). Next, all functional annotations absent (dashed box with yellow nodes) in the model are input into UniFuncNet. Lastly, all of the metabolic model's entities are connected to UniFuncNet's output (yellow and grey nodes connected with non-dashed edges). Optionally, the user may also add all remaining nodes in UniFuncNet's network (yellow nodes connected with dashed edges). B Workflow II: UniFuncNet is used to identify the proteins of an organism involved in the metabolism of specific compounds. First, proteomes for all organisms were collected (represented by the first dashed box with black dots), these were then functionally annotated with Mantis (represented by the second dashed box with black dots, note the lower number of nodes in each proteome, which represents the lack of functional annotations for some proteins). Using UniFuncNet, we created a network with the reactions and respective proteins associated with each input compound. Lastly, using the previously created network, we linked the compounds with their respective proteins in each proteome.



Figure 2. UniFuncNet overview. The input of UniFuncNet is a list of IDs, ID types, entity types, and search modes, which are processed line by line. In this example, UniFuncNet starts by collecting data on the first query (Q1), which is a gene. According to the search mode "gprc" it then searches for data for the connected proteins, reactions and compounds. For the second query (Q2) - a reaction, UniFuncNet first collects data on the reaction and then on the associated compounds (search mode "rc"). UniFuncNet then outputs the results for each collected entity in the respective tsv file, as well as the resulting network in SIF format.

structure of each database; i.e., biological databases entities are generally connected in two directions: 164 $q \rightarrow p \rightarrow r \rightarrow c$ and $c \rightarrow r \rightarrow p \rightarrow q$. 165

UniFuncNet can process entities in 14 possible search modes, i.e., "gp", "gpr", "gpr", "pg", "pr", "pr", "pr", 166 "rpg", "rp", "rc", "cr", "crp", "crpg", "", and "global". Each letter in the search mode corresponds to one of 167 the four different entity types. The "global" search mode corresponds to a search in both directions, e.g., 168 while searching for a given protein, UniFuncNet retrieves information on the associated genes - "pg", as well 169 as the associated reactions and compounds - "prc". The "" search mode corresponds to an "in situ" search 170 on the same entity, i.e., UniFuncNet retrieves information on the given input IDs without connecting them 171 to additional other entities, e.g., when one aims to fetch ChEBI IDs from compound synonyms or for ID 172 conversion. Figure 3 represents a generic example of multiple search modes and how these drive network 173 generation. 174

The user input and search mode are inherently linked to the data that is collected, i.e., the user input ID 175 is used as a seed for data retrieval and to generate an entity, whereas the search mode is used to impose a 176 direction and stop criterion on the data retrieval process. If, for example, the user inputs a reaction ID - the 177 resulting entity will contain the database IDs associated with this reaction. During data retrieval this entity 178 may also be connected to different types of entities, e.g., a reaction entity is usually associated with two or 179 more compound entities. The IDs of these connected entities are then used for posterior data retrieval and 180 generation of the respective entities (Figure 3). The user is able to input multiple search modes (comma 181 separated) for the same input ID, which may be useful, e.g., for connecting a reaction entity to its respective 182 compound and protein entities. 183

UniFuncNet to Neo4j API 184

In order to provide users with the possibility to efficiently query and manage the UniFuncNet results (for 185 example during network analysis), an API importing UniFuncNet's output into Neo4j a highly-flexible graph 186 database management system, was implemented. 187

UniFuncNet's output can be depicted as a multipartite graph, which is a graph whose nodes can be split 188 into multiple independent sets. In the case of UniFuncNet each output file contains multiple entities (e.g., 189 proteins) with entity related annotations (e.g., EC numbers (ECs)) (Figure 4). Since Neo4j is a highly flexible 190 graph-based database it provides a natural integration of UniFuncNet's data models. 191

The API takes as input a folder containing all the UniFuncNet output tsy files and stores the data in a 192 Neo4j database. This database can then be queried using Cypher (Neo4j's querying language) or using any 193 programming language Neo4j API (e.g., the Python or Java drivers). Additionally, we added the option to 194 input Mantis consensus annotations to query the Neo4j database and create the respective SIF networks. 195

Use cases 196

UniFuncNet is a flexible network annotation framework, being usable within diverse contexts. We provide 197 two case scenarios, the first using UniFuncNet for the expansion of GSMMs, and the second for linking 198 compounds with specific organisms. 199

Use case I 200

212

In this use case we used UniFuncNet to expand GSMMs built with CarveMe[34], exploring how many putative 201 connections UniFuncNet could add to the original GSMM. To that end, Mantis is used to provide additional 202 functional annotations, and UniFuncNet to map those functional annotations to the GSMM (Figure 1.A). 203

As an example, we expanded the GSMMs of multiple organisms that have been shown to be relevant 204 in multiple ecosystems: (i) Akkermansia muciniphila has been shown to play an important role in human 205 intestinal health as part of the gut microbiome [47]; (ii) Bradyrhizobium japonicum, has been shown to be a 206 key organism in nitrogen fixation, essential for e.g. soybean plant growth [48]; (iii) Microthrix parvicella, has 207 been shown to be the dominant species involved in the bulking of activated sludge and lipid accumulation in 208 wastewater treatment plants [49]; (iv) Pelagibacter ubique, has been shown to be an ubiquitous ocean-dwelling 209 bacterium that belongs to the SAR11 clade, which is reported to account for 25% of all cells in the ocean 210 [38], and (v) Streptococcus pneumoniae, has been shown to be a key human pathogen, which is one of the 211 leading causes of pneumonia, bacterial meningitis, and sepsis [50].

This workflow compiled a list of all ECs and MetaCyc protein IDs found by Mantis that are not part of 213 the original GSMM (generated by CarveMe). A non-redundant list of IDs over all species was generated. 214

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Figure 3. UniFuncNet search modes: Example of three different search modes available in UniFuncNet and how they sequentially link entities, generating a connected network. The first input line contains a gene ID with search mode "gprc", UniFuncNet searches first for information on this gene and subsequently the directly or indirectly connected entities (one protein, one reaction and three compounds). The second input line contains a protein ID with the search mode "prc". UniFuncNet retrieves first information on the protein, then on two reactions and four compounds; notice how one of the compounds found in the second search is linked to the network created already during the processing of the first input. The third input line contains a compound ID, and the search mode "crp", UniFuncNet then retrieves information on four compounds, three reactions and four proteins. Again, since one of the proteins was already searched during the processing of the second input line, the resulting network will connect these inputs' entities.

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Figure 4. UniFuncNet results as a multipartite graph. The output from UniFuncNet can be represented as a multipartite graph, where the central layers correspond to the entity types (e.g., proteins), and the outer layers to the annotations (e.g., IDs or synonyms). The protein layer contains a protein complex (red dashed circle), comprised of multiple subunits (i.e., protein nodes).

This list was converted to a UniFuncNet input file, which contained 1329 unique EC numbers and 1052 unique MetaCyc protein IDs. UniFuncNet were then run for all these IDs with the "prc" search mode to connect the (p)rotein function annotations to the (r)eactions and (c)ompounds.

UniFuncNet collected 5244 putative reactions, which were then filtered according to multiple steps: (i) filter for proteins associated to at least one reaction (ii) filter for proteins that were also absent in the original GSMM (iii) extract all reactions connected to these proteins, (iv) exclude reactions that were already present in the original GSMM, and (v) match the compounds obtained from UniFuncNet with the GSMM compounds

²²² to match reactions and exclude redundant reactions.

For each proteome, a baseline directed network from the initial GSMM was created, where reactions and their respective substrates and products are represented as nodes (i.e., reactant(s) \rightarrow reaction \rightarrow product(s)).

We then expanded the network by adding UniFuncNet's nodes, either by adding new connections to the baseline network or adding new nodes.

The draft GSMMs and expanded networks were evaluated according to: (i) % of reactions in the largest network component (%RLC); (ii) % of dead end metabolites (%DEM), i.e., metabolites without a transporter reaction that are produced but not consumed or consumed but not produced [51]; (iii) % of newly connected dead end metabolites (%CDEM); and (iv) the amount of new putative reactions that could be added to the GSMM (NR).

On average %RLC decreased from 99.6% (sd=0.4%) to 95.0% (sd=0.7%), &DEMs increased from 4.2% (sd=0.6%) to 12.8% (sd=0.6%), and 0.1% (sd=0.06%) of DEMs were successfully connected in the expanded network. Finally, on average 1005 (sd=485.5) reactions could be added per proteome.

The expanded networks resulted in a substantial enzyme-specific enrichment (i.e., ECs), the most enriched ones being transferases, oxidoreductases and hydrolases. We also analysed the ECs functional redundancy of the each organisms' baseline and expanded networks, i.e., each baseline network was compared to all others baseline networks, and the same was repeated for the expanded networks. On average, we found that the ECs functional redundancy for the "only baseline", "only expanded", "baseline+expanded" networks was 0.74, 0.44, and 0.66 (0-1, 1 being equal), respectively.

When analysing KEGG pathways, the most enriched metabolic capacities corresponded to the metabolism 241 of carbohydrates, lipids, and cofactors and vitamins (from least to most enriched). In the Akkermansia 242 muciniphila expanded network, the biosynthesis and metabolism of glycans was among the metabolic capacities 243 most enriched by the network expansion (161 ECs in the baseline to 166 additional ECs in the expanded 244 network mapped to the kegg pathway "Glycan biosynthesis and metabolism"). In the Microthrix parvicella 245 expanded network, the metabolism of lipids was the metabolic capacity most enriched by the network 246 expansion (31 ECs in the baseline to 218 additional ECs in the expanded network mapped to the kegg 247 pathway "Lipid metabolism"). 248

These results are available in Supplementary table "results.ods" available at https://gitlab.lcsb.uni. lu/pedro.queiros/benchmark_unifuncnet.

251 Use case II

In order to understand how UniFuncNet could be used to link different omics levels, we used it to connect functionally annotated reference proteomes to a metabolomics dataset, i.e., linking metabolism related

²⁵⁴ proteins to their reactions and respective compounds (Figure 1.B).

As an example, we used a metabolomics study [41] that cultured four organisms in artificial sputum and nutrient broth mediums and sampled their headspaces for 13 compounds. These compounds were used as potential biomarkers in order to determine the most appropriate antimicrobial therapy in the treatment of ventilator-associated pneumonia.

In order to find the reactions and proteins associated with each compound, UniFuncNet ran with the search mode "crp". The proteins found to be connected with the compounds via UniFuncNet were then intersected with the functional annotations of each proteome, thus allowing for the identification of the enzymes within each organism involved in the metabolism of these compounds.

After running this workflow we successfully retrieved information on 11 of 13 compounds, eight of these were linked to a total of 30 reactions. These reactions were then connected to a total of 17 proteins. We then linked the proteins connected to reactions (n=17) to the functional annotations of each organism, finding which of these organisms could potentially be involved in the metabolism of studied compounds. We found that all organisms were involved in the metabolism of indole and that *Pseudomonas aeruginosa* was additionally involved with the metabolism of 2-furanmethanol.

269 Discussion and conclusion

Here we present UniFuncNet, a network annotation framework that collects and integrates data from multiple biological databases. UniFuncNet can be used to search for information and generate annotated networks in a flexible manner (i.e., various search modes and input ID types). UniFuncNet automates data collection into a human-readable output, by connecting the different biological entities (i.e., genes, proteins, reactions, and compounds), and it provides a network-structured output, which can be easily used in network-based downstream analysis. An added benefit of UniFuncNet is the standardization of the search methodology, potentially decreasing the accidental omission of information during manual collection/curation.

UniFuncNet collects data from live websites/application programming interfaces (API) and allows the 277 user to update their own local flat files (e.g., MetaCyc or Rhea). UniFuncNet ensures that the collected 278 data is up to date, which represents a limitation in similar projects [46]), since they require regular database 279 maintenance. However, UniFuncNet faces its own challenges: (i) a website's HTML structure or API may 280 change over time, which requires maintenance of UniFuncNet's data collection protocols, (ii) live retrieval of 281 information tends to be slower than using a centralized source of data, and (iii) websites may block scraping 282 attempts if these are done too frequently, which is circumvented by UniFuncNet by having 10 second waiting 283 period between each web query to the same database. While reliable and large data collection is provided by 284 UniFuncNet, as a framework that can speed-up the work of researchers requiring comprehensively annotated 285 networks, it is advisable to perform manual curation during downstream data integration. Overall though, we 286 believe that the benefits of having a lightweight framework with very low storage footprint, always retrieving 287 the latest information, clearly outweigh the aforementioned downsides. 288

Current automated reconstruction tools are capable of generating GSMMs ready for modelling. However, 289 divergent implementations [52, 34, 53] may lead to different outcomes (i.e., the models) due to multiple factors, 290 e.g.: (i) different gene predictions, (ii) different functional annotation reference databases, and (iii) different 291 automated curation implementations [54, 55]. While automated curation offers a good modelling basis, it 292 is unlikely that the current methods will ever be able to encompass the complexity of *in vivo* biological 293 networks. As such, manual curation and expansion of GSMMs remain essential; the latter is routinely done 294 through the iterative analysis of the subsystems for genes, proteins, and reaction(s) of interest. In particular, 295 the end-user searches for information regarding a certain ontology ID, such as KEGG [18] orthology IDs, 296 ECs, or others, in highly comprehensive (and partially redundant) biological databases. To avoid introducing 297 redundancy, cross-linking entities between databases is necessary, which can be done manually or partially 298 automated through ID mapping tools offered by MetaNetX [46] or UniProt [27]. To this end, we implemented 299 a workflow that uses UniFuncNet to facilitate the cross-linking and expansion of GSMMs. 300

We have shown that the networks enriched with UniFuncNet's workflow were better able to capture organism-specific characteristics, e.g., in *Microthrix parvicella* the metabolism of lipids was the most enriched KEGG pathway, which agrees with the findings of Sheik et al. [49]. Similarly, in *Akkermansia muciniphila* the metabolism and biosynthesis of glycans was amongst the most enriched KEGG pathways, which supports the hypothesis that this organism and glycans play an important role in human gut health [47, 56]. Lastly, the metabolism of cofactors and vitamins was, on average, the most enriched KEGG pathway among all organisms.

In general, we found that this workflow could add a substantial amount of reactions to the GSMMs, which. 308 as previously shown [31], is likely due to the more comprehensive reference databases used (Mantis with 309 the KOfam, Pfam, and MetaCyc databases and CarveMe with the BIGG database [57]). In addition, we 310 also found that the similarity between the functional profiles (i.e., ECs functional redundancy) between each 311 organism's network was substantially lower in the expanded networks, highlighting the benefit of applying 312 UniFuncNet to discover functions unique to each organism. It is important to emphasize that the expanded 313 networks would still require curation; indeed the aim of this workflow is not to directly output an expanded 314 GSMM ready for modelling, but to provide the user with a framework that automates some of the most 315 time-consuming curation steps, i.e., expanding and enriching the network. Altogether, these results show 316 the potential of UniFuncNet to support the expansion of GSMMs, provided additional curation steps are 317 implemented by the end-users. While in this manuscript we have shown how UniFuncNet can be used in a 318 targeted manner, it could also be used for the generation of genome-scale metabolic networks. 319

We have also shown how UniFuncNet can be used to link different datasets, in particular how it can be used for linking different omics, which should prove useful for multi-omics network-based analysis. Specifically, in the second workflow, we have shown how UniFuncNet may be used for the mapping of compounds to specific organisms. The results shown in the use case II were not able to connect the organisms and compounds in the same resolution as the respective study [41], which further highlights the need to create and use more comprehensive functional annotation reference databases. However, we found that indole's metabolism was

³²⁵ comprehensive functional annotation reference databases. However, we found that indole's metabolism was ³²⁶ shared among all organisms, which is a clear indication of conservation of function in prokaryotes[58, 59, 60].

shared among all organisms, which is a clear indication of conservation of function in prokaryotes [58, 59, 60]. Despite this, we believe this workflow could be combined with more resolved input proteomes (i.e., using

³²⁷ Despite tins, we believe this worknow could be combined with more resorved input proteomes (i.e., using ³²⁸ proteomics data instead of reference proteomes) and as such could be an even more powerful screening tool

¹for more thorough investigations.

In conclusion, in this article we have highlighted UniFuncNet's ability to automatically and comprehensively

annotate networks. Additionally, we have showcased two use cases which could be used as baseline examples

- 332 for more intricate analysis. We believe that UniFuncNet's flexible search modes and varied input formats
- ³³³ expands its utility into a variety of analysis well beyond the ones shown in this paper.

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340 Conflict of interest statement.

³⁴¹ None declared.

References

- [1] Mikaela Koutrouli et al. "A guide to conquer the biological network era using graph theory". In: Frontiers in bioengineering and biotechnology 8 (2020), p. 34.
- [2] Emilie EL Muller et al. "Using metabolic networks to resolve ecological properties of microbiomes". In: *Current Opinion in Systems Biology* 8 (2018), pp. 73–80.
- [3] Ralf Steuer et al. "Observing and interpreting correlations in metabolomic networks". In: *Bioinformatics* 19.8 (2003), pp. 1019–1026.
- [4] Hugo Roume et al. "Comparative integrated omics: identification of key functionalities in microbial community-wide metabolic networks". In: *npj Biofilms and Microbiomes* 1.1 (2015), pp. 1–11.
- [5] Beatriz García-Jiménez, Jesús Torres-Bacete, and Juan Nogales. "Metabolic modelling approaches for describing and engineering microbial communities". In: *Computational and Structural Biotechnology Journal* 19 (2021), pp. 226–246.
- [6] Willi Gottstein et al. "Constraint-based stoichiometric modelling from single organisms to microbial communities". In: Journal of the Royal Society Interface 13.124 (2016), p. 20160627.
- [7] Adam M Feist et al. "Reconstruction of biochemical networks in microorganisms". In: *Nature Reviews Microbiology* 7.2 (2009), pp. 129–143.
- [8] Ines Thiele and Bernhard Ø Palsson. "A protocol for generating a high-quality genome-scale metabolic reconstruction". In: *Nature protocols* 5.1 (2010), pp. 93–121.
- [9] Zoran Nikoloski et al. "Metabolic networks are NP-hard to reconstruct". In: Journal of theoretical biology 254.4 (2008), pp. 807–816.
- [10] Tunahan Çakır and Mohammad Jafar Khatibipour. "Metabolic network discovery by top-down and bottom-up approaches and paths for reconciliation". In: *Frontiers in Bioengineering and Biotechnology* 2 (2014), p. 62.
- Sebastián N Mendoza et al. "A systematic assessment of current genome-scale metabolic reconstruction tools". In: *Genome biology* 20.1 (2019), pp. 1–20.
- [12] Gayathri Sambamoorthy and Karthik Raman. "Understanding the evolution of functional redundancy in metabolic networks". In: *Bioinformatics* 34.17 (2018), pp. i981–i987.

- [13] Masaaki Kotera and Susumu Goto. "Metabolic pathway reconstruction strategies for central metabolism and natural product biosynthesis". In: *Biophysics and physicobiology* 13 (2016), pp. 195–205.
- [14] Kirstin Scherlach and Christian Hertweck. "Mining and unearthing hidden biosynthetic potential". In: *Nature Communications* 12.1 (2021), pp. 1–12.
- [15] Peter Cimermancic et al. "Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters". In: Cell 158.2 (2014), pp. 412–421.
- [16] Michael E Pyne, Lauren Narcross, and Vincent JJ Martin. "Engineering plant secondary metabolism in microbial systems". In: *Plant physiology* 179.3 (2019), pp. 844–861.
- [17] Satria A Kautsar et al. "MIBiG 2.0: a repository for biosynthetic gene clusters of known function". In: Nucleic acids research 48.D1 (2020), pp. D454–D458.
- [18] Minoru Kanehisa and Susumu Goto. "KEGG: kyoto encyclopedia of genes and genomes". In: Nucleic acids research 28.1 (2000), pp. 27–30.
- [19] Ron Caspi et al. "The MetaCyc database of metabolic pathways and enzymes-a 2019 update". In: Nucleic acids research 48.D1 (2020), pp. D445–D453.
- [20] Robert Stevens et al. "Ontologies in bioinformatics". In: Handbook on ontologies. Springer, 2004, pp. 635–657.
- [21] Michael Ashburner et al. "Gene Ontology: tool for the unification of biology". In: Nature genetics 25.1 (2000), pp. 25–29. ISSN: 1061-4036. DOI: 10.1038/75556.
- [22] Gene Ontology Consortium. "The gene ontology resource: 20 years and still GOing strong". In: Nucleic acids research 47.D1 (2019), pp. D330–D338.
- [23] Damian Szklarczyk et al. "STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets". In: Nucleic Acids Research 47 (D1 2019), pp. D607–D613. ISSN: 1362-4962. DOI: 10.1093/nar/gky1131.
- [24] Parit Bansal et al. "Rhea, the reaction knowledgebase in 2022". In: Nucleic acids research (2021).
- [25] Janna Hastings et al. "ChEBI in 2016: Improved services and an expanding collection of metabolites". In: Nucleic acids research 44.D1 (2016), pp. D1214–D1219.
- [26] David S Wishart et al. "HMDB 4.0: the human metabolome database for 2018". In: Nucleic acids research 46.D1 (2018), pp. D608–D617.
- [27] UniProt Consortium. "UniProt: a worldwide hub of protein knowledge". In: Nucleic acids research 47.D1 (2019), pp. D506–D515.
- [28] Sunghwan Kim et al. "PubChem substance and compound databases". In: Nucleic acids research 44.D1 (2016), pp. D1202–D1213.
- [29] Leonard Richardson. "Beautiful soup documentation". In: April (2007).
- [30] Paul Shannon et al. "Cytoscape: a software environment for integrated models of biomolecular interaction networks". In: Genome research 13.11 (2003), pp. 2498–2504.
- [31] Pedro Queirós et al. "Mantis: flexible and consensus-driven genome annotation". In: *GigaScience* 10.6 (2021), giab042.
- [32] Takuya Aramaki et al. "KofamKOALA: KEGG Ortholog assignment based on profile HMM and adaptive score threshold". In: *Bioinformatics* 36.7 (2020), pp. 2251–2252. ISSN: 1367-4803. DOI: 10. 1093/bioinformatics/btz859. (Visited on 06/25/2020).
- [33] Sara El-Gebali et al. "The Pfam protein families database in 2019". In: Nucleic Acids Res. 47 (2019), pp. 427–432.
- [34] Daniel Machado et al. "Fast automated reconstruction of genome-scale metabolic models for microbial species and communities". In: *Nucleic acids research* 46.15 (2018), pp. 7542–7553.
- [35] Mark WJ Van Passel et al. "The genome of Akkermansia muciniphila, a dedicated intestinal mucin degrader, and its use in exploring intestinal metagenomes". In: *PloS one* 6.3 (2011), e16876.
- [36] Arthur Fernandes Siqueira et al. "Comparative genomics of Bradyrhizobium japonicum CPAC 15 and Bradyrhizobium diazoefficiens CPAC 7: elite model strains for understanding symbiotic performance with soybean". In: *BMC genomics* 15.1 (2014), pp. 1–21.

- [37] Simon Jon McIlroy et al. "Metabolic model for the filamentous 'Candidatus Microthrix parvicella'based on genomic and metagenomic analyses". In: *The ISME journal* 7.6 (2013), pp. 1161–1172.
- [38] Stephen J Giovannoni et al. "Genome streamlining in a cosmopolitan oceanic bacterium". In: science 309.5738 (2005), pp. 1242–1245.
- [39] JoAnn Hoskins et al. "Genome of the bacterium Streptococcus pneumoniae strain R6". In: Journal of bacteriology 183.19 (2001), pp. 5709–5717.
- [40] Kenneth Haug et al. "MetaboLights: a resource evolving in response to the needs of its scientific community". In: *Nucleic acids research* 48.D1 (2020), pp. D440–D444.
- [41] Oluwasola Lawal et al. "Headspace volatile organic compounds from bacteria implicated in ventilatorassociated pneumonia analysed by TD-GC/MS". In: Journal of breath research 12.2 (2018), p. 026002.
- [42] Rodney A Welch et al. "Extensive mosaic structure revealed by the complete genome sequence of uropathogenic Escherichia coli". In: Proceedings of the National Academy of Sciences 99.26 (2002), pp. 17020–17024.
- [43] Michael McClelland et al. "Complete genome sequence of Salmonella enterica serovar Typhimurium LT2". In: Nature 413.6858 (2001), pp. 852–856.
- [44] C K. Stover et al. "Complete genome sequence of Pseudomonas aeruginosa PAO1, an opportunistic pathogen". In: *Nature* 406.6799 (2000), pp. 959–964.
- [45] Allison F Gillaspy et al. "The Staphylococcus aureus NCTC 8325 genome". In: Gram-Positive Pathogens (2006), pp. 381–412.
- [46] Sébastien Moretti et al. "MetaNetX/MNXref: Unified namespace for metabolites and biochemical reactions in the context of metabolic models". In: Nucleic Acids Research 49.D1 (2021), pp. D570–D574.
- [47] Jing Ouyang et al. "The bacterium Akkermansia muciniphila: a sentinel for gut permeability and its relevance to HIV-related inflammation". In: *Frontiers in immunology* 11 (2020), p. 645.
- [48] Hauke Hennecke. "Nitrogen fixation genes involved in the Bradyrhizobium japonicum-soybean symbiosis". In: FEBS letters 268.2 (1990), pp. 422–426.
- [49] Abdul R Sheik et al. "In situ phenotypic heterogeneity among single cells of the filamentous bacterium Candidatus Microthrix parvicella". In: *The ISME journal* 10.5 (2016), pp. 1274–1279.
- [50] Debby Bogaert, Ronald de Groot, and PWM Hermans. "Streptococcus pneumoniae colonisation: the key to pneumococcal disease". In: *The Lancet infectious diseases* 4.3 (2004), pp. 144–154.
- [51] Amanda Mackie et al. "Dead end metabolites-defining the known unknowns of the E. coli metabolic network". In: *PloS one* 8.9 (2013), e75210.
- [52] Johannes Zimmermann, Christoph Kaleta, and Silvio Waschina. "gapseq: Informed prediction of bacterial metabolic pathways and reconstruction of accurate metabolic models". In: *Genome biology* 22.1 (2021), pp. 1–35.
- [53] Oscar Dias et al. "Reconstructing genome-scale metabolic models with merlin". In: Nucleic acids research 43.8 (2015), pp. 3899–3910.
- [54] José P Faria et al. "Methods for automated genome-scale metabolic model reconstruction". In: Biochemical Society Transactions 46.4 (2018), pp. 931–936.
- [55] Almut Heinken et al. "DEMETER: efficient simultaneous curation of genome-scale reconstructions guided by experimental data and refined gene annotations". In: *Bioinformatics* 37.21 (2021), pp. 3974– 3975.
- [56] Nicole M Koropatkin, Elizabeth A Cameron, and Eric C Martens. "How glycan metabolism shapes the human gut microbiota". In: *Nature Reviews Microbiology* 10.5 (2012), pp. 323–335.
- [57] Jan Schellenberger et al. "BiGG: a Biochemical Genetic and Genomic knowledgebase of large scale metabolic reconstructions". In: BMC bioinformatics 11.1 (2010), pp. 1–10.
- [58] Stefanía Magnúsdóttir et al. "Generation of genome-scale metabolic reconstructions for 773 members of the human gut microbiota". In: *Nature biotechnology* 35.1 (2017), pp. 81–89.
- [59] Liang Tian et al. "Deciphering functional redundancy in the human microbiome". In: Nature communications 11.1 (2020), pp. 1–11.

[60] JIA Yu and Joann K Whalen. "A new perspective on functional redundancy and phylogenetic niche conservatism in soil microbial communities". In: *Pedosphere* 30.1 (2020), pp. 18–24.

Input

	ID	ID type	entity type	search mode
Q1	gene ID	DB	gene	gprc
Q2	reaction ID	DB	reaction	rc

Data retrieval



Output

G1 DB:ID1 DB:ID2 G2 DB:ID3 DB:ID4	
proteins.tsv	G1 ap P1
P1 DB:ID5 DB:ID6 P2 DB:ID7 DB:ID8	P1 pr R1
eactions.tsv	R1 rc C1
R1 DB:ID9 DB:ID10 R2 DB:ID11 DB:ID12	G2 gp P2
	P2 nr R2



Annotations

Entities

Annotations



Input

ID	ID type	entity type	search mode	
ID1 ID2 ID3	DB1 DB2 DB3	gene protein compound	grpc prc crp	

Data retrieval

