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Gene-environment interaction and Mendelian randomisation



P. Kolber a,b, R. Krüger a,b,c,*

- ^a Luxembourg Centre for Systems Biomedicine, Clinical and Experimental Neuroscience, University of Luxembourg, 4362 Belval, Esch-sur-Alzette, Luxembourg
- ^b Neurology, Centre Hospitalier de Luxembourg, Luxembourg, Luxembourg

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ABSTRACT

Genetic factors only account for up to a third of the cases of Parkinson's disease (PD), while the remaining cases are of unknown aetiology. Environmental exposures (such as pesticides or heavy metals) and the interaction with genetic susceptibility factors (summarized in the concept of impaired xenobiotic metabolism) are believed to play a major role in the mechanisms of neurodegeneration. Beside of the classical association studies (e.g. genomewide association studies), a novel approach to investigate environmental risk factors are Mendelian randomisation studies. This review explores the gene-environment interaction and the gain of Mendelian randomisation studies in assessing causalities of modifiable risk factors for PD.

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1. Introduction

Parkinson's disease (PD) is the most common movement disorder and the second most frequent neurodegenerative disease after Alzheimer's disease (AD) [1,2]. First described two centuries ago, the underlying cause of PD still remains unknown for the majority of the cases, however, there are recognized factors which increase the risk for PD, including aging, environmental exposure (pesticides or heavy metals) and genetic factors (genetic risk variants and monogenic Mendelian traits) – currently up to 30% of all PD cases can be referred to a genetic contribution [3].

Fundamentally, the motor symptoms of PD – as a prototype for neurodegenerative disorders– are mainly characterized by a progressive loss of dopaminergic neurons linked with an accumulation of aggregation-prone alpha-synuclein protein. These intracellular alpha-synuclein inclusions are a more generalized process affecting different types of neurons in the central and peripheral nervous system contributing to the spectrum of non-motor symptoms in PD and forming the pathognomonic Lewy bodies in affected brain regions (reviewed in [4]). Therefore, specific clearance pathways are involved in PD, that include on one hand impaired protein degradation via the ubiquitin-proteasome system [5] and on another hand impaired lysosomal clearance of proteins and

E-mail address: rejko.krueger@uni.lu (R. Krüger).

^cLuxembourg Institute of Health, Luxembourg, Luxembourg

^{*} Corresponding author at: Luxembourg Centre for Systems Biomedicine (LCBS), Clinical and Experimental Neuroscience, University of Luxembourg, 6 avenue du Swing, 4362 Belval, Esch-sur-Alzette, Luxembourg.

organelles via autophagy [6,7]. The subsequent aggregation of misfolded alpha-synuclein and its prion-like spread within the central and autonomous nervous system is supposed to be a critical step in disease progression [4,8,9]. Whether these aggregates or the preceding (proto-)fibrillary structures of alpha-synuclein interfere with the neuron's vital functions is currently subject of debate [9]. However, the sequestration of other proteins and transcription factors into aggregates, as well as impaired mitochondrial function also increase the cells' vulnerability to excitotoxicity and oxidative stress, leading to energy depletion. In addition, due to deficiency of the ubiquitinproteasome system, the autophagy lysosome pathway plays an important role for degradation of misfolded proteins and defect mitochondria [10]. These mechanisms lead in fine to the activation of the apoptotic cascade and cell death related to neurodegeneration. Some neuronal populations seem to more vulnerable to these pathogenic mechanisms than others within the nervous system, thus determining the clinical phenotype of the disease according to the preferential sites of neuronal dysfunction and subsequent degeneration [11,12].

In the last two decades the investigation of rare familial forms of PD with monogenetic cases showing classical Mendelian inheritance provided the first major insights into the underlying molecular mechanisms involved in PD, by analysing in vitro and in vivo the resulting dysfunction of the proteins encoded by these genes, e.g. alpha-synuclein. This allowed to dissect pathophysiological mechanisms of impaired protein degradation, oxidative stress and mitochondrial dysfunction, and therefore rare monogenic forms of PD served as an entry point to understand the more common sporadic form of PD [13]. Indeed, some of the mutations identified in monogenic forms of PD were also observed in patients without a positive family history or sometimes in unaffected individuals, so apparently not sufficient to cause the disease. This indicates reduced penetrance for some of the mutations. Carriers of the G2019S mutation in the LRRK2 gene are a good example, where patients were diagnosed with sporadic PD mirroring the reduced penetrance of certain genetic risk factors [14-20]. Unbiased genome-wide association studies (GWAS) have demonstrated a role of more common genetic variants in the pathogenesis of idiopathic

PD [20–25]. The "graded risk" concept (Fig. 1) includes Mendelian mutations, low frequency genetic variants and common polymorphisms and introduces the concept of a continuum from more common genetic variants to rare disease-causing mutations with an associated more or less strong impact on the expressivity and contribution to the development of the disease [3,26].

Besides the participation of genetic susceptibility factors in the pathogenesis of neurodegeneration, these findings on reduced penetrance and potential polygenic contribution to disease risk indicate an important role of additional, most probably environmental, risk factors and an important interplay between both genetic and environmental factors in modulating the clinical expression of the disease [27].

2. Gene-environment interactions

2.1. Aging and DNA methylation

Aging, at the cellular level, reflects an accumulation of changes affecting the physiological functioning of the cell and the whole organism, leading to an increased vulnerability to death. "Chronologic age" does not necessarily reflect necessarily "biological age", as additional variables such as individual genetic background, lifestyle and disease processes influence cellular aging. Hallmarks of aging are genetic alterations including telomere attrition, increased genomic instability and epigenetic alterations involving changes in DNA methylation patterns, histone modifications and chromatin remodeling [28].

Different biomarkers previously proposed for defining the biological age of a specific cell or tissue, such as telomere length [29] or age-dependent deletions of mitochondrial DNA [30], were insufficient in their precision (e.g. sensitivity, specificity) and practicality (e.g. accessibility, cost-effectiveness) [31]. Epigenetic changes, especially the DNA methylation status provides a more reliable biomarker in this regard [32–34]. Especially some CpG sites (5'-cytosine-phosphate-guanine-3' sites) exhibit more linear DNA methylation changes throughout aging and are thus valuable biomarkers for age prediction [35–37].

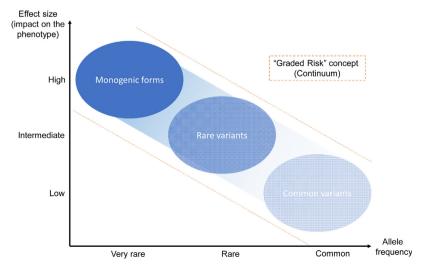


Fig. 1 - The graded-risk concept (adapted from Manolio et al., 2009; [26]).

Interestingly, "epigenetic drift" – the divergence of the epigenome due to random changes in methylation (or errors in the maintenance of the epigenome) over a lifespan – is influenced substantially by environmental factors, as shown in twin studies [36,37]. This is believed to cause the individual susceptibilities, prognoses, treatment outcomes and in a larger sense the individual clinical phenotypes of common diseases.

Over time, the accumulation of epigenetic changes – or epimutations – is believed to induce instability of the genome, to facilitate genetic mutations and finally pathological processes defining a disease. These changes in DNA methylation and their effect on gene expression give us new insight in agerelated diseases such as PD [38,39].

2.2. Gene-environment interaction

Within the last two decades, genetic causes for PD have more and more come to the forefront in PD research [40]. However, most cases cannot be explained yet by a monogenetic form of PD, nor by risk variants. Inversely, even though environmental factors are recognized to contribute to PD, not everyone with PD has been exposed to one of the currently known environmental risk factors and vice versa not everyone exposed to an environmental risk factor is developing the disease. Commonly, pesticides are thought to alter mitochondrial function and increase oxidative stress in dopaminergic neurons, but also to accelerate the formation of alpha-synuclein fibrils and interfere with the function of the ubiquitin-proteasome system [41-43]. Professional or occupational pesticide usage may increase expression of the dopamine transporter (DAT) and lead to an accumulation of toxicants in dopaminergic neurons, affecting dopaminergic neurotransmission in subjects carrying DAT susceptibility alleles [44,45]. Exposure to organophosphates of subjects carrying polymorphisms in the ACHE/PON1 locus might increase the risk for insecticide-induced PD via a neurotransmission imbalance due to impaired acetylcholinesterase (AChE) and paraoxonase (PON1) activity (and in fine impaired organophosphate degradations) [46,47]. The combination and interaction of genetic susceptibility factors and environmental exposure is summarized in the concept of impaired xenobiotic metabolism and sheds first light into the mechanisms of neurodegeneration. These initial insights may translate into new therapies for PD patients that take into account the individual genetic backgrounds reflected in enzymatic activities [48-50].

Even once associations of PD with environmental factors were established, it was yet not clear whether there is (inverse) causality between PD and these factors, e.g. the reverse association between PD and smoking [51]. One way to investigate environmental factors and their causality for PD might be by using so called Mendelian randomisation (MR) methods.

3. Mendelian randomisation

3.1. Principles of Mendelian randomisation

Association studies are a common tool to investigate aetiologies of pathologies, however it is now known that

association and causality are not equivalent by no means, as an association between a disease and a risk factor can be caused by unknown factors (confounders) or the disease can cause the increased presence of a risk factor (reverse causation), as represented in Fig. 2A. In order to avoid confounding factors (selection bias, cofounders, reverse causation...), randomised control trials (RCT), by randomly assigning study participants in two or more groups, are considered as gold standard for investigating causality in prospective clinical trials. Besides the ethical concerns of exposing subjects deliberately to the investigated risk factor, these studies have the downside that they are expensive and time consuming as a longitudinal observation period is needed. Additionally, RCTs are limited in the number of risk factors investigated and they only reflect the effect of an exposure during a certain period of time in the life of the study participant.

The aim of a MR study is to assess whether a non-genetic/ modifiable environmental exposure is associated with the investigated pathology by introducing a randomisation method into an observational study and avoiding the above-mentioned downsides of RCTs [52]. As shown in a directed acyclic graph (Fig. 2A), MR is able to test the null hypothesis that the pathology (outcome) is not caused by an exposure, by assuming that genetic variants used as instrumental variables (IV) are robustly associated with the investigated exposure, are independent of confounders and are associated with the investigated outcome solely (and linearly, unaffected by statistical interactions) via the exposure [53]. The randomisation method in MR studies is based on Mendelian principles of inheritance [54]. MR studies use genetic variants, such as single nucleotide polymorphisms (SNPs) as IVs to measure the effect of an non-genetic exposure on an outcome (Fig. 2A) [55,56]. This randomisation method of using germline genetic variants (which are supposed to be randomly distributed during conception in the general population) as variables, is used for proving causal associations and to accurately estimate the effect of a lifelong exposure to an environmental/ lifestyle risk factor on an outcome [57], by being less susceptible to the biases of observational studies [58].

3.2. Mendelian randomisation in Parkinson's disease

In the past decades, classical epidemiological studies (including potential recall biases) could determine some environmental risk factors for PD, for example pesticide usage or head trauma [59–62]. But also, protective factors having a negative association with the PD risk, such as smoking, caffeine drinking or elevated urate levels in serum were identified in such studies. These negative associations (or potential protective factors) are widely discussed and MR studies provide a way to analyse these associations for their causalities and to shed light on potential protective factors.

Potentially due to its antioxidative effect, elevated plasma urate levels were shown to have a negative association with PD risk [63]. A Japanese clinical study had already investigated whether inosine (a urate precursor) would increase the plasma urate level of PD patients, without proving causality [64]. Two independent Danish studies recently used MR methods to assess causality of PD incidence and metabolic data such as

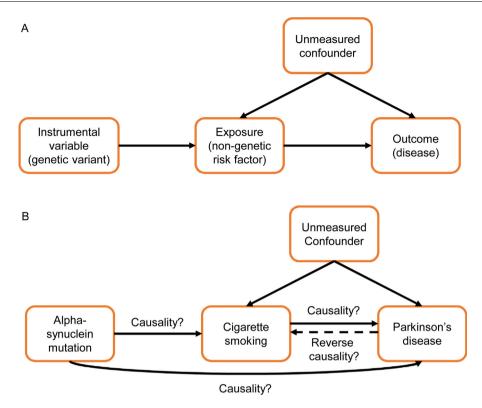


Fig. 2 – The principles of Mendelian randomisation represented in a directed acyclic graph (A). The example of Parkinson's disease is represented in B. Genetic variants of alpha-synuclein (or other) can potentially cause the PD phenotype or even a "primary personality" for a tendency to expose to non-genetic factors (such as cigarette smoking). If cigarette smoking is "protective factor" for PD (causality) or due to this "primary personality" (reverse causality) is debated.

plasma urate levels [65,66]. The epidemiological association between elevated plasma urate and a reduced risk for PD was shown, but MR analysis remained negative and provided no evidence for a causal relationship between this modifiable risk factor and the outcome. The initially demonstrated epidemiological association might therefore be attributed to other unknown factors than the elevated urate levels, making the rationale of clinical inosine studies potentially less relevant [67].

The negative association of PD risk and coffee drinking or nicotine smoking was previously shown in different epidemiological studies [68], raising the controversy over whether, for example, smoking cigarettes reduces the risk for PD, or PD patients are less likely to become addicted to smoking because of their underlying "primary personality" (Fig. 2B). Serum caffeine and its metabolites were even investigated as potential diagnostic biomarkers for early PD, consistent with its supposed neuroprotective effect [69]. However, the underlying causality is not known yet (neuroprotective substances absorbed by the lungs or digestive tract are speculated) and an RCT exposing intentionally subjects to cigarette smoking to investigate the risk for PD would be highly unethical. Many MR studies have been conducted as a new approach to investigate potential protective environmental factors, such as coffee consumption, and their effect on health [70]. Also high serum iron levels and a high body mass index (BMI) were shown in MR studies to have a negative causal effect on PD risk [71,72]. Low LDL cholesterol levels on the other hand, might have a

causal effect on lowering the risk for AD, without an effect on PD risk [73].

A recent MR study on daily coffee consumption and PD risk could not define a causal effect between these two, but did find a positive causal association between being a "morning person" and increased PD risk [74]. The circadian rhythm is highly determined by genetic variants and the presumption is that the resulting phenotype "morning person" is more active in the morning and does not need to drink as much coffee to start its day. However, the "night owl" (having a lower risk for PD) would need more coffee in the morning to cope with the "8 a.m. to 5 p.m." work schedule dictated by the society, explaining the reduced risk for PD associated with coffee consumption as an epiphenomenon. This causal association of the sleep-wake rhythm and PD (or neurodegenerative diseases in general) makes sense in light of studies showing amyloid-beta accumulation in sleep deprived brains [75].

3.3. Limitations of Mendelian randomisation

Complementing the classical observational studies by avoiding their biases, MR is not without its limitations. In order to be used to assess the effect of a non-genetic risk factor on an outcome, MR relies on the availability of robust genetic data. Many GWAS studies have enlightened in the last decades not only associations with diseases, but also complex measurable parameters such as body mass index (BMI; [76]), serum level of metabolites (such as serum uric acid or cholesterol levels;

[77,78]), cognitive features or many other behaviour patterns like diets and habits [79–81]. For example, Yengo and colleagues analysed GWAS data of about seven hundred subjects using MR and identified hundreds of exposure-associated SNPs (data available because of the increasing number of genetic association studies and data banks) [76]. The discovery of new genetic traits and SNPs and the assessment of causalities are more accurate if a large sample of genetic data is available. Therefore, MR studies cannot be used if no SNPs have been identified to be associated for the exposure, or they don't have sufficient power to detect causality if the SNPs don't explain the integral exposure variation [57].

4. Conclusion

Mendelian randomisation studies are an efficient method, complementary to classical association studies, to assess causalities of modifiable environmental risk factors if sufficient genetic data is available. Therefore, cohorts assessing a huge array of different clinical data and providing a detailed genetic characterization are needed in the future to enhance the power of these MR studies [82]. Avoiding confounder and reverse causality biases of association studies, MR thus enables to investigate gene-environmental interactions and determine potential protective factors to be implemented in future preventive medicine campaigns. They might offer a good tool for guiding future clinical RCTS, by avoiding errancy and waste of research resources.

5. Disclosure of interest

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REFERENCES

- [1] Ascherio A, Schwarzschild MA. The epidemiology of Parkinson's disease: risk factors and prevention. Lancet Neurol 2016;15:1257–72. http://dx.doi.org/10.1016/S1474-4422(16)30230-7.
- [2] Poewe W, Seppi K, Tanner CM, Halliday GM, Brundin P, Volkmann J, et al. Parkinson disease. Nat Rev Dis Prim 2017;3:17013.
- [3] Larsen SB, Hanss Z, Krüger R. The genetic architecture of mitochondrial dysfunction in Parkinson's disease. Cell Tissue Res 2018;373:21–37. <u>http://dx.doi.org/10.1007/s00441-017-2768-8</u>.
- [4] Goedert M, Clavaguera F, Tolnay M. The propagation of prion-like protein inclusions in neurodegenerative diseases. Trends Neurosci 2010;33:317–25. http://dx.doi.org/10.1016/j.tins.2010.04.003.
- [5] Atkin G, Paulson H. Ubiquitin pathways in neurodegenerative disease. Front Mol Neurosci 2014;7:63. http://dx.doi.org/10.3389/fnmol.2014.00063.

- [6] Cuervo AM, Wong ESP, Martinez-Vicente M. Protein degradation, aggregation, and misfolding. Mov Disord 2010;25:S49–54. http://dx.doi.org/10.1002/mds.22718.
- [7] Pickrell AM, Youle RJ. The Roles of PINK1. Parkin, and Mitochondrial Fidelity in Parkinson's Disease. Neuron 2015;85:257–73. <u>http://dx.doi.org/10.1016/j.neuron.2014.12.007</u>.
- [8] Recasens A, Dehay B. Alpha-synuclein spreading in Parkinson's disease. Front Neuroanat 2014;8:159. http://dx.doi.org/10.3389/fnana.2014.00159.
- [9] Goedert M. Alzheimer's and Parkinson's diseases: The prion concept in relation to assembled Aβ, tau, and αsynuclein. Science 2015;80:349. http://dx.doi.org/10.1126/ science.1255555.
- [10] Pan T, Kondo S, Le W, Jankovic J. The role of autophagy-lysosome pathway in neurodegeneration associated with Parkinson's disease. Brain 2008;131:1969–78. http://dx.doi.org/10.1093/brain/awm318.
- [11] Braak H, Del Tredici K, Rüb U, De Vos RAI, Jansen Steur ENH, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. Neurobiol Aging 2003;24:197–211. http://dx.doi.org/10.1016/S0197-4580(02)00065-9.
- [12] Braak H, Rüb U, Gai WP, Del Tredici K. Idiopathic Parkinson's disease: possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. J Neural Transm 2003;110:517–36. http://dx.doi.org/10.1007/s00702-002-0808-2.
- [13] Schiesling C, Kieper N, Seidel K, Krüger R. Review: Familial Parkinson's disease–genetics, clinical phenotype and neuropathology in relation to the common sporadic form of the disease. Neuropathol Appl Neurobiol 2008;34:255–71. http://dx.doi.org/10.1111/j.1365-2990.2008.00952.x.
- [14] Maraganore DM, De Andrade M, Elbaz A, Farrer MJ, Ioannidis JP, Krüger R, et al. Collaborative analysis of α-synuclein gene promoter variability and Parkinson disease. J Am Med Assoc 2006;296:661–70. http://dx.doi.org/10.1001/jama.296.6.661.
- [15] Abou-Sleiman PM, Muqit MMK, McDonald NQ, Yang YX, Gandhi S, Healy DG, et al. A heterozygous effect for PINK1 mutations in Parkinson's disease? Ann Neurol 2006;60:414–9. http://dx.doi.org/10.1002/ana.20960.
- [16] Goldwurm S, Zini M, Mariani L, Tesei S, Miceli R, Sironi F, et al. Evaluation of LRRK2 G2019S penetrance. Neurology 2007;68:1141–3. http://dx.doi.org/10.1212/01.wnl.0000254483.19854.ef.
- [17] Di Fonzo A, Rohé CF, Ferreira J, Chien HF, Vacca L, Stocchi F, et al. A frequent LRRK2 gene mutation associated with autosomal dominant Parkinson's disease. Lancet 2005;365:412–5. http://dx.doi.org/10.1016/S0140-6736(05)17829-5.
- [18] Latourelle JC, Sun M, Lew MF, Suchowersky O, Klein C, Golbe LI, et al. The Gly2019Ser mutation in LRRK2is not fully penetrant in familial Parkinson's disease: the GenePD study. BMC Med 2008;6:32. http://dx.doi.org/10.1186/1741-7015-6-32.
- [19] Mueller JC, Fuchs J, Hofer A, Zimprich A, Lichtner P, Illig T, et al. Multiple regions of α-synuclein are associated with Parkinson's disease. Ann Neurol 2005;57:535–5341. http://dx.doi.org/10.1002/ana.20438.
- [20] Foroud T, Uniacke SK, Liu L, Pankratz N, Rudolph A, Halter C, et al. Heterozygosity for a mutation in the parkin gene leads to later onset Parkinson disease. Neurology 2003;60:796–801. http://dx.doi.org/10.1212/ 01.WNL.0000049470.00180.07.
- [21] Simón-Sánchez J, Schulte C, Bras JM, Sharma M, Gibbs JR, Berg D, et al. Genome-wide association study reveals genetic risk underlying Parkinson's disease. Nat Genet 2009;41:1308.

- [22] International Parkinson Disease Genomics Consortium, Nalls MA, Plagnol V, Hernandez DG, Sharma M, Sheerin UM, et al. Imputation of sequence variants for identification of genetic risks for Parkinson's disease: a meta-analysis of genome-wide association, studies. Lancet 2011;377:641–9. http://dx.doi.org/10.1016/S0140-6736(10)62345-8.
- [23] Beecham AH, Patsopoulos NA, Xifara DK, Davis MF, Kemppinen A, et al. Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. Nat Genet 2013;45:1353.
- [24] Lambert J-C, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet 2013;45:1452.
- [25] Saad M, Lesage S, Saint-Pierre A, Corvol JC, Zelenika D, Lambert JC, et al. Genome-wide association study confirms BST1 and suggests a locus on 12q24 as the risk loci for Parkinson's disease in the European population. Hum Mol Genet 2011;20:615–27. http://dx.doi.org/10.1093/hmg/ddo497.
- [26] Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ, et al. Finding the missing heritability of complex diseases. Nature 2009;461:747.
- [27] Robak LA, Jansen IE, van Rooij J, Uitterlinden AG, Kraaij R, Jankovic J, et al. Excessive burden of lysosomal storage disorder gene variants in Parkinson's disease. Brain 2017;140:3191–203. http://dx.doi.org/10.1093/brain/awx285.
- [28] López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell 2013;153:1194–217. http://dx.doi.org/10.1016/j.cell.2013.05.039.
- [29] Blasco MA. Telomeres and human disease: ageing, cancer and beyond. Nat Rev Genet 2005;6:611–22. http://dx.doi.org/10.1038/nrg1656.
- [30] Cortopassi GA, Shibata D, Soong NW, Arnheim N. A pattern of accumulation of a somatic deletion of mitochondrial DNA in aging human tissues. Proc Natl Acad Sci 1992;89:7370–4. http://dx.doi.org/10.1073/pnas.89.16.7370.
- [31] Meissner C, Ritz-Timme S. Molecular pathology and age estimation. Forensic Sci Int 2010;203:34–43. http://dx.doi.org/10.1016/j.forsciint.2010.07.010.
- [32] Fraga MF, Esteller M. Epigenetics and aging: the targets and the marks. Trends Genet 2007;23:413–8. http://dx.doi.org/10.1016/j.tig.2007.05.008.
- [33] Weidner CI, Lin Q, Koch CM, Eisele L, Beier F, Ziegler P, et al. Aging of blood can be tracked by DNA methylation changes at just three CpG sites. Genome Biol 2014;15:R24. http://dx.doi.org/10.1186/gb-2014-15-2-r24.
- [34] Horvath S. DNA methylation age of human tissues and cell types. Genome Biol 2013;14:3156. http://dx.doi.org/10.1186/gb-2013-14-10-r115.
- [35] Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sadda S, et al. Genome-wide methylation profiles reveal quantitative views of human aging rates. Mol Cell 2013;49:359–67. http://dx.doi.org/10.1016/j.molcel.2012.10.016.
- [36] Bocklandt S, Lin W, Sehl ME, Sánchez FJ, Sinsheimer JS, Horvath S, et al. Epigenetic Predictor of Age. PLoS One 2011;6:1–6. http://dx.doi.org/10.1371/journal.pone.0014821.
- [37] Koch CM, Wagner W. Epigenetic-aging-signature to determine age in different tissues. Aging (Albany NY) 2011;3:1018–27. http://dx.doi.org/10.18632/aging.100395.
- [38] Tan Q, Heijmans BT, Hjelmborg JVB, Soerensen M, Christensen K, Christiansen L. Epigenetic drift in the aging genome: a ten-year follow-up in an elderly twin cohort. Int J Epidemiol 2016;45:1146–58. http://dx.doi.org/10.1093/ije/dyw132.
- [39] Renani PG, Taheri F, Rostami D, Farahani N, Abdolkarimi H, Abdollahi E, et al. Involvement of aberrant regulation of

- epigenetic mechanisms in the pathogenesis of Parkinson's disease and epigenetic-based therapies. J Cell Physiol n.d.;0. https://doi.org/10.1002/jcp.28622.
- [40] Singleton AB, Hardy JA, Gasser T. The birth of the modern era of Parkinson's Disease Genetics. J Parkinsons Dis 2017;7:S87–93. <u>http://dx.doi.org/10.3233/jpd-179009</u>.
- [41] Abdollahi M, Ranjbar A, Shadnia S, Nikfar S, Rezaiee A. Pesticides and oxidative stress: a review. Med Sci Monit 2004;10:RA141–7.
- [42] Betarbet R, Canet-Aviles RM, Sherer TB, Mastroberardino PG, McLendon C. Kim J-H, et al. Intersecting pathways to neurodegeneration in Parkinson's disease: Effects of the pesticide rotenone on DJ-1, (-synuclein, and the ubiquitin – proteasome system). Neurobiol Dis 2006;22:404–20. http://dx.doi.org/10.1016/j.nbd.2005.12.003.
- [43] Uversky VN, Li J, Fink AL. Pesticides directly accelerate the rate of alpha-synuclein fibril formation: a possible factor in Parkinson's disease. FEBS Lett 2001;500:105–8.
- [44] Kelada SNP, Checkoway H, Kardia SLR, Carlson CS, Costa-Mallen P, Eaton DL, et al. 5' and 3' region variability in the dopamine transporter gene (SLC6A3), pesticide exposure and Parkinson's disease risk: A Hypothesis-Generating Study. Hum Mol Genet 2006;15:3055–62. http://dx.doi.org/10.1093/hmg/ddl247.
- [45] Ritz BR, Manthripragada AD, Costello C, Lincoln SJ, Farrer LJ, Cockburn M, et al. Dopamine Transporter Genetic Variants and Pesticides in Parkinson's Disease. Environ Health Perspect 2009;117:964–9. http://dx.doi.org/10.1289/ehp.0800277.
- [46] Benmoyal-Segal L, Vander T, Shifman S, Bryk B, Ebstein RP, Marcus E-L, et al. Acetylcholinesterase/paraoxonase interactions increase the risk of insecticide-induced Parkinson's disease. FASEB J 2005;19:452–4. http://dx.doi.org/10.1096/fj.04-2106fje.
- [47] Gatto NM, Cockburn M, Bronstein J, Manthripragada AD, Ritz B. Well-Water Consumption and Parkinson's Disease in Rural California. Environ Health Perspect 2009;117:1912–8. http://dx.doi.org/10.1289/ehp.0900852.
- [48] Le Couteur DG, McLean AJ, Taylor MC, Woodham BL, Board PG. Pesticides and Parkinson's disease. Biomed Pharmacother 1999;53:122–30. http://dx.doi.org/10.1016/ S0753-3322(99)80077-8.
- [49] Bjorklund G, Stejskal V, Urbina MA, Dadar M, Chirumbolo S, Mutter J. Metals and Parkinson's Disease: Mechanisms and Biochemical Processes. Curr Med Chem 2018;25:2198– 214. http://dx.doi.org/10.2174/ 0929867325666171129124616.
- [50] Hamza TH, Chen H, Hill-Burns EM, Rhodes SL, Montimurro J, Kay DM, et al. Genome-Wide Gene-Environment Study Identifies Glutamate Receptor Gene GRIN2A as a Parkinson's Disease Modifier Gene via Interaction with Coffee. PLOS Genet 2011;7:1–15. http://dx.doi.org/10.1371/journal.pgen.1002237.
- [51] Elbaz A, Carcaillon L, Kab S, Moisan F. Epidemiology of Parkinson's disease. Rev Neurol (Paris) 2016;172:14–26. http://dx.doi.org/10.1016/j.neurol.2015.09.012.
- [52] Taylor AE, Davies NM, Ware JJ, VanderWeele T, Smith GD, Munafò MR. Mendelian randomization in health research: Using appropriate genetic variants and avoiding biased estimates. Econ Hum Biol 2014;13:99–106. http://dx.doi.org/10.1016/j.ehb.2013.12.002.
- [53] Lawlor DA, Harbord RM, Sterne JAC, Timpson N, Davey Smith G. Mendelian randomization: Using genes as instruments for making causal inferences in epidemiology. Stat Med 2008;27:1133–63. http://dx.doi.org/10.1002/sim.3034.
- [54] Cornish AJ, Tomlinson IPM, Houlston RS. Mendelian randomisation: A powerful and inexpensive method for identifying and excluding non-genetic risk factors for

- colorectal cancer. Mol Aspects Med 2019;69:41–7. $\underline{\text{dx.doi.org/10.1016/j.mam.2019.01.002}}.$
- [55] Smith GD, Ebrahim S. "Mendelian randomization": can genetic epidemiology contribute to understanding environmental determinants of disease? Int J Epidemiol 2003;32:1–22. http://dx.doi.org/10.1093/ije/dyg070.
- [56] Sheehan NA, Didelez V, Burton PR, Tobin MD. Mendelian Randomisation and Causal Inference in Observational Epidemiology. PLOS Med 2008;5:1–6. http://dx.doi.org/10.1371/journal.pmed.0050177.
- [57] Smith GD, Ebrahim S. Mendelian randomization: prospects, potentials, and limitations. Int J Epidemiol 2004. http://dx.doi.org/10.1093/ije/dyh132.
- [58] Smith GD, Hemani G. Mendelian randomization: geneticanchorsfor causal inference in epidemiological studies. Hum Mol Genet 2014;23:R89–98. http://dx.doi.org/10.1093/hmg/ddu328.
- [59] Vlaar T, Kab S, Schwaab Y, Fréry N, Elbaz A, Moisan F. Association of Parkinson's disease with industry sectors: a French nationwide incidence study. Eur J Epidemiol 2018;33:1101–11. http://dx.doi.org/10.1007/s10654-018-0399-3.
- [60] Delamarre A, Meissner WG. Epidemiology, environmental risk factors and genetics of Parkinson's disease. Presse Med 2017;46:175–81. http://dx.doi.org/10.1016/j.lpm.2017.01.001.
- [61] Nicoletti A, Vasta R, Mostile G, Nicoletti G, Arabia G, Iliceto G, et al. Head trauma and Parkinson's disease: results from an Italian case-control study. Neurol Sci 2017;38:1835–9. http://dx.doi.org/10.1007/s10072-017-3076-5.
- [62] Factor SA, Weiner WJ. Prior history of head trauma in Parkinson's disease. Mov Disord 1991;6:225–9. http://dx.doi.org/10.1002/mds.870060306.
- [63] Weisskopf MG, O'Reilly E, Chen H, Schwarzschild MA, Ascherio A. Plasma urate and risk of Parkinson's disease. Am J Epidemiol 2007;166:561–7. http://dx.doi.org/10.1093/aje/kwm127.
- [64] Iwaki H, Ando R, Miyaue N, Tada S, Tsujii T, Yabe H, et al. One year safety and efficacy of inosine to increase the serum urate level for patients with Parkinson's disease in Japan. J Neurol Sci 2017;383:75–8. http://dx.doi.org/10.1016/j.jns.2017.10.030.
- [65] Kobylecki CJ, Nordestgaard BG, Afzal S. Plasma urate and risk of Parkinson's disease: A mendelian randomization study. Ann Neurol 2018;84:178–90. http://dx.doi.org/10.1002/ana.25292.
- [66] Kia DA, Noyce AJ, White J, Speed D, Nicolas A, collaborators I, et al. Mendelian randomization study shows no causal relationship between circulating urate levels and Parkinson's disease. Ann Neurol 2018;84:191–9. http://dx.doi.org/10.1002/ana.25294.
- [67] Schwarzschild MA, Ascherio A, Beal MF, Cudkowicz ME, Curhan GC, Hare JM, et al. Inosine to increase serum and cerebrospinal fluid urate in parkinson disease a randomized clinical trial. JAMA Neurol 2014;71:141–50. http://dx.doi.org/10.1001/jamaneurol.2013.5528.
- [68] Hernán MA, Takkouche B, Caamaño-Isorna F, Gestal-Otero JJ. A meta-analysis of coffee drinking, cigarette smoking, and the risk of Parkinson's disease. Ann Neurol 2002;52:276–84. http://dx.doi.org/10.1002/ana.10277.
- [69] Fujimaki M, Saiki S, Li Y, Kaga N, Taka H, Hatano T, et al. Serum caffeine and metabolites are reliable biomarkers of early Parkinson disease. Neurology 2018;90:e404–11. http://dx.doi.org/10.1212/WNL.000000000004888.

- [70] Cornelis MC, Munafo MR. Mendelian randomization studies of coffee and caffeine consumption. Nutrients 2018;10:1343. http://dx.doi.org/10.3390/nu10101343.
- [71] Pichler I, Del Greco MF, Gögele M, Lill CM, Bertram L, Do CB, et al. Serum Iron Levels and the Risk of Parkinson Disease: A Mendelian Randomization Study. PLOS Med 2013;10:1–13. http://dx.doi.org/10.1371/journal.pmed.1001462.
- [72] Noyce AJ, Kia DA, Hemani G, Nicolas A, Price TR, De Pablo-Fernandez E, et al. Estimating the causal influence of body mass index on risk of Parkinson disease: A Mendelian randomisation study. PLOS Med 2017;14:1–19. http://dx.doi.org/10.1371/journal.pmed.1002314.
- [73] Benn M, Nordestgaard BG, Frikke-Schmidt R, Tybjærg-Hansen A, Low LDL. cholesterol, PCSK9 and HMGCR genetic variation, and risk of Alzheimer's disease and Parkinson's disease: Mendelian Randomisation Study. BMJ 2017;357:j1648. http://dx.doi.org/10.1136/bmj.j1648.
- [74] Noyce AJ, Kia D, Heilbron K, Jepson J, Hemani G, Consortium IPDG, et al. Tendency towards being a "Morning person" increases risk of Parkinson's disease: evidence from Mendelian randomisation. BioRxiv 2018;288241. http://dx.doi.org/10.1101/288241.
- [75] Shokri-Kojori E, Wang G-J, Wiers CE, Demiral SB, Guo M, Kim SW, et al. β-Amyloid accumulation in the human brain after one night of sleep deprivation. Proc Natl Acad Sci 2018;115:4483–8. http://dx.doi.org/10.1073/ pnas.1721694115.
- [76] Yengo L, Sidorenko J, Kemper KE, Zheng Z, Wood AR, Weedon MN, et al. Meta-analysis of genome-wide association studies for height and body mass index in 700,000 individuals of European ancestry. Hum Mol Genet 2018;27:3641–9. http://dx.doi.org/10.1093/hmg/ddy271.
- [77] Dehghan A, Köttgen A, Yang Q, Hwang SJ, Kao WL, Rivadeneira F, et al. Association of three genetic loci with uric acid concentration and risk of gout: a genome-wide association study. Lancet 2008;372:1953–61. http://dx.doi.org/10.1016/S0140-6736(08)61343-4.
- [78] Kathiresan S, Melander O, Guiducci C, Surti A, Burtt NP, Rieder MJ, et al. Six new loci associated with blood lowdensity lipoprotein cholesterol, high-density lipoprotein cholesterol or triglycerides in humans. Nat Genet 2008;40:189–97. http://dx.doi.org/10.1038/ng.75.
- [79] Figueiredo JC, Hsu L, Hutter CM, Lin Y, Campbell PT, Baron JA, et al. Genome-Wide Diet-Gene Interaction Analyses for Risk of Colorectal Cancer. PLoS Genet 2014;10:e1004228. http://dx.doi.org/10.1371/journal.pgen.1004228.
- [80] Trampush JW, Yang MLZ, Yu J, Knowles E, Davies G, Liewald DC, et al. GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: A report from the COGENT consortium. Mol Psychiatry 2017;22:1651–2. http://dx.doi.org/10.1038/mp.2016.244.
- [81] De Moor MHM, Costa PT, Terracciano A, Krueger RF, De Geus EJC, Toshiko T, et al. Meta-analysis of genome-wide association studies for personality. Mol Psychiatry 2012;17:337–49. http://dx.doi.org/10.1038/mp.2010.128.
- [82] Hipp G, Vaillant M, Diederich NJ, Roomp K, Satagopam VP, Banda P, et al. The Luxembourg Parkinson's Study: a Comprehensive Approach for Stratification and Early Diagnosis. Front Aging Neurosci 2018;10:326. http://dx.doi.org/10.3389/FNAGI.2018.00326.