

ScienceDirect



Reservoirs of antimicrobial resistance in the context of One Health

Milena Despotovic¹, Laura de Nies¹, Susheel Bhanu Busi¹ and Paul Wilmes^{1,2}



The emergence and spread of antimicrobial resistance (AMR) and resistant bacteria, are a global public health challenge. Through horizontal gene transfer, potential pathogens can acquire antimicrobial resistance genes (ARGs) that can subsequently be spread between human, animal, and environmental reservoirs. To understand the dissemination of ARGs and linked microbial taxa, it is necessary to map the resistome within different microbial reservoirs. By integrating knowledge on ARGs in the different reservoirs, the One Health approach is crucial to our understanding of the complex mechanisms and epidemiology of AMR. Here, we highlight the latest insights into the emergence and spread of AMR from the One Health perspective, providing a baseline of understanding for future scientific investigations into this constantly growing global health threat.

Addresses

- ¹ Systems Ecology Group, Luxembourg Centre for Systems Biomedicine, 7 Avenue des Hauts Fourneaux, L-4362 Esch-sur-Alzette, Luxembourg
- ² Department of Life Sciences and Medicine, Faculty of Science, Technology and Medicine, University of Luxembourg, 6, avenue du Swing, Belvaux, L-4367, Luxembourg

Corresponding author: Wilmes, Paul (paul.wilmes@uni.lu)

Current Opinion in Microbiology 2023, 73:102291

This review comes from a themed issue on **Microbiota**Edited by **Christopher Stewart** and **Maria Carmen Collado**

Available online xxxx

https://doi.org/10.1016/j.mib.2023.102291

1369–5274/© 2023 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Introduction

Globalization and enhanced mobility, a growing human population, close contact with animals and their environment, intensive farming, pollution, ecosystem degradation, and climate changes have led to the emergence of new pathogens and the spread of antimicrobial resistance (AMR). Throughout history,

bacterial infections have been a major cause of human and animal diseases. The discovery and use of antibiotics enabled their effective management from a clinical point of view, but at the same time resulted in increased AMR. If no measures are undertaken to mitigate the current rates of emergence and spread of AMR, it is estimated that AMR will result in a financial burden of 100 trillion dollars at the global level and cause over 10 million deaths per year by 2050 [1].

Human health is tightly linked to the health of animals and the ecosystems they share, enabling resistant bacteria or antimicrobial resistance genes (ARGs) to spread between different human, animal, and environmental reservoirs. These phenomena have the potential to rapidly trigger a pandemic, whereby AMR is no longer constrained by either geographic or human-animal borders [2]. In view of the resulting limitations to conventional approaches for the prevention and control of infectious diseases, the One Health approach has emerged. One Health represents a transdisciplinary approach that shifts the focus from disease treatment and control to disease prevention and surveillance. By integrating research on resistant microorganisms circulating in humans, animals, and the environment, One Health is crucial to enhancing our understanding of the complex epidemiology of AMR [2].

In this review, we highlight the latest insights into the emergence and spread of AMR from the One Health perspective, thereby providing a baseline reference of understanding for future scientific investigations into this leading and constantly growing global health threat.

Mechanisms of antimicrobial resistance

Bacteria have evolved various counteractive mechanisms to confer resistance to antimicrobial agents and assure their survival in a competitive environment [3]. Bacterial resistance can be classified as either natural or acquired. Natural resistance is either constitutively expressed in a bacterial species (i.e. intrinsic), or expressed upon exposure to antibiotics (i.e. induced) [4]. Acquired resistance refers to the acquisition of resistance-conferring genetic material through horizontal gene transfer (HGT). Employing HGT via mobile genetic elements (MGEs), bacteria can acquire ARGs through plasmid-mediated conjugation, transduction via bacteriophages,

or integron-mediated transfer of genetic information between bacteria [3,4]. Alternatively, resistance can be acquired via mutations in the chromosomal DNA following antibiotic exposure [5]. Besides encoding for resistance to most, if not all, major classes of antibiotics, multiple genes conferring resistance to different antibiotic categories can be encoded on the same plasmid. This is especially evident in multidrug-resistant *Kleb*siella pneumoniae [6]. Furthermore, plasmids encoding ARGs are not only found within pathogenic bacteria, but can also be detected in commensals [7]. Additionally, the environment within bacterial biofilms, one of the common modes of microbial life, may promote HGT. In particular, it has been recently shown that pathogenic methicillin-resistant Staphylococcus aureus (MRSA) can transfer MGEs, which are too large to be packed into phages, to methicillin-sensitive Staphylococcus aureus strains by natural transformation in biofilms [8].

Microbial reservoirs of antimicrobial resistance

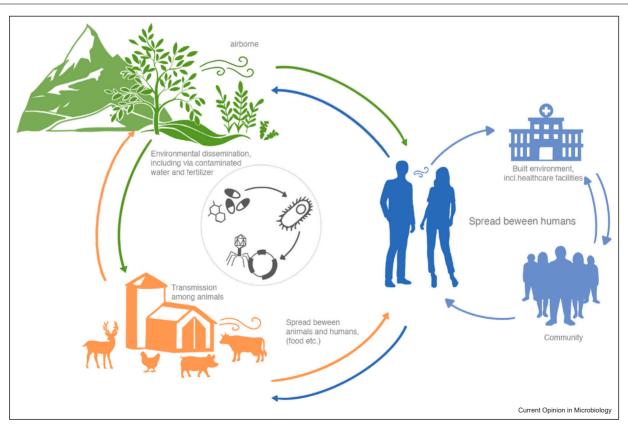
Resistant bacteria residing within human, animal, and environmental reservoirs may spread from one to another, at

both local and global levels (Fig. 1). The role of the resistome (i.e. the collection of ARGs in a given environment or organism) and the differences between ecosystems are of great importance not only to understand the AMR dissemination, but also to identify pools of potentially novel resistance mechanisms.

Many studies have focused specifically on the *Enterococcus* Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, Enterobacter spp., and Escherichia coli pathogens that are highly resistant to last-resort drugs and extensively described in different microbial reservoirs [9]. Additionally, MRSA has been reported to be both human- and animal-associated with a high risk for zoonotic transmission [10,11]. Recent research has been extended to other pathogens posing a threat to human health, such as resistant Campylobacter jejuni for which infections have been reported in humans, animals, and the environment [12]. Similarly, multidrug-resistant Salmonella has been identified in human [13], animal [14], and environmental reservoirs [15].

In the context of One Health, natural microbial communities, or microbiomes, may also have an important

Figure 1



AMR dissemination in One Health. MGE-mediated (i.e. phage, plasmids, and integrons) dissemination of AMR across different microbial reservoirs.

role in the dissemination of AMR. The structure of human and animal microbiomes is shaped by several factors, including exposure to microorganisms through contacts with exogenous sources (e.g. animals, environment), specific host-microbe interactions, and the outcome of competitive, cooperative, and/or predatory (phage) interactions [16]. Recent evidence suggests that ARGs in environmental bacteria can be rapidly acquired by human-associated and pathogenic bacteria [17]. thereby posing a considerable threat to human health.

Human

The prevalence of AMR and the studies thereof have mostly been limited to clinically relevant pathogenic bacteria. These include but are not limited to extendedspectrum beta-lactamase (ESBL)-producing and carbapenem-resistant Klebsiella pneumoniae, ESBL-, AmpC-, and carbapenemase-producing Escherichia coli, carbapenem-resistant Acinetobacter baumannii and Pseudomonas aeruginosa, vancomycin-resistant Enterococcus faecium, MRSA, penicillin-resistant Streptococcus pneumoniae, as well as fluoroquinolone-resistant Salmonella and Shigella species. More recently, lesser-known human pathogens such as Corynebacterium diphtheriae isolates have been reported to carry penicillin, macrolide, and multidrug resistance [18]. However, AMR is also associated with the human microbiome. Although most of the microorganisms constituting the human microbiome are commensals, they have an important role in AMR dissemination. The transfer of AMR can occur from pathogenic bacteria to commensals [19], and from commensals or environmental bacteria to the members of the microbial community [20]. Once ARGs are acquired, commensal organisms may mediate the dissemination of AMR to the microorganisms with pathogenic potential. Interestingly, resistance potential in the gut microbiome exhibits significant differences between geographical areas resulting from differences in antibiotic usage as well as those linked to medicine and food production [21]. In this context, the oral cavity is an important gateway as it constantly reloads the gut microbiome by oral-to-gut transmission [22], and represents an additional microbial reservoir contributing to the resistome [23]. Similarly, due to the constant shedding of microbiota in the environment, the human skin is also an important AMR reservoir [19]. The treatment with systemic antibiotics is associated with long-lasting changes of the human skin microbiome composition potentially contributing to the AMR [24]. Additionally, bacterial transmission within the host may contribute to AMR at smaller spatial scales or in case of a low mutation rate [25]. For example, Wheatley et al. revealed a Pseudomonas aeruginosa-resistant lineage within the gut of a critically ill individual demonstrating local resistance adaptation due to the organ-specific selective pressure. Subsequently, the same strain was found in the lung suggesting within-host transmission as one of the AMR dissemination mechanisms [25].

Transmission of AMR has been extensively discussed in the context of sanitary conditions, such as open defecation or access to clean water [26]. However, recent evidence suggests that transmission of ARGs via air (i.e. bioaerosols) may lead to an increased prevalence of HGT [27,28]. The spread of bioaerosols especially came into focus with the COVID-19 pandemic that has also influenced global AMR spreading. For example, in the WHO European region, reporting on resistant Escherichia coli and Streptococcus pneumoniae was decreased during 2020, likely due to the sanitary measures implemented to prevent the spread of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). However, Acinetobacter spp. and Enterococcus faecium, resistant bacteria usually present in the hospital environments, were more frequently observed during the same period [29].

Animals

Antibiotics are extensively used in livestock and poultry, especially in food production, leading to microbial community compositional shifts and potential increase in ARGs. Treatments using antibiotics in the animal industry also raise the risk of emergence of resistant bacteria due to longer-term selective pressures. In this context, an increased AMR abundance was detected in the gut of farm animals (chicken, turkey, and pig) compared with wild animals (boars, foxes, and rodents) [30]. Similarly, bovine fecal and nasopharyngeal microbiome changes accompanied with increased abundance of ARGs were detected after prophylactic application of antibiotics [31]. A further five-year longitudinal study in pigs and broilers treated with antibiotics for growth promotion highlighted the concomitant increase in AMR specifically in *Enterococcus* spp. [32].

Though the emergence of resistant pathogens is a critical consideration, the spread of ARGs from animal to the human microbiome is of more immediate concern. Such spread can occur via multiple routes, including the direct transmission through food products. Multiple studies have reported food animals as a source of AMR, including multidrug-resistant Salmonella from poultry [33], cephalosporin-resistant Escherichia coli from veal calves [34], and carbapenem-resistant Escherichia coli from pigs [35,36]. Additionally, a number of carbapenem-resistant bacteria, including Pseudomonas, Stenotrophomonas, and Myroides species, were identified in a variety of seafood products [37], underlining the argument that nonpathogenic bacteria, regularly excluded from surveillance programs, may serve as a reservoir for AMR along food supply chains [37,38]. Furthermore, resistant bacteria may spread from animals to humans through direct contact such as in the agricultural sector [16]. For example, livestock-associated MRSA was identified in workers at an industrial livestock operation, but not in workers at an antibiotic-free livestock operation [39]. These reports underline the need for a more comprehensive analysis and monitoring of livestock reservoirs of AMR.

Extensively used concentrated animal feeding operations have also been recognized as AMR reservoirs and a source of resistant bacteria (e.g. *Enterococcus* spp., *Salmonella* spp., *and Vibrio* spp.) in migratory birds [40,41]. They were recently discovered as a source of ESBL-producing *Escherichia coli* in Bangladesh [42]. Given the propensity for these birds to be in contact with humans in populated countries such as Bangladesh, it is likely that ARGs may in turn affect human health or likely disseminate within the human population.

Environment

The role of human-influenced environments in sustaining and disseminating AMR is largely unexplored. Polluted environments (e.g. with heavy metals) contribute to the evolution and spread of AMR through coselection. Heavy metal contamination, for example, coelects antibiotic and metal resistance by cross-resistance, where single genetic mutation may mediate resistance to both metals and antibiotics, or co-resistance, where both metal- and antibiotic-resistance genes are localized on the same MGE [43]. The level of AMR in a specific environment is highly impacted by interactions between different environments. Built environments, for example, hospitals and extended care facilities, where bacteria are exposed to high and repeated doses of antibiotics, represent hotspots for AMR. Furthermore, sewage from both the hospital and the general population is ultimately transported to wastewater treatment plants that therefore provide a vast reservoir for AMR [44]. Importantly, the transmission of ARGs via MGEs was further highlighted recently by de Nies et al. who reported the segregation of ARG categories between plasmids and phages in wastewater treatment plants [45]. Recent evidence suggests that anthropogenic forcing of environments has led to increased AMR in the environment, such as Antarctica, that was previously recognized as pristine [46]. Therefore, the role and potential of the environment as a reservoir cannot be discounted in AMR stewardship in relation to the One Health triad.

Approaches in assessing antimicrobial resistance: a One Health perspective

Traditionally, culture-based methods are used in clinical settings to investigate AMR and resistant bacteria [37] that are readily culturable using standard cultivation

methods. However, sequencing-based methodologies allow for the genomic analysis of all organisms within a microbial ecosystem [47] providing a comprehensive view on all ARGs within different microbial reservoirs. Metagenomic studies that are focused on multiple microbial reservoirs still largely target only one side of the One Health triad, for example, human-animal, animal-environment, or environment-human [48–50]. Nonetheless, some studies have pursued a complete One Health AMR approach [51,52], showing a widespread occurrence of vancomycin-resistance genes in all environments, except from river sediments and drinking water [52]. Additionally, a number of ARGs corresponding to aminoglycoside, macrolide, beta-lactam, and tetracycline resistance were found to be widespread and present in almost all of the investigated environments [51].

To investigate the presence of AMR and MGEs within metagenomes, different bioinformatic workflows, read-, and de novo assembly-based methods [53] have been developed. In the context of One Health, it is crucial to study the prevalence and spread of AMR simultaneously. However, methods to systematically assess AMR within and between biomes have long remained elusive [54]. Tools such as MOCAT2 (metagenomics analysis toolkit) [55] and HUMAnN3 (HMP Unified Metabolic Analysis Network) [56] enable AMR gene identification, but do not provide any information with respect to MGE contextualization. To precisely address the gap in available methodologies, PathoFact [57], which genomically contextualizes ARGs, including their localization on MGEs, was developed. By combining effective study designs with computational analysis methods, PathoFact enables tracing the origins and dissemination of AMR from one reservoir to another using metagenomic sequencing coupled with de novo reconstruction of genomic fragments. Though available studies report the cross-reservoir similarities and likely transmission of AMR in a One Health setting, there is a need for more in-depth characterization of AMR transmission mechanisms, including methods to determine and classify transmission.

Conclusions

AMR is an ever-present concern, not necessarily due to the use of antibiotics alone, but also due to anthropogenic impact and rapid globalization. A major challenge still faced by most One Health studies is attributing the directionality of ARGs between the different metagenomes. To accurately reconstruct the patterns of transmission, especially the directionality of transmission, approaches combining (meta)genomic data analysis, including phylogenetic analysis, with epidemiological approaches and time series, are needed. Since it is evident that AMR reservoirs may affect each other

and given the potential role of human and animal microbiomes in AMR, understanding the interactions/mechanisms and role of each component contributing to the spread of AMR is a critical step in monitoring and addressing this challenge for human health and well-being. Recognizing the microbial reservoirs of AMR is an important first step toward this goal. Furthermore, combined methods incorporating the identity of ARGs, modes of transmission, and integration into the individual reservoirs, alongside crossover mechanisms, may be needed for comprehensive characterization of AMR dissemination. Given the dynamic interactions between humans, animals, and the environment, information on directionality in the One Health context will propel better understanding and management of the different reservoirs, especially in terms of anthropogenic effects. This will allow better antibiotic stewardship contributing to effective treatments using existing antibiotics.

Ethics approval

Not applicable.

Funding

P.W. acknowledges the European Research Council [ERC-CoG 863664]. L.dN. and P.W. were supported by the Luxembourg National Research Fund (FNR) [PRIDE17/11823097]. S.B.B. was supported by the Synergia grant [CRSII5_180241] through the Swiss National Science Foundation. P.W. and M.D. are supported by the Luxembourg Government through the CoVaLux programme.

Consent to participate

Not applicable.

Consent for publication

The authors consent to publication.

CRediT authorship contribution statement

LdN designed and created the overview figure. MD, SBB, LdN, and PW conceptualized the review and contributed to the writing.

Conflict of interest statement

The authors do not have any conflicts of interest relating to this work.

Data Availability

No data were used for the research described in the article.

Acknowledgements

We are grateful for the feedback and input by Dr. Deepthi Budagavi. The authors acknowledge the European Union's Horizon 2020 research and innovation program of the European Research Council (ERC-CoG 863664), Luxembourg National Research Fund (FNR, PRIDE17/11823097), the Synergia grant of the Swiss National Science Foundation (CRSII5_180241), and the Luxembourg Government (CoVaLux programme).

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest
- Brogan DM, Mossialos E: A critical analysis of the review on antimicrobial resistance report and the infectious disease financing facility. Glob Health 2016, 12:8.
- Nadeem SF, Gohar UF, Tahir SF, Mukhtar H, Pornpukdeewattana S, Nukthamna P, Moula Ali AM, Bavisetty SCB, Massa S: Antimicrobial resistance: more than 70 years of war between humans and bacteria. Crit Rev Microbiol 2020, 46:578-599.
- Revgaert WC: An overview of the antimicrobial resistance mechanisms of bacteria. AIMS Microbiol 2018, 4:482-501.
- Martinez JL: General principles of antibiotic resistance in bacteria. Drug Discov Today Technol 2014, 11:33-39.
- MacLean RC, San, Millan A: The evolution of antibiotic resistance. Science 2019, 365:1082-1083.
- Bassetti M, Righi E, Carnelutti A, Graziano E, Russo A: Multidrugresistant Klebsiella pneumoniae: challenges for treatment, prevention and infection control. Expert Rev Anti-Infect Ther 2018. 16:749-761.
- Salinas L, Cárdenas P, Johnson TJ, Vasco K, Graham J, Trueba G: Diverse commensal escherichia coli clones and plasmids disseminate antimicrobial resistance genes in domestic animals and children in a semirural community in Ecuador. mSphere 2019, 4:e00316-e00319.
- Maree M, Thi Nguyen LT, Ohniwa RL, Higashide M, Msadek T. Morikawa K: Natural transformation allows transfer of SCCmecmediated methicillin resistance in Staphylococcus aureus biofilms. Nat Commun 2022, 13:247

The study demonstrates experimental evidence of (Staphylococcal cassette chromosome mec) SCCmec (i.e. a large MGE involved in resistance to β-lactam antibiotics) transfer from MRSA, and from methicillin-resistant coagulase-negative Staphylococci, to methicillinsensitive Staphylococcus aureus, suggesting that natural transformation have an important role in the transfer of MGEs in Staphylococcus aureus

- Ngoi ST, Chong CW, Ponnampalavanar SSLS, Tang SN, Idris N, Abdul Jabar K, Gregory MJ, Husain T, Teh CSJ: Genetic mechanisms and correlated risk factors of antimicrobial resistant ESKAPEE pathogens isolated in a tertiary hospital in Malaysia. Antimicrob Resist Infect Control 2021, 10:70.
- 10. De Boeck I, van den Broek MFL, Allonsius CN, Spacova I, Wittouck S, Martens K, Wuyts S, Cauwenberghs E, Jokicevic K Vandenheuvel D, et al.: Lactobacilli have a niche in the human nose. Cell Rep 2020, 31:107674.
- 11. Lewis HC, Mølbak K, Reese C, Aarestrup FM, Selchau M, Sørum M, Skov RL: Pigs as source of methicillin-resistant Staphylococcus aureus CC398 infections in humans, Denmark. Emerg Infect Dis 2008. 14:1383-1389.
- 12. Dahl LG, Joensen KG, Østerlund MT, Kill K, Nielsen EM: Prediction of antimicrobial resistance in clinical Campylobacter jejuni isolates from whole-genome sequencing data. Eur J Clin Microbiol Infect Dis 2021, 40:673-682.
- 13. Mangat CS, Bekal S, Avery BP, Côté G, Daignault D, Doualla-Bell F, Finley R, Lefebvre B, Bharat A, Parmley EJ, et al.: **Genomic** investigation of the emergence of invasive multidrug-resistant Salmonella enterica Serovar Dublin in humans and animals in Canada. Antimicrob Agents Chemother 2019, 63:e00108-e00119.
- 14. Zhang X-S, Li J, Krautkramer KA, Badri M, Battaglia T, Borbet TC, Koh H, Ng S, Sibley RA, Li Y, et al.: Antibiotic-induced

- - acceleration of type 1 diabetes alters maturation of innate intestinal immunity. eLife 2018, 7:e37816.
- 15. Dionisi AM, Lucarelli C, Benedetti I, Owczarek S, Luzzi I: Molecular characterisation of multidrug-resistant Salmonella enterica serotype Infantis from humans, animals and the environment in Italy. Int J Antimicrob Agents 2011, 38:384-389.
- 16. Trinh P, Zaneveld JR, Safranek S, Rabinowitz PM: One health relationships between human, animal, and environmental microbiomes: a mini-review. Front Public Health 2018, 6:235, https://doi.org/10.3389/fpubh.2018.00235
- 17. Stanton IC, Bethel A, Leonard AFC, Gaze WH, Garside R: What is the research evidence for antibiotic resistance exposure and transmission to humans from the environment? A systematic map protocol. Environ Evid (1) 2020, 9:12, https://doi.org/10.1186/
- Hennart M, Panunzi LG, Rodrigues C, Gaday Q, Baines SL, Barros-Pinkelnig M, Carmi-Leroy A, Dazas M, Wehenkel AM, Didelot X, et al.: Population genomics and antimicrobial resistance in Corynebacterium diphtheriae. Genome Med 2020, 12:107.
- 19. Montassier E, Valdés-Mas R, Batard E, Zmora N, Dori-Bachash M, Suez J, Elinav E: Probiotics impact the antibiotic resistance gene reservoir along the human GI tract in a person-specific and antibiotic-dependent manner. Nat Microbiol 2021, 6:1043-1054.
- 20. Brinkac L, Voorhies A, Gomez A, Nelson KE: The threat of antimicrobial resistance on the human microbiome. Micro Ecol 2017, 74:1001-1008.
- 21. Forslund K, Sunagawa S, Coelho LP, Bork P: Metagenomic insights into the human gut resistome and the forces that shape it: prospects & overviews. BioEssays 2014, 36:316-329.
- 22. Schmidt TS, Hayward MR, Coelho LP, Li SS, Costea PI, Voigt AY, Wirbel J, Maistrenko OM, Alves RJ, Bergsten E, et al.: Extensive transmission of microbes along the gastrointestinal tract. eLife 2019, 8:e42693.
- 23. Carr VR, Witherden EA, Lee S, Shoaie S, Mullany P, Proctor GB,
 Gomez-Cabrero D, Moyes DL: Abundance and diversity of resistomes differ between healthy human oral cavities and gut. Nat Commun 2020, 11:693.

The authors revealed country- and body site-specific differences in the prevalence of ARGs and their classes, as well as the differences in the resistance mechanisms. By demonstrating lower diversity of ARGs of the oral cavity samples in comparison to the paired gut samples, they highlight an ultimate need for extensive surveillance studies to understand the mechanisms and composition of the resistome across different human microbial habitats.

Jo J-H, Harkins CP, Schwardt NH, Portillo JA, NISC Comparative
 Sequencing Program, Zimmerman MD, Carter CL, Hossen MA, Peer CJ, Polley EC, et al.: Alterations of human skin microbiome and expansion of antimicrobial resistance after systemic antibiotics. Sci Transl Med 2021, 13:eabd88077.

Alterations in skin microbiome after administration of systemic antibiotics to healthy human volunteers were investigated in this prospective, randomized study, showing long-lasting changes of the skin microbial communities after antibiotic treatment. The study highlights the role of skin microbiome as a potential source of AMR.

Wheatley RM, Caballero JD, van der Schalk TE, De Winter FHR, Shaw LP, Kapel N, Recanatini C, Timbermont L, Kluytmans J, Esser M, et al.: Gut to lung translocation and antibiotic mediated selection shape the dynamics of Pseudomonas aeruginosa in an ICU patient. Nat Commun 2022, 13:6523.

By combining clinical and genomic data the authors identified gut to lung transmission of Pseudomonas aeruginosa resistant lineage in a critically ill individual at the intensive care unit, suggesting that prevention of gut colonization or gut to lung transmission may represent an important strategy in preventing *Pseudomonas* infection in critically ill individuals. This study paves the way for future studies to confirm gut to lung transmission as a potentially crucial mechanism of Pseudomonas aeruginosa respiratory tract colonization in critically ill individuals.

26. Graham D, Giesen M, Bunce J: Strategic approach for prioritising local and regional sanitation interventions for reducing global antibiotic resistance. Water (1) 2019, 11:27,

- 27. Lee G, Yoo K: A review of the emergence of antibiotic resistance in bioaerosols and its monitoring methods. Rev Environ Sci Biotechnol 2022. 21:799-827.
- 28. Yu Y, Liang Z, Liao W, Ye Z, Li G, An T: Contributions of meat waste decomposition to the abundance and diversity of pathogens and antibiotic-resistance genes in the atmosphere. Sci Total Environ 2021, 784:147128.
- 29. European Centre for Disease Prevention and Control, World Health Organization: Antimicrobial resistance surveillance in Europe: 2022: 2020 data. Publications Office; 2022.
- Skarżyńska M, Leekitcharoenphon P, Hendriksen RS, Aarestrup FM, Wasyl D: A metagenomic glimpse into the gut of wild and domestic animals: quantification of antimicrobial resistance and more. PLoS One 2020, 15:e0242987.
- 31. Holman DB, Yang W, Alexander TW: Antibiotic treatment in feedlot cattle: a longitudinal study of the effect of oxytetracycline and tulathromycin on the fecal and nasopharyngeal microbiota. Microbiome (1) 2019, 7:86, https:// doi.org/10.1186/s40168-019-0696-4
- 32. Aarestrup FM, Seyfarth AM, Emborg H-D, Pedersen K, Hendriksen RS, Bager F: **Effect of abolishment of the use of antimicrobial** agents for growth promotion on occurrence of antimicrobial resistance in Fecal Enterococci from food animals in Denmark. Antimicrob Agents Chemother 2001, 45:2054-2059.
- **33.** Alvarez J, Lopez G, Muellner P, Frutos C, Ahlstrom C, Serrano T, Moreno MA, Duran M, Saez JL, Dominguez L, *et al.*: **Identifying** emerging trends in antimicrobial resistance using Salmonella surveillance data in poultry in Spain. Transbound Emerg Dis 2020, 67:250-262.
- 34. Gay E, Bour M, Cazeau G, Jarrige N, Martineau C, Madec J-Y, Haenni M: Antimicrobial usages and antimicrobial resistance in commensal Escherichia coli from Veal Calves in France: evolution during the fattening process. Front Microbiol 2019,
- 35. Diaconu EL, Carfora V, Alba P, Di Matteo P, Stravino F, Buccella C, Dell'Aira E, Onorati R, Sorbara L, Battisti A, *et al.*: Novel IncFII plasmid harbouring *bla* NDM-4 in a carbapenem-resistant Escherichia coli of pig origin, Italy. J Antimicrob Chemother 2020 75:3475-3479
- 36. Irrgang A, Tausch SH, Pauly N, Grobbel M, Kaesbohrer A, Hammerl JA: First detection of GES-5-producing Escherichia coli from livestock-an increasing diversity of Carbapenemases recognized from german pig production. Microorganisms 2020,
- 37. Morrison BJ, Rubin JE: Carbapenemase producing bacteria in the food supply escaping detection. PLoS One 2015, 10:e0126717.
- 38. Barza M: Potential mechanisms of increased disease in humans from antimicrobial resistance in food animals. Clin Infect Dis 2002, 34:S123-S125.
- 39. Rinsky JL, Nadimpalli M, Wing S, Hall D, Baron D, Price LB, Larsen J, Stegger M, Stewart J, Heaney CD: Livestock-associated methicillin and multidrug resistant Staphylococcus aureus is present among industrial, not antibiotic-free livestock operation workers in North Carolina. PLoS One 2013, 8:e67641.
- 40. Anders J, Bisha B: High-throughput detection and characterization of antimicrobial resistant Enterococcus sp. isolates from GI tracts of European starlings visiting concentrated animal feeding operations. Foods 2020, 9:890.
- 41. Saiful Islam Md, Paul A, Talukder M, Roy K, Abdus Sobur Md, levy S, Mehedi Hasan Nayeem Md, Rahman S, Nazmul Hussain Nazir KHM, Tofazzal Hossain M, et al.: Migratory birds travelling to Bangladesh are potential carriers of multi-drug resistant Enterococcus spp., Salmonella spp., and Vibrio spp. Saudi J Biol Sci 2021, **28**:5963-5970.
- Islam MdS, Sobur MdA, Rahman S, Ballah FM, levy S, Siddique MP, Rahman M, Kafi MdA, Rahman MdT: Detection of blaTEM, blaCTX-M, blaCMY, and blaSHV genes among extendedspectrum beta-lactamase-producing Escherichia coli isolated

- from migratory birds travelling to Bangladesh, Microb Ecol 2022. 83:942-950
- 43. Thomas JC, Oladeinde A, Kieran TJ, Finger JW, Bayona-Vásquez NJ, Cartee JC, Beasley JC, Seaman JC, McArthur JV, Rhodes OE, et al.: Co-occurrence of antibiotic, biocide, and heavy metal resistance genes in bacteria from metal and radionuclide contaminated soils at the Savannah River Site. Micro Biotechnol 2020, **13**:1179-1200.
- 44. Barancheshme F, Munir M: Strategies to combat antibiotic resistance in the wastewater treatment plants. Front Microbiol
- 45. Nies L de, Busi SB, Kunath BJ, May P, Wilmes P: Mobilome-driven segregation of the resistome in biological wastewater treatment, eLife 2022, 11:e81196, https://doi.org/10.7554/eLife.

The study analyzed metagenomic, metatranscriptomic and metaproteomic data systematically collected over 1.5 years from biological wastewater treatment plants. By combining analysis of gene abundance, expression, and association with MGEs and bacterial taxa, the study identified a core group of 15 categories of AMR providing evidence for further monitoring of ARGs in the environmental settings

- Hwengwere K, Paramel Nair H, Hughes KA, Peck LS, Clark MS, Walker CA: Antimicrobial resistance in Antarctica: is it still a pristine environment? Microbiome (1) 2022, 10:71, https://doi.org/ 10.1186/s40168-022-01250-x
- 47. Lepage P, Leclerc MC, Joossens M, Mondot S, Blottière HM, Raes J, Ehrlich D, Doré J: A metagenomic insight into our gut's microbiome. Gut 2013, 62:146-158.
- Duarte ASR, Röder T, Van Gompel L, Petersen TN, Hansen RB, Hansen IM, Bossers A, Aarestrup FM, Wagenaar JA, Hald T: Metagenomics-based approach to source-attribution of antimicrobial resistance determinants - identification of reservoir resistome signatures. Front Microbiol 2021, 11:601407.
- 49. Qian X, Gunturu S, Guo J, Chai B, Cole JR, Gu J, Tiedje JM: Metagenomic analysis reveals the shared and distinct features of the soil resistome across tundra, temperate prairie, and tropical ecosystems. Microbiome 2021, 9:108.

- 50. Bai Y, Ruan X, Xie X, Yan Z: Antibiotic resistome profile based on metagenomics in raw surface drinking water source and the influence of environmental factor: a case study in Huaihe River Basin, China. Environ Pollut 2019, 248:438-447.
- 51. Pal C, Bengtsson-Palme J, Kristiansson E, Larsson DGJ: The structure and diversity of human, animal and environmental resistomes. Microbiome 2016, 4:54.
- 52. Li B, Yang Y, Ma L, Ju F, Guo F, Tiedje JM, Zhang T: Metagenomic and network analysis reveal wide distribution and cooccurrence of environmental antibiotic resistance genes. ISME J 2015. 9:2490-2502.
- 53. Boolchandani M, D'Souza AW, Dantas G: Sequencing-based methods and resources to study antimicrobial resistance. Nat Rev Genet 2019. 20:356-370. https://doi.org/10.1038/s41576-019-
- 54. Kim D-W. Cha C-J: Antibiotic resistome from the One-Health perspective: understanding and controlling antimicrobial resistance transmission. Exp Mol Med 2021, 53:301-309.
- Kultima JR, Coelho LP, Forslund K, Huerta-Cepas J, Li SS, Driessen M, Voigt AY, Zeller G, Sunagawa S, Bork P: MOCAT2: a metagenomic assembly, annotation and profiling framework. Bioinformatics 2016, 32:2520-2523.
- 56. Beghini F, McIver LJ, Blanco-Míguez A, Dubois L, Asnicar F, Maharjan S, Mailyan A, Manghi P, Scholz M, Thomas AM, et al.: Integrating taxonomic, functional, and strain-level profiling of diverse microbial communities with bioBakery 3. eLife 2021,
- 57. de Nies L. Lopes S. Busi SB. Galata V. Heintz-Buschart A. Laczny CC, May P, Wilmes P: PathoFact: a pipeline for the prediction of virulence factors and antimicrobial resistance genes in metagenomic data. Microbiome 2021, 9:49.

The authors developed an easy-to-use modular pipeline for the metagenomic analyses of toxins, virulence factors and AMR. It combines the prediction of these pathogenic factors with the identification of MGEs providing further depth to the analysis by considering the localization of the genes on MGEs and the chromosome.