

RESEARCH

Open Access



A year of monitoring 20 mesophilic full-scale bioreactors reveals the existence of stable but different core microbiomes in bio-waste and wastewater anaerobic digestion systems

Magdalena Calusinska^{1*} , Xavier Goux¹, Marie Fossépré¹, Emilie E. L. Muller^{2,3}, Paul Wilmes² and Philippe Delfosse¹

Abstract

Background: Anaerobic digestion (AD) is a microbe-driven process of biomass decomposition to CH₄ and CO₂. In addition to renewable and cost-effective energy production, AD has emerged in the European Union as an environmentally friendly model of bio-waste valorisation and nutrient recycling. Nevertheless, due to the high diversity of uncharacterised microbes, a typical AD microbiome is still considered as “dark matter”.

Results: Using the high-throughput sequencing of small rRNA gene, and a monthly monitoring of the physico-chemical parameters for 20 different mesophilic full-scale bioreactors over 1 year, we generated a detailed view of AD microbial ecology towards a better understanding of factors that influence and shape these communities. By studying the broadly distributed OTUs present in over 80% of analysed samples, we identified putatively important core bacteria and archaea to the AD process that accounted for over 70% of the whole microbial community relative abundances. AD reactors localised at the wastewater treatment plants were shown to operate with distinct core microbiomes than the agricultural and bio-waste treating biogas units. We also showed that both the core microbiomes were composed of low (with average community abundance $\leq 1\%$) and highly abundant microbial populations; the vast majority of which remains yet uncharacterised, e.g. abundant candidate *Cloacimonetes*. Using non-metric multidimensional scaling, we observed microorganisms grouping into clusters that well reflected the origin of the samples, e.g. wastewater versus agricultural and bio-waste treating biogas units. The calculated diversity patterns differed markedly between the different community clusters, mainly due to the presence of highly diverse and dynamic transient species. Core microbial communities appeared relatively stable over the monitoring period.

Conclusions: In this study, we characterised microbial communities in different AD systems that were monitored over a 1-year period. Evidences were shown to support the concept of a core community driving the AD process, whereas the vast majority of dominant microorganisms remain yet to be characterised.

Keywords: Biogas, Microbial communities, Small rRNA gene amplicon high-throughput sequencing

*Correspondence: magdalena.calusinska@list.lu

¹ Environmental Research and Innovation Department, Luxembourg Institute of Science and Technology, 41 rue du Brill, 4422 Belvaux, Luxembourg

Full list of author information is available at the end of the article



Background

Microbial resource management (MRM), which refers to an optimal management of microbes with the aim to develop new products and to improve existing bio-processes, is at basis of numerous strategies in different domains, including renewable energy production, nutrient and water recycling, environmental safety and health [1, 2]. Anaerobic digestion is an example of an efficient MRM application, which relies on different classes of fermentative and syntrophic bacteria's interactions with methanogens to decompose organic compounds in an anaerobic environment into CO₂ and CH₄; the latter being an energy carrier [3]. With an estimated number of over 30,000 industrial installations worldwide, AD provides a permanent power production of 10,000 MW [4], and thus stable operation is highly desired. In addition to renewable and cost-effective energy production, AD has emerged in the European Union as an environmentally friendly model of bio-waste valorisation allowing both nutrient recovery from digestion residue, and a reduction in greenhouse gas emissions (in comparison to, e.g. aerobic composting or landfilling [5]).

In Luxembourg, the government has supported research and encouraged farmers and industries to produce energy from biomass through the AD process. More recently, the Third Industrial Revolution (TIR) strategy study by J. Rifkin mentions the important role that biogas can play in Luxembourg to complement the intermittent solar and wind power, especially through energy storage, on-demand energy production, or through the power-to-gas technology (<http://www.troisiemerevolutionindustrielle.lu/>). The same study encourages the circular bio-economy to be included in the food production systems, where biogas would be at the heart of a waste-as-nutrients circular process for a sustainable and circular agriculture. Anaerobic digestion has long been used to stabilize organic materials, mainly manure and sewage sludge, but nowadays its range of feeding substrate has been broadened to bio-waste (e.g. agricultural, municipal and food) and dedicated energy crops [6]. Whereas energy crops, including maize and immature cereals, are in most cases highly digestible substrates, competition may arise with their use as feed or food. On the contrary, according to the European Biogas Industry Association (EUBIA), estimated biogas yield (can vary according to the total solid content and biomass composition) from municipal solid waste (MSW) is similar to ley crops and can be twice higher than animal manure (<http://www.eubia.org/cms/wiki-biomass/anaerobic-digestion/>). Therefore, having seen the growing streams of organic waste as opportunity, in Luxembourg MSW is now converted into clean biogas which is directly injected into the local gas network. As an example, around 35,000 tonnes

of organic waste corresponding to 44% of organic household waste are converted yearly into methane in Mondercange biogas plant in southern Luxembourg.

Anaerobic digestion is a microbe-driven process; however, typical AD reactor microbiome is still considered as “dark matter”, mainly due to the huge diversity of uncharacterised microbes representing in many cases candidate phyla [7]. Recently, through the use of 16S ribosomal RNA (rRNA) gene high-throughput amplicon sequencing and/or metagenomics, a number of studies have attempted to explore the microbial community ecology of AD, e.g. [3, 8–14]. Although these studies have provided great insights into the structure, dynamics and functionality of microbial communities, most of them were limited to the municipal/industrial wastewater installations or farm reactors where animal manure and/or energy crops were the main feeding substrates. MSW-supplemented AD reactors were given much less attention (e.g. [12]) most probably due to the lower number of waste-valorising installations. Linking the dynamics of AD microbial community structures (e.g. using microbial ecology parameters such as community richness, diversity and evenness) with the performance of the AD system has also yielded conflicting results [15–17]. On the one hand, higher microbial diversity is considered as a reservoir of microbes with redundant metabolic pathways, which is desirable to ensure a functional stability of microbial communities in case of the changing environment [18]. Thus, higher species diversity is often correlated with more stable AD reactors [17]. On the other hand, functional stability might also be conferred by less diverse communities possibly expressing complementary pathways (niche complementarity [19]). In this sense, by avoiding direct competition over the exploitation of available resources, lower species richness could sustain higher structural stability of such microbial communities. While, highly enriched communities might be more quickly destabilised by, e.g. viral predation [20], a stable core community composed of roughly 63 abundant genus-level operational taxonomic units (OTUs), accounting for 68% of relative community abundance, has recently been reported for activated sludge ecosystem [21]. Six-year monitoring survey of 32 Danish full-scale AD reactors also showed that out of the huge diversity of microbes merely 300 OTUs represented 80% of total rRNA reads across reactors [8]. While, the high overall diversity of microbes reported in different studies (e.g. in a range of 3000–5000 OTUs [8, 9, 11]) mitigates our efforts of their functional characterisation which is practical for only a limited fraction of species in a given system, the question to ask is whether the whole diversity really matters in anaerobic reactors? Can we select a core of microorganisms common to most AD systems (e.g.

valorising agricultural by-products, energy crops, animal manure, sewage sludge and MSW, etc.) that would be structurally stable in different reactors? If yes, their further characterisation should become a research priority to show whether these core microbes reflect the population of putative key microbes to the AD process.

To address these questions, we first aimed to characterise the structures and stabilities of microbial communities, including bacteria and archaea, involved in biogas production in 20 full-scale AD bioreactors that were monitored over 1-year period at regular monthly time intervals. Second, using the core community concept, we intended to indicate putatively important organisms to the AD process. As a result, we confirmed that though being phylogenetically similar (represented by the same phyla) the different microbiomes were biogas unit-specific and structurally stable over the monitoring period. Furthermore, by associating the abundance and the distribution of microorganisms in the studied reactors we revealed the existence of a microbial core specific to the AD system. However, the microbial core of the full-scale bio-waste (agricultural and MSW) energy units largely differed from that of the AD reactors located at WWTPs. This confirms that these two main types of AD systems are operated by different microbial communities.

Results and discussion

Characteristics of the studied AD reactors

In total, 20 different full-scale mesophilic (temp. range from 33 to 44 °C) anaerobic reactors operating at 10 different biogas units (U-1–U-10) located in Belgium and Luxembourg were monitored during 1 year in regular monthly time series (Table 1). According to the main feeding substrates, the studied units were aggregated into four main AD categories, including: farm reactors fed mainly with agricultural residues (U-1 and U-2; acronym farm), reactors treating bio-waste (a mixture of agricultural residues including manure supplemented with MSW; U-3–U-6; bio-waste), a biogas unit treating uniquely sorted MSW including green waste (mostly garden and park residues; U-7; MSW), and anaerobic digesters of WWTPs (fed with sewage-activated sludge; U-8–U-10; WWTP-ADs). This classification scheme is stipulated and it aimed at reflecting the ordering based on the complexity/specificity of the feeding substrates (for the categories farm, bio-waste and MSW it reflects the decreasing ratio of manure towards MSW utilisation in the reactors; Fig. 1). All the reactors excluding U-7 (MSW) and WWTP-ADs were supplemented with different ratio of cattle manure (Table 1, Fig. 1). Reactor categories corresponding to farm, bio-waste and MSW will be commonly referred to as agricultural and bio-waste treating units, through the manuscript. Respectively,

WWTP-ADs will often be discussed as a separate category. Except for U-7 which was operated as horizontal plug-flow-type (PFR), the other reactors were all operated as completely stirred tank reactors (CSTRs, Additional file 1: Figure S1). U-7 was also the only unit equipped with a separate hydrolysis box (partially covered, non-heated storage room where MSW was stored for 2–3 days before being fed to the reactor), and the anaerobic digestion process was a dry-type (solid content over 15%). In the case of units U-3, U-4 and U-5 several digesters and post-digesters located at the same biogas unit, as well as storage tanks were included in the analysis. For the horizontal rectangular PFR-type U-7, the inlet (MK-beg), outlet (MK-end) and the middle part (MK-mid) were monitored, showing no significant changes in species composition between the different parts (see below). During the time of the monitoring, all reactors were stably operated and none of them reported overt failures, including the MSW-supplemented reactor receiving the highly diversified feeding substrates (Table 1 and personal communication with plant managers). The sludge retention time was in a range of 2–3 weeks in the case of the WWTP-ADs and 5–12 weeks for the remaining reactors. A wide range in physicochemical parameters was observed between all the reactors with pH ranging between 7.1 (WWTP-ADs) and 8.0 (farm), total inorganic carbon (TIC) between 1.2 (WWTP-ADs) and 6.2 (bio-waste; m³ of CO₂/m³ of slurry), ammonium–nitrogen (NH₄–N) between 0.4 (WWTP-ADs) and 5.0 (bio-waste; kg NH₄–N/m³ of sludge), total solids (TS) between 2.5 (WWTP-ADs) and 22.2 (MSW; % of TS/fresh sludge [w/w]), and volatile solids (VS) between 47.7 (WWTP-ADs) and 72.9 (farm; % of VS/% of total solid [w/w]). Details of these parameters values and volatile fatty acids (VFAs) concentrations at different sampling points are provided in Additional file 2: Table S1.

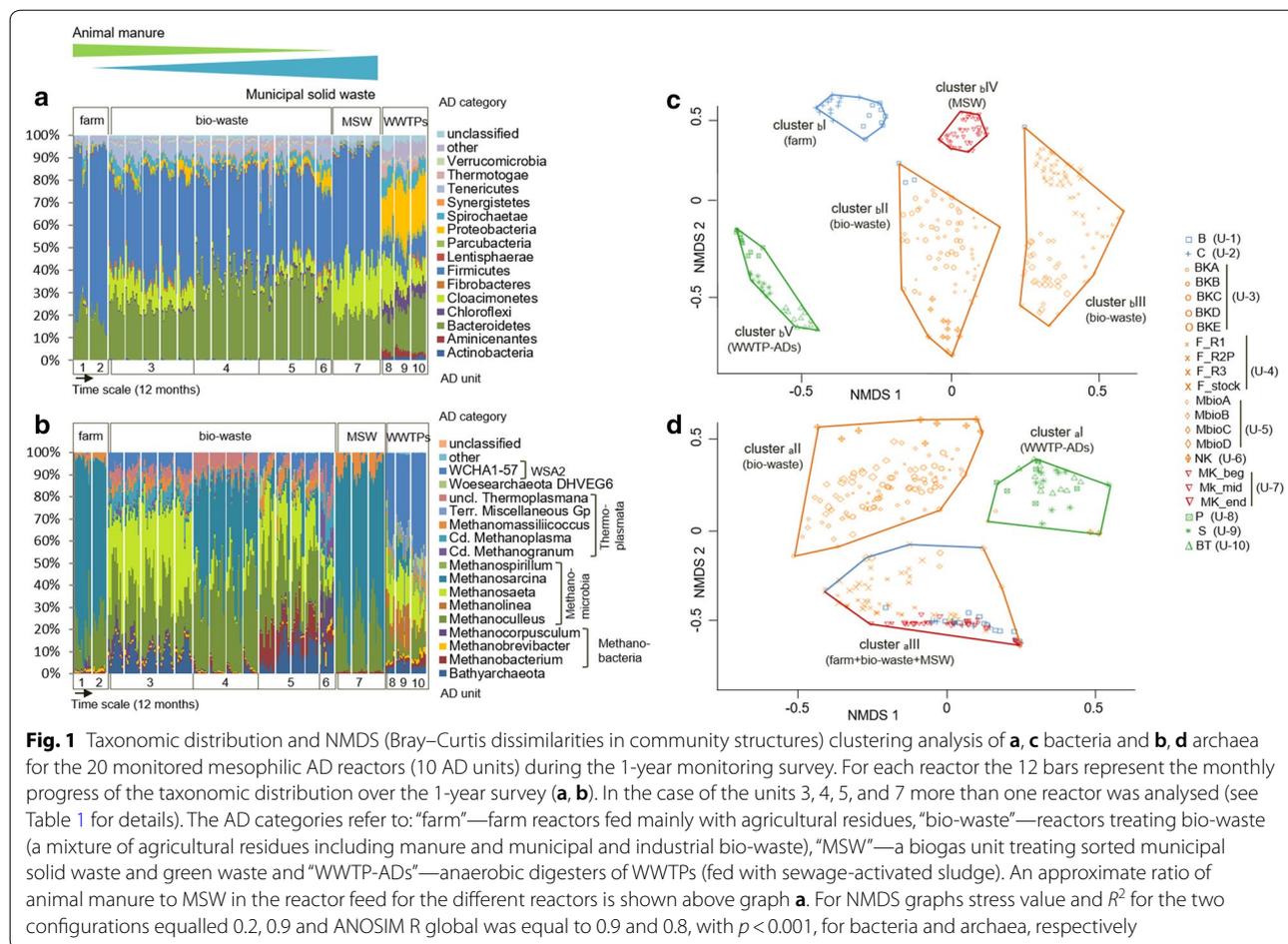
Taxonomic distribution of bacteria and archaea in AD reactors based on the 16S rRNA gene amplicons

In our study, a year of monitoring microbial communities (using a newly optimised protocols for bacterial and archaeal 16S rRNA gene amplicon sequencing; Additional files 3 and 4) revealed the existence of different but structurally stable microbiomes in wastewater and agricultural and bio-waste anaerobic digestion systems. Sequencing of bacterial 16S rRNA gene amplicons from over 250 samples (subsampling to 11,550 quality-trimmed reads per sample), resulted in a total of 5938 bacterial OTUs (₆OTUs defined at 97% sequence similarity and represented by more than one sequence), representing 55 known and candidate phyla (Additional file 5: Table S5). Rarefaction curves observed based on the species richness did not reach the plateau; however, the

Table 1 Studied AD reactors and their physicochemical characteristics

Reactor type	Main feeding substrate	AD unit	Reactor acronym/m	Reactor category	pH	Temp. range °C	TIC	NH ₄ -N	TS	VS
On-farm AD reactors fed mainly with agricultural residues	Maize and grass silages, cow manure and slurry (around 70%), bread	U-1	B	Fd	7.9±0.2	39–40	3.4±0.9	3.3±0.9	8.6±2.2	72.1±4.8
	Maize silage, cow manure (around 70%), minor food waste	U-2	C	Fd	8.0±0.1	42–44	4.7±0.6	3.9±1.1	10.2±1.4	72.6±2.9
AD reactors treating a mixture of agricultural residues (mainly manure) and bio-waste	Cow manure and slurry, grass, maize and cereal silages, dairy waste	U-3	BKA	Fd-1	7.6±0.2	33–39	2.7±0.4	2.1±0.6	8.3±0.6	72.9±2.9
			BKB	Fd-2	7.7±0.2		2.9±0.6	2.1±0.6	8.0±0.9	72.5±2.9
			BKC	Pd-1	7.9±0.2		3.3±0.3	2.6±0.9	7.2±0.3	69.5±1.9
	Cow manure and slurry, cereal waste, waste from food industries, maize silage, grass		BKD	Pd-2	7.9±0.2		3.3±0.4	2.6±0.7	7.3±0.4	69.1±1.8
			BKE	St	7.8±0.2	nh	3.6±0.3	2.9±0.5	6.7±0.5	67.1±1.9
			F-R1	Fd-1	7.8±0.1	39	3.9±0.5	3.5±0.8	10.1±0.8	65.9±1.9
Municipal and green waste treating AD reactors	Manure, cereal waste, grass silage, organic waste from supermarkets and canteens		F-R2P	Pd	7.9±0.1		5.6±0.5	4.5±0.8	8.5±0.6	61.2±1.2
			F-R3	Fd-3	7.9±0.1		4.8±0.6	3.8±0.8	8.8±0.5	62.5±1.6
	Manure, green waste, maize silage, waste from food industries		F-stock	St	7.9±0.1	nh	6.2±0.5	5.0±0.9	7.6±0.4	57.6±2.5
			MbioA	Fd-1	7.5±0.2	37–38	3.5±0.7	3.6±1.5	6.7±0.9	66.5±2.5
			MbioB	Fd-2	7.6±0.2		2.3±0.4	1.9±1.1	10.8±3.7	70.6±2.6
WWTP-specific AD reactors	Manure, green waste, maize silage, waste from food industries		MbioC	Pd	7.6±0.1		3.3±0.5	2.7±1.1	8.8±0.9	68.1±2.2
			MbioD	St	7.6±0.2	nh	3.5±0.4	2.8±0.6	7.7±1.7	65.7±2.4
	Yard green waste (leaves, trees), organic waste from households and canteens		NK	Fd	7.6±0.3	36–37	3.4±0.6	2.4±0.8	7.8±0.9	66.2±2.2
			MK-beg	Fd	7.8±0.2	39	4.9±0.8	3.6±1.5	21.6±1.6	50.8±2.7
Thickened sewage sludge from wastewater treatment	Yard green waste (leaves, trees), organic waste from households and canteens		MK-mid		7.8±0.2		4.6±0.8	3.4±1.5	22.2±1.3	51.7±3.2
			MK-end		7.8±0.1		4.6±0.9	3.5±1.6	21.8±1.7	50.7±2.8
	Thickened sewage sludge from wastewater treatment	U-8	P	Fd	7.2±0.1	35	1.3±0.3	0.4±0.5	2.5±0.4	47.7±3.3
		U-9	S	Fd	7.1±0.1	36	1.2±0.2	0.7±0.5	3.8±0.4	55.6±1.8
	U-10	BT	Fd	7.2±0.1	36	1.4±0.2	1.3±0.9	3.0±0.2	49.0±1.6	

Fd fed digester, Pd post-digester, St storage tank, TIC total inorganic carbon (m³ of CO₂/m³ of sludge), NH₄-N total ammonium-nitrogen (kg NH₄-N/m³ of sludge), TS total solids (% of total solid/fresh sludge [w/w]), VS volatile solids (% of volatile solid/% of total solid [w/w]), nh non-heated



rank-abundance curves showed that OTUs representing at least 0.008% of the bacterial community were detected (Additional file 6: Figure S6). In line with our results, previous studies of full-scale AD reactors reported a total number of bacterial OTUs to be in a range of 3000–5000 [9, 11]. Twelve bacterial phyla were represented in all analysed samples, and nine were at $\geq 1\%$ average 16S rRNA gene amplicons reads abundance (decreasing order: *Firmicutes*, *Bacteroidetes*, *Cloacimonetes* previously known as “candidate phylum WWE1”, *Proteobacteria*, *Tenericutes*, *Spirochaetes*, *Aminicenantes*, *Chloroflexi*, *Parcubacteria*). Bacterial communities of agricultural and bio-waste treating units were mainly dominated by *Firmicutes* ($46.4\% \pm 11.8$ of sequencing reads per sample) and *Bacteroidetes* ($28.5\% \pm 8.5$) with a minor proportion of reads assigned to *Cloacimonetes* ($8.0\% \pm 7.5$) and *Tenericutes* ($5.3\% \pm 3.0$). WWTP-ADs had distinct bacterial communities with *Proteobacteria* ($22.6\% \pm 4.3$), *Bacteroidetes* ($20.3\% \pm 4.5$), *Cloacimonetes* ($12.0\% \pm 6.2$) and *Firmicutes* ($9.5\% \pm 2.1$; Fig. 1a) as principal phyla. While, in general, a more dominant phylum was represented by a higher number of β OTUs, some phyla like *Cloacimonetes*

and *Proteobacteria* did not follow this trend. Only 18.7% of β OTUs were common to the two anaerobic digestion systems, while 53.7 and 27.6% of bacterial OTUs were specific to agricultural and bio-waste treating units and WWTP-ADs, respectively. The predominance of *Firmicutes* and *Bacteroidetes* has been previously shown (using both 16S rRNA amplicon sequencing and metagenomics) in many full-scale (mainly manure supplemented) AD reactors [3, 9, 13, 22]; while other taxonomic divisions including *Proteobacteria* and *Spirochaetes* [11] or *Chloroflexi* and *Proteobacteria* [23] dominated mainly WWTP-specific (sewage sludge-supplemented) AD reactors previously studied. The presence of candidate phylum *Cloacimonetes* has less often been reported in the literature, even though it clearly dominated bacterial communities in some full-scale digesters [24–26] and lab-scale reactors [27, 28].

Concerning archaea, the high-throughput analysis of quality-trimmed reads subsampled to 2070 reads per sample resulted in 89 archaeal OTUs (α OTU), out of which 66 were assigned to *Euryarchaeota* (86.6% of total reads), six to *Woesearchaeota* (0.6%), five to

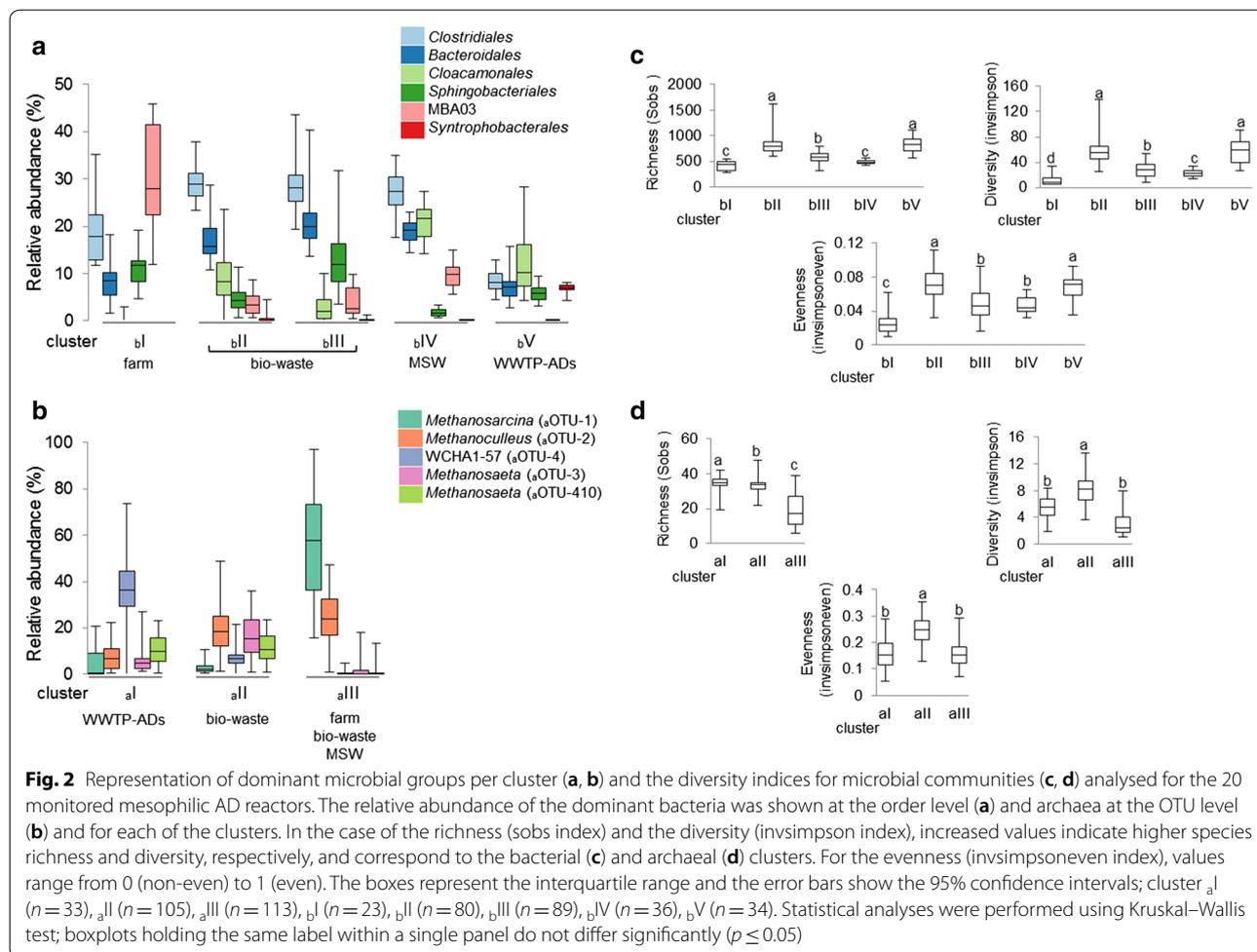
Bathyarchaeota (4.8%) and three to WSA2 (7.9%; Fig. 1b). The rarefaction curves based on the species richness reached the plateau, suggesting that the description of the archaeal diversity was nearly exhaustive (Additional file 6: Figure S6). *Methanomicrobia* (71.2% ± 19.2) and *Thermoplasmata* (11.0% ± 7.3; in U-6 they constituted 31.6% ± 5.8 of archaeal community) were prevailing classes in all AD units, while *Methanobacteria* accounted for around 4.3% ± 4.6 of all archaea. The highest abundance of *Bathyarchaeota* was detected in the ADs located at U-3 (8.3% ± 4.7) and U-5 (10.0% ± 6.2). Next to *Methanomicrobia* (44.2% ± 13.8), WCHA1-57 (39.0% ± 10.3) dominated in WWTP-ADs. In contrast to bacteria, around 56.2% of archaeal OTUs were common to the two main anaerobic digestion systems. In total 31.5% of aOTUs were specific to agricultural and bio-waste treating units and only 12.3% to WWTP-ADs.

Diversity versus stability of bacterial populations in AD reactors

To analyse the variation in microbial communities within a single reactor and across different biogas units we used the non-metric multidimensional scaling (NMDS) of the calculated Bray–Curtis dissimilarities in community structures at the OTU level (Fig. 1c). The NMDS clustering of bacterial communities visibly separated (ANOSIM R global equalled 0.9 with $p < 0.001$) AD reactors treating activated sludge located at the WWTPs (cluster $_bV$) from agricultural and bio-waste treating units (clusters $_bI$ to $_bIV$), confirming the presence of different microbiomes in the two AD systems [13]. Additionally, clustering of samples resulted in biogas unit- and sampling time-specific segregation indicating the strong influence of operational conditions and confirming previous observations [3, 9, 10, 13]. The formation of the tight collections of points indicated that the variation over time of microbial communities within a reactor and/or AD plant is lower than in the cross-section. Former clustering analysis of 38 samples collected from 29 different full-scale AD installations indicated the dominance of either *Bacteroidales* or *Clostridiales* in two different clusters grouping mesophilic sludge digesters [9]. In our study, next to mainly *Clostridiales* and *Bacteroidales*-dominated clusters $_bII$ and $_bIII$ (Fig. 2a), we show the presence of MBA03-dominated (Firmicutes) cluster $_bI$ specific to farm reactors, cluster $_bIV$ (MSW) where three bacterial orders including *Clostridiales*, *Cloacamonales* and *Bacteroidales* were co-abundant, and WWTP AD-specific cluster $_bV$ dominated by *Cloacamonales*, *Clostridiales*, *Bacteroidales*, *Sphingobacteriales* and *Syntrophobacteriales*. The co-occurrence of MBA03 which was evidenced to be an electroactive genus [29]

and *Methanosarcina* (see below), might suggest the direct interspecies electron transfer (DIET) to potentially be an effective form of syntrophy in farm methanogenic reactors [30].

In terms of bacterial diversity, ADs reactors specific to WWTPs (cluster $_bV$) were similar to the bio-waste units grouping in cluster $_bII$ (Fig. 2c; Additional file 6: Figure S7). Bacterial communities from farm reactors (cluster $_bI$) and MSW (cluster $_bIV$) were the least diverse and evenly distributed ($p \leq 0.05$), even though U-7 was receiving highly diversified over 1-year MSW feed. In these reactors, bacterial diversity was significantly negatively correlated to mainly TS and NH_4-N , TIC, total VFAs (mainly acetate) and pH ($p \leq 0.01$; Fig. 3; Additional file 7: Table S6). While it has been shown that the initial community evenness favours community functionality under selective stress [18], communities dominated by single species could assure similar resistance to the perturbation, provided that they are tolerant to it. Both strategies were observed in the studied reactors: (1) less diverse communities that were generally dominated by single OTUs (clusters $_bI$ and $_bIV$; interestingly they were correlated with higher TS), and (2) the more diverse that were characterised by a higher OTUs evenness (clusters $_bII$ and $_bIII$ and $_bV$). Interestingly, the stabilities of these communities were similar (Additional file 6: Figure S8), assuring stable operation (i.e. $7.0 \leq pH \leq 8.0$, low VFAs concentrations and stable biogas production as reported by all plants operators) of the studied AD reactors during the monitoring period (Table 1 and Additional file 2: Table S1). Therefore, to what extent increased species diversity increases the functional stability of the AD system? What are the mechanisms that optimise diversity within a given biological community? According to the insurance hypothesis proposed by Yachi and Loreau [31], species richness at which specific ecosystem becomes functionally redundant largely depends on the way the different species interact and respond to the changing environment. Moreover, according to [32], the more a species is functionally dependent on the activity of another, fewer species are necessary to maintain ecosystem stability. In this sense, even highly productive communities might exhibit reduced species diversity [19], since the high functional redundancy required in more variable environments (e.g. soil, ocean) would no longer be essential in a relatively stable and highly specialised methanogenic world. These observations may suggest that a more diverse community does not necessarily mean a better adapted to anaerobic digestion. Indeed, U-7 receiving MSW was characterised with the highest community structural stability (Additional file 6: Figure S8A), that was at the same time one of the least diverse



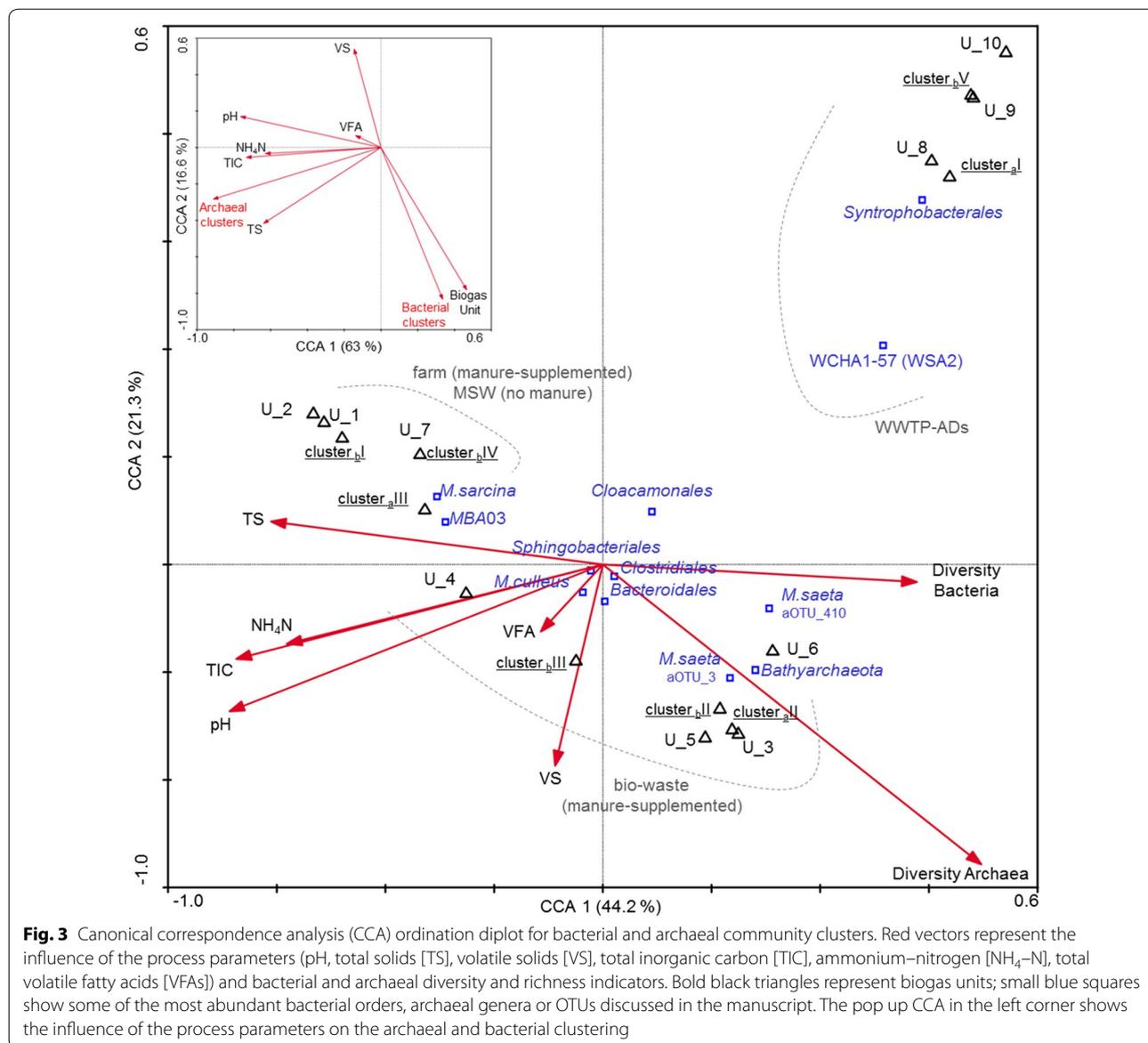
and even (in this case the dominant b_{OTU} represented the candidate phylum *Cloacimonetes*).

Archaeal community clusters and diversity patterns

Archaeal communities of WWTPs (cluster a_I) differed significantly (ANOSIM R global equalled 0.9 with $p < 0.001$) from agricultural and bio-waste treating units (clusters a_{II} and a_{III}) and the separation was mainly attributed to the different NH_4-N and TIC parameters (Figs. 1d and 3). The diversity patterns varied between the different AD reactors (Fig. 2d; Additional file 6: Figure S9); however, similar trends of diversity and richness were observed for archaeal communities from reactors characterised by the same type of a feeding regime. In general, archaeal cluster a_{II} (bio-waste) was characterised by the highest species diversity and evenness in contrast to the cluster a_{III} (mainly regrouping farm and MSW) that was lower (Fig. 2d). Interestingly, an increased relative abundance of *Bathyarchaeota* (a_{OTU-6}) was positively correlated ($p < 0.001$) with increased archaeal

diversity in the studied reactors (Fig. 3; Additional file 7: Table S6).

On average *Methanosarcina* was the most abundant and omnipresent archaeon (Figs. 1b and 2b); however, it dominated over *Methanosaeta* only in 46.8% of analysed samples (mainly cluster a_{III}). As previously reported, its dominance was correlated with a higher concentration of acetate and ammonium–nitrogen (Additional file 7: Table S6; [12]), and a higher TS content of a reactor (Fig. 3). In the other half of the samples, two *Methanosaeta* a_{OTU-3} and $a_{OTU-410}$ were co-abundant (mainly cluster a_{II} and to a lower extent a_I), and correlated with the increased abundance of *Bathyarchaeota*. Co-occurrence of *Bathyarchaeota* with some *Methanomicrobia*, including *Methanosaeta* has been previously reported across different terrestrial settings [33]. Miscellaneous *Crenarchaeota* Group (MCG) renamed to *Bathyarchaeota* has recently been shown to encode in its genome genes necessary for hydrogenotrophic and methylotrophic methanogenesis, including the methyl Co-A reductase [34]. Its co-dominance with *Methanosaeta*



would thus assure the presence of the gene pool assuring the methane production by all three known methanogenic pathways, including acetoclastic, hydrogenotrophic and methylotrophic. The dominance of the unknown WSA2 archaeon (WCHA1-57, _aOTU-4) in all studied WWTP-ADs (cluster _aI) opposes to the study of 32 Danish full-scale reactors located at 20 WWTPs, where between 60 and 80% of reads were assigned to *Methanosaeta* [35]. Our result is unlikely to be an artefact resulting from the assay design, since this archaeon was not dominating elsewhere in the studied reactors. Moreover, the most represented archaeal group in the study of Rivière et al. [36] was affiliated with WSA2 as well.

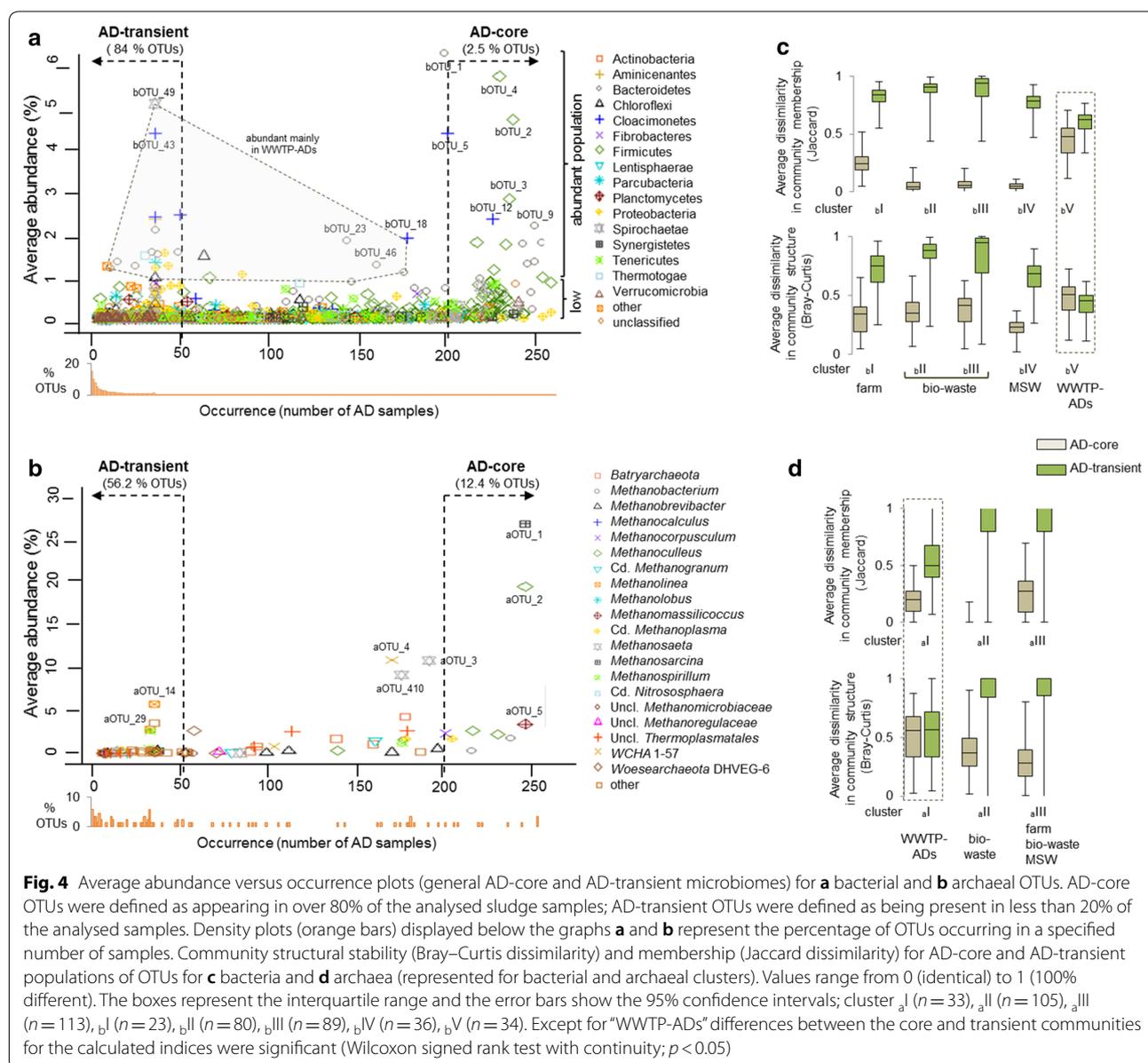
Distribution and stability of bacterial _bOTUs across the studied full-scale AD reactors

It has been proposed, that commonly occurring organisms appearing in most of the microbial communities associated with a particular environment, are likely important to the functioning of the whole community [37]. Revealing these stable and consistent components across different anaerobic digestion systems and defining the core (structural and functional) microbiome is important to the understanding of the whole process and could guide future manipulation of communities to attain a desired outcome. Detailed analysis of the clusters revealed that some OTUs were shared between all the reactors (except for the WWTP ADs), regardless the

feeding regime (i.e. the ratio of manure to MSW in the feed). In line with [1], we initially defined as a general AD-core microorganisms occurring in 80% of the studied AD samples. Both abundant and rare ($\leq 1\%$ average reads abundance) OTUs were considered, and not only the top abundant per sample as previously proposed [38]. Except for WWTP-ADs that were shown to operate with a distinct core microbiome (see below), 2.5% of bacterial OTUs assigned to AD-core accounted for $70.3\% \pm 12.5\%$ average bacterial read abundance in the reactors. AD-core OTUs were typical AD members, mainly assigned to *Firmicutes* (82 OTUs) and *Bacteroidetes* (28 OTUs). As a general trend we confirmed that most of the dominant

(with the highest average abundances across samples) bacterial OTUs were at the same time the most broadly distributed (Fig. 4a; [38]). However, there was also a considerable population of low abundant bacteria having the core community characteristics.

In line with the Pareto concept of species distribution in relation to their ecological tasks [39] applied in different microbial ecology studies [40, 41], abundant microorganisms (up to 20% top abundant species) would process 80% of the energy flux in the environment. In this sense, abundant AD-core microbes could be regarded as “key players” of the process, presumably performing the most important environmental tasks in their niche



[1]. Indeed, an optimal microbial metabolism operating close to the Pareto value has recently been proposed for AD [42]. Nevertheless, the dominance in the community might not correlate with the activity of a specific microbe, meaning that bacterial abundances not always reflect growth rates or activity [43]. In that study, some of the low abundant bacteria seemed the most active in the community. Accordingly, low abundant AD-core populations of microbes might exert a disproportionately large effect on the functionality of the community by, e.g. performing a very specific task. Consequently, their persistence might be occasionally favoured; however, at a frequency high enough to retain their community membership [17]. As an example, syntrophic acetate-oxidizing bacteria (SAOB) have been shown to occupy a unique functional niche in the microbiome of some ADs (mainly operated at high $\text{NH}_4\text{-N}$ load), at the same time representing a low abundant community of microbes [44].

Consistent with previous reports [38], 84.0% of all bacterial OTUs were assigned to AD-transient bOTUs (defined as being present in less than 20% of analysed samples and mostly detected at single measurement points), representing $3.5\% \pm 3.8$ of the relative bacterial abundances. They belonged mainly to *Firmicutes* (1742 OTUs), *Proteobacteria* (802 OTUs) and *Bacteroidetes* (615 OTUs). The remaining 790 bacterial OTUs that were neither classified as core nor transient were only shared between some bioreactors, and on average constituted $26.2\% \pm 10.0$ of bacterial abundances in the different reactors. Roughly 150 of these bOTUs were common to all the bacterial clusters, and most of them did not persist in the reactor over time. A few of them were locally abundant (also in WWTP-ADs-core), including bOTU-18 classified to *Cloacimonetes* and several other *Bacteroidetes* bOTUs , e.g. bOTU-23 or bOTU-46 .

To evaluate the extent to which the AD-core and AD-transient bacteria contribute to the observed bacterial diversity patterns and community structural stability in the studied AD reactors, we calculated separately for the core and transient populations the dissimilarity in community membership and structure over time. As a result, AD-core bacterial communities appeared quite stable in the studied reactors over the year, both in terms of the community membership and structure (except for WWTP-ADs; Fig. 4c; Additional file 6: Figure S10). By contrast, concerning the AD-transient communities, the dissimilarities in the community membership and structure were very high for all studied reactors. Except for WWTP-ADs, only 16 out of 4997 AD-transient bOTUs were abundant ($\geq 1\%$ of relative abundance) in at least one sample. Out of these, nine appeared at the same time point in serial reactors located at the same biogas unit, suggesting that they were supplied to the reactor with

the influent streams. Only two of these transient bOTUs were detected in more than two consecutive samplings, suggesting that most of the AD-transient microbes were not able to develop on a longer term (at least not over 1 month period) in AD environment, and contrasting with previous findings revealed for WWTP ADs [38]. The observed differences might result from the sampling intervals that equalled 1 month (during 1 year) in our study versus 2–7 days (during 1 month) [38]. Therefore, while our core concept relates to the microbes that were present at least during 1 year in a reactor, what was observed in the other study can only be valid at monthly basis.

As stated by [1] and the Hubbell theory, plenty of rare species happen to be there, sometimes without much relation to the functionality of a particular environment. Therefore, even if we cannot exclude some of the transient microbes being important to the process, at this stage of our knowledge, we should probably first focus on characterising the most abundant and/or widespread core species.

Distribution and stability of archaeal aOTUs across the studied full-scale AD reactors

AD-core archaea representing 12.4% of aOTUs (assigned exclusively to *Euryarchaeota*) accounted for $75.3\% \pm 19.4$ of the relative archaeal reads abundances across samples (Fig. 4b). The presence of three aOTUs in all analysed reactors (at every sampling point) corresponding to *Methanosarcina* (aOTU-1), *Methanoculleus* (aOTU-2) and *Methanomassiliicoccus* (aOTU-5) assures the potential to produce methane through any of the three methanogenic pathways [45]. Out of the 50 aOTUs assigned as AD-transient archaea, some were locally abundant only in WWTP-ADs, confirming that this type of reactors operates with different archaeal core communities. They only accounted for an average of $0.4\% \pm 0.3$ of the relative archaeal abundance across the agricultural and bio-waste treating units. The abundance of aOTUs that was neither assigned to AD-core nor AD-transient populations varied from 83.5% in the case of some WWTP-ADs, to as little as 0.1% for unit U-7 (MSW). For agricultural and bio-waste treating units these OTUs accounted for an average of $0.4\% \pm 0.3$. This observation shows that in contrast to bacteria, mainly process-specific archaea are present in AD reactors.

Based on the calculated Jaccard and Bray–Curtis indices, core archaeal communities were less stable than bacterial cores (Fig. 4c, d). However, the higher standard variations mostly resulted from the lower species diversity and sparse occurrence of some archaea in the different reactors.

Bacterial and archaeal core communities specific to WWTP-ADs

Until now, most attempts to characterise core bacterial communities of anaerobic reactors were mainly restricted to WWTP AD facilities. The very first trial was limited by the low throughput of Sanger sequencing [36]. According to that report, bacterial core was composed of six β OTUs affiliated with *Chloroflexi*, *Betaproteobacteria*, *Bacteroidetes* and *Synergistetes*. More recently, using a high-throughput 16S rRNA gene amplicon sequencing of 40 sludge samples collected from seven WWTP-ADs located in China, a total of 31 β OTUs (mainly assigned to *Bacteroidetes*, *Proteobacteria* and *Firmicutes*) with abundance greater than 3% of total reads in each digester were assigned to the core community [46]. Similarly to our results, by analysing 32 ADs located at 20 WWTPs in Denmark, Kirkegaard and collaborators [8] identified 300 abundant core organisms that accounted for 80% of reads. In that study, a few candidate phyla, including *Fermentibacteria* (Hyd-24-12), *Aminicenantes* (OP8) and *Atribacteria* (OP9) were present along with the commonly described *Firmicutes*, *Proteobacteria* and *Bacteroidetes*.

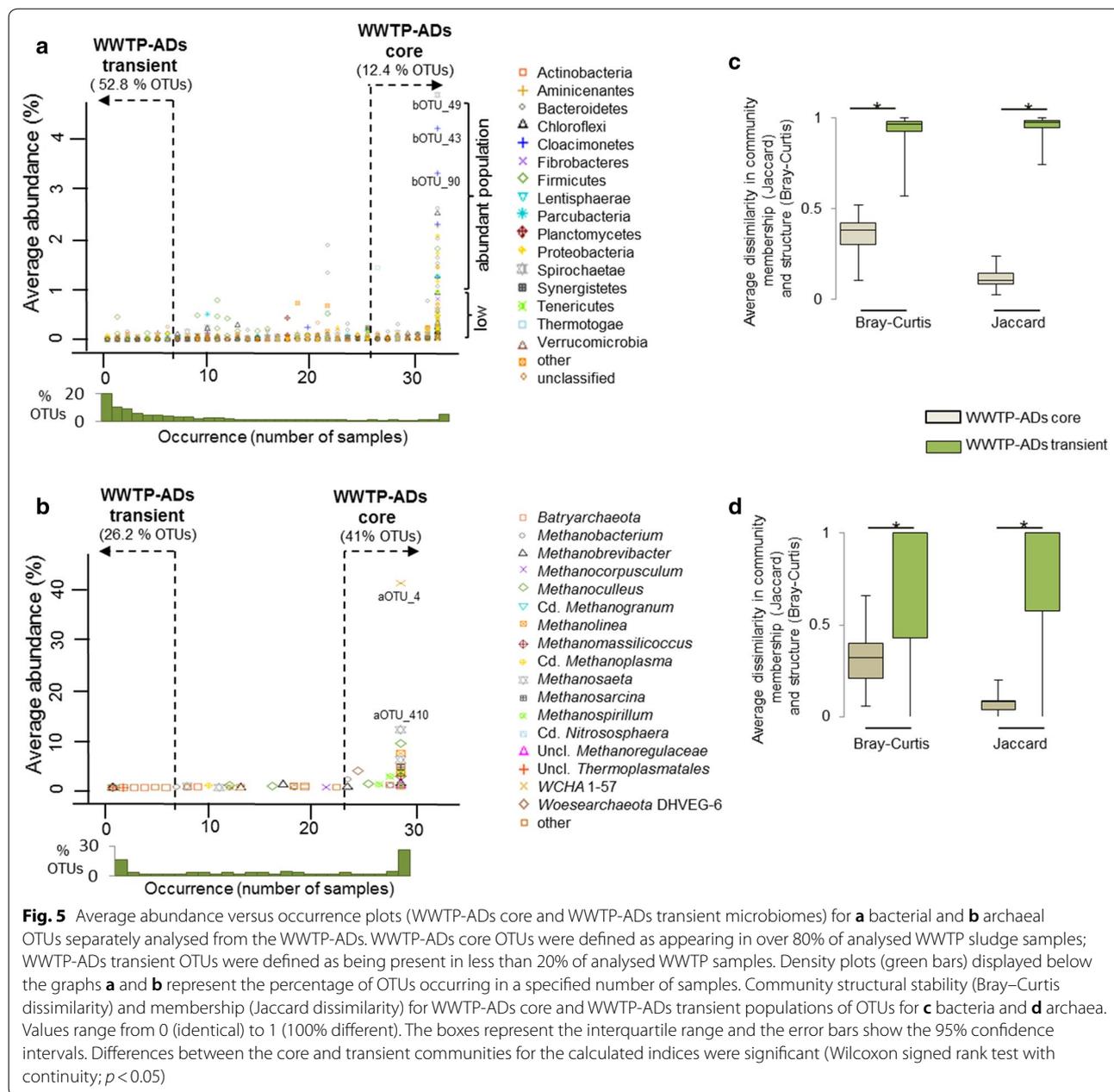
Since the observed AD-core did not show any core characteristics for the studied WWTPs ADs, we evaluated these reactors separately. The resulting WWTP-ADs core β OTUs represented 12.4% of bacterial richness and accounted for $82.0\% \pm 6.2$ of relative bacterial abundance (Fig. 5a). Their taxonomic composition was much different from the general AD-core and the majority of WWTP-ADs core β OTUs was assigned to *Proteobacteria* (103 β OTUs), *Bacteroidetes* (57 β OTUs), *Firmicutes* (36 β OTUs) and *Chloroflexi* (30 β OTUs). In terms of the relative abundance *Proteobacteria*, *Bacteroidetes* and *Cloacimonetes* were the dominant phyla. Out of the 342 WWTP-ADs core β OTUs, 24 represented candidate bacterial phyla and all together accounted for an average of 17.8% of the relative bacteria abundance in the studied WWTP ADs (Additional file 5: Table S5). There were only 27 β OTUs common to the general AD-core and WWTP-ADs core, mainly assigned to *Bacteroidetes*, *Firmicutes* and *Proteobacteria*; showing little species overlap between the two AD systems. Three most abundant (relative abundance $\geq 3\%$) WWTP-ADs core β OTUs were not AD-core, and were assigned to *Cloacimonetes* (β OTU-43 and β OTU-90) and one to *Spirochaetes* (β OTU-49). Around 89% of β OTUs (representing on average $1.44\% \pm 0.8$ of the reads) were classified to WWTP-ADs transient β OTUs, and were mainly assigned to *Firmicutes* (328 OTUs), *Proteobacteria* (297 OTUs) and *Chloroflexi* (107 OTUs). Even though, according to [38] the major fraction of microbial populations found in WWTP-AD reactors is able to actively grow, we think

that the occurrence of WWTP-ADs transient OTUs is rather related to the reactor feed residue and not to competitive metabolism. Interestingly, it has been shown for other WWTP-ADs that even some of the most abundant microbes (e.g. *Ca. Microthrix*) were related to influent streams as well [35]. In our study, three *Ca. Microthrix* β OTUs (β OTU-351, 882 and 1418) were assigned to the WWTP-ADs core; however, they represented the low abundant population of bacteria. *Ca. Microthrix* is a known aerobe and was shown to temporarily dominate aerobic tanks of the WWTPs in Luxembourg [47]. Regarding the relatively short sludge retention time and the fact that sewage-activated sludge is the only feeding substrate of the WWTP-AD reactors, it may happen that some of the WWTP-ADs core bacteria do not actively contribute to the AD process and they are present because they were fed into the reactor.

Archaeal core specific to WWTP-ADs was composed of 25 α OTUs (21 classified to *Euryarchaeota*, three to *Bathyarchaeota* and one to *Woesearchaeota*) and represented $97.9\% \pm 0.8$ of the relative archaeal abundance per sample. Five WWTP-ADs core α OTUs were also designated AD-core OTUs, including α OTU-2 (*Methanoculleus* sp.), α OTU-1 (*Methanosarcina* sp.), α OTU-5 (*Methanomassiliicoccus* sp.), α OTU-48 (*Methanobacterium* sp.) and α OTU-3 (*Methanosaeta* sp.). Widespread in WWTP-ADs and the most abundant α OTU-4 (relative abundance in WWTP-ADs was $38.4\% \pm 10.5$) assigned to an unclassified WSA2, was only present in half of the samples from agricultural and bio-waste treating AD units, and at much lower abundance (Fig. 2b). While there were no α OTUs representing the phylum *Bathyarchaeota* in the general AD-core, two were classified as WWTP-ADs core α OTUs (α OTU-6 and α OTU-15). Similar to the general AD-core, WWTP-ADs transient α OTUs were scarce in the reactors ($0.1\% \pm 0.1$ of the relative archaea abundance). Twenty α OTUs were neither core nor transient, and accounted for $2.1\% \pm 0.7$ of the relative archaea abundance. They were mainly assigned to *Euryarchaeota*. Core archaeal population of WWTP ADs seemed more stable than the general AD-core (Fig. 5d). This might be related to the biogranules which form in WWTP systems, thus providing a protective microenvironment to the methanogens situated inside the granules; in comparison to the non-layered microbial distribution observed in non-WWTP AD systems [48].

Abundance of candidate phyla in AD reactors

In total, around 450 β OTUs were assigned to candidate bacterial phyla and their abundance differed significantly in the different AD systems. Their relative abundance was the lowest in farm reactors (on average 0.93% of bacterial community), followed by the bio-waste (7.56%)



and WWTP-ADs (19.9%). Candidate phyla were the most abundant in a MSW-fed reactor (U-7), where they accounted for 21.15% of the whole bacterial community (mainly attributed to the dominance of *Cloacimonetes* _bOTU-5). Five OTUs representing candidate phyla *Parcubacteria* (formerly known as OD1; 1.9% ± 0.8 average bacterial abundance) and two *Aminicenantes* (OP8; 2.5% ± 1.1 average bacterial abundance) formed part of the abundant WWTP-ADs core population of bacteria. Except for farm reactors, *Cloacimonetes* was dominant between the candidate phyla (from 60.1 to 89.8% of

average abundance). *Cloacimonales* were the most abundant bacterial order in WWTP-ADs, but the dominant *Cloacimonetes* OTUs were different from those dominating in agricultural and bio-waste treating units (Fig. 6a). A neighbor-joining tree constructed using the 16S rRNA *Cloacimonetes* OTUs identified in this study and other phylogenetically similar sequences supported the existence of a high diversity within the *Cloacimonetes* phylum. Ten major phlotypes were distinguished, clearly separating _bOTUs according to the anaerobic digestion system. Phlotype A grouped _bOTUs that were dominating in

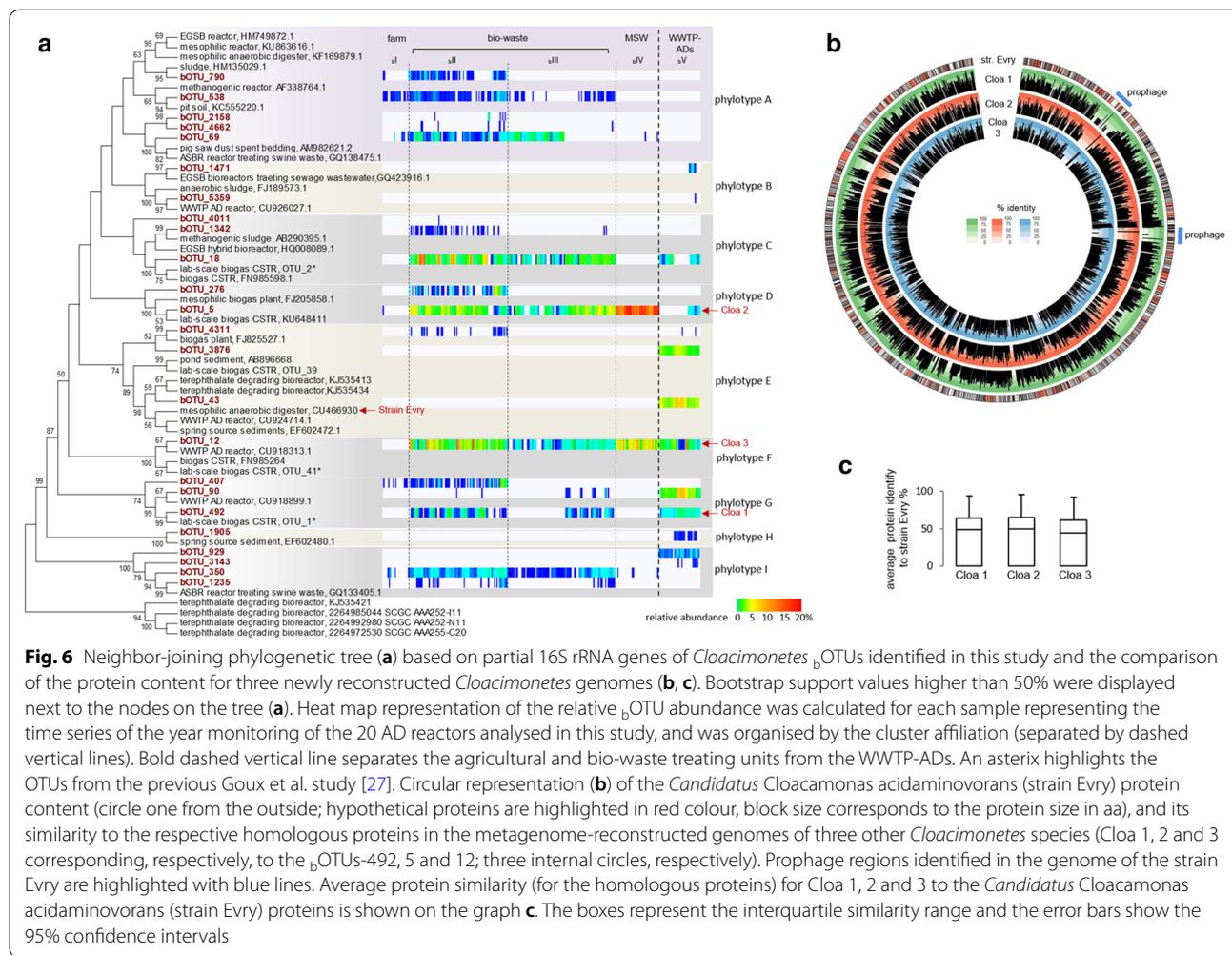


Fig. 6 Neighbor-joining phylogenetic tree **(a)** based on partial 16S rRNA genes of *Cloacimonetes* _bOTUs identified in this study and the comparison of the protein content for three newly reconstructed *Cloacimonetes* genomes **(b, c)**. Bootstrap support values higher than 50% were displayed next to the nodes on the tree **(a)**. Heat map representation of the relative _bOTU abundance was calculated for each sample representing the time series of the year monitoring of the 20 AD reactors analysed in this study, and was organised by the cluster affiliation (separated by dashed vertical lines). Bold dashed vertical line separates the agricultural and bio-waste treating units from the WWTP-ADs. An asterisk highlights the OTUs from the previous Goux et al. study [27]. Circular representation **(b)** of the *Candidatus Cloacimonas acidaminovorans* (strain Evry) protein content (circle one from the outside); hypothetical proteins are highlighted in red colour, block size corresponds to the protein size in aa, and its similarity to the respective homologous proteins in the metagenome-reconstructed genomes of three other *Cloacimonetes* species (Cloa 1, 2 and 3 corresponding, respectively, to the _bOTUs-492, 5 and 12; three internal circles, respectively). Prophage regions identified in the genome of the strain Evry are highlighted with blue lines. Average protein similarity (for the homologous proteins) for Cloa 1, 2 and 3 to the *Candidatus Cloacimonas acidaminovorans* (strain Evry) proteins is shown on the graph **c**. The boxes represent the interquartile similarity range and the error bars show the 95% confidence intervals

agricultural and bio-waste treating units. Phylotypes B, E and H were mainly found in WWTP-ADs. *Cloacimonetes* phylotypes C, D, E, G and I were equally abundant in both anaerobic digestion systems. Such distribution of species could indicate the niche specialisation in the case of some *Cloacimonetes*, and it has not been discussed before in the literature.

Cloacimonetes were first discovered in a municipal WWTP in Evry, where they represented 85% of the generated clone library [49]. Preliminary analyses of their genomic content suggesting their involvement in cellulose degradation were further confirmed experimentally [50]. Moreover, reconstructed genomes of *Cloacimonetes* candidates contain most of the genes encoding enzymes involved in propionate oxidation via the methylmalonyl-CoA pathway, suggesting that they might be involved in a syntrophic oxidation of propionate [51]. Sequence-based homology search revealed that the partially reconstructed genomes corresponding to the *Cloacimonetes* _bOTU_5 (Cloa 1), _bOTU-12 (Cloa 2) and _bOTU-492

(Cloa 3) had 65.5% ± 3.9 (average protein identity below 50%) of their proteins similar to the *Candidatus Cloacimonas acidaminovorans* (strain Evry; Fig. 6b, c). This could indicate similar metabolisms. However, most (around 70%) of the non-homologous proteins (on average representing one-third of the protein pool) were assigned as hypothetical, thus functionally unknown. The putative prophages were also missing. Due to the lack of isolates, it is, therefore, impossible based on a simple genome comparison, to discuss the differences between the metabolic capabilities of the proposed *Cloacimonetes* phylotypes. However, according to our previous bio-augmentation success with *Cloacimonetes*-enriched sludge [52], we think *Cloacimonetes* might be one of the key-stone species that according to the concept of the centres of concentrated activity [42] might take a central role in the anaerobic digestion process.

Candidate WSA2 archaeon dominated the WWTP ADs representing 38.4% ± 10.3 of archaeal population in these reactors. Recent phylogenetic analyses indicate

that this group is different from other *Euryarchaeota*, and reconstructed genome suggests WSA2 to be the first methanogen restricted to methanogenesis through methylated thiol reduction [53]. Using acetate, malonate and especially propionate as carbon source, it could be regarded as a beneficial microbe to the stability of the AD process (i.e. increased structural stability of WWTP-core archaea towards general AD-core might be related to the presence of these archaea), as propionate tends to accumulate under sub-optimal conditions in AD reactors [27]. Moreover, using the methylated thiol reduction, WSA2 may potentially compete with sulfate reducers thus controlling the amount of the produced H_2S in the reactor.

Core microbiomes in a global context

Taking into consideration the different assay designs (e.g. different primer pair used, handling of sampled material, thresholds used during the bioinformatics analyses, etc.) undertaken in different studies it is difficult to directly compare the results from the literature and verify the universality of the proposed core community concept in different digestion systems worldwide. Even if 16S rRNA do not reassemble well in metagenomes, as preliminary analysis, we blasted the 16S rRNA sequences generated in this study over the 8000 assembled genomes recently reconstructed from 1500 public metagenomes [54]. As a result, 123 OTUs matched with at least 97% identity to 103 unique genomes (Additional file 8: Table S7). Most of them corresponded to the transient OTUs, and only eight genomes represented AD-core OTUs and 31 WWTP-ADs core bacteria. Therefore, further sequencing efforts and cultivation studies are necessary, especially with regards to the microbes present in agricultural and bio-waste treating AD reactors (largely underrepresented in public databases) to better uncover the real metabolic capabilities of the AD-core microbes.

Still, some of the dominant AD-core bacteria described here (e.g. β OTU-9 and β OTU-4) were previously discovered in other ADs, including thermophilic anaerobic digesters in Japan [55] or CSTR bioreactors in Germany [56], suggesting their worldwide distribution. In the case of the agricultural biogas plants in Germany, more than 75% of reactors utilise maize as a major feeding substrate [57]. Therefore, we could expect the bacterial cores in these reactors to be less diverse than our AD-core, as they have specialised in processing mainly this explicit biomass. In Luxembourg and Belgium, most of the reactors receive a very complex and highly diversified along the year feed (e.g. MSW), therefore, we believe that the diverse microbial communities that have established might be considered more representative of the different anaerobic digestion systems worldwide.

Conclusions

Globally, the growing appreciation of understanding microbial ecology to improve the stability and the efficiency of full-scale bioenergy production systems encourages researchers to study the AD microbial communities in a profound manner. In this study, by monitoring 20 AD reactors over a period of 1 year, we demonstrated that the diversity of microbial communities was high and varied significantly between differentially operated AD reactors, mainly due to the presence of highly diverse and dynamic transient β OTUs. However, using the core community concept, we have shown that the ubiquitously distributed OTUs (roughly 2.5 and 12.4% of bacterial and archaeal OTUs, respectively), accounted for over 70% of the community abundance in agricultural and bio-waste treating units. Regardless the reactor operational conditions and feedstock composition (i.e. agricultural waste, manure, MSW, etc.), core communities were very stable over the year in terms of community membership and species dynamics. Proper installation design (e.g. U-7) has proved that even the microbial communities fed with highly diversified MSW might be very stable, opening the doors for an increased MSW valorisation in AD reactors. WWTP-ADs were shown to operate with distinct microbiomes where 12.4% of bacterial and 41% of archaeal OTUs were classified as a WWTP-ADs core. Based on our results, we hypothesize that the core microbial community of an AD process might have a more reduced complexity in terms of species richness than previously expected. Indeed, according to the “insurance hypothesis” [31], the greater the diversity of the species responses, the lower the species richness at which the ecosystem becomes redundant. In this sense, future research should prioritise the characterisation of the largely unknown and often representing candidate phyla AD-core and WWTP-ADs core microbes to unravel the complexity of their metabolic pathways and their real potential in a methanogenic reactor [58]. Characterisation of these putative key process microbes and a better comprehension of the interactions between them will help to establish the relation between species diversity (composition) and community functionality and dynamics. This would allow us to understand whether highly diverse communities with the portfolio of serial pathways (continuous competition between highly diverse species can; however, destabilise the community structure) or less diverse microbiomes with complementarity of functions (lower species competition should assure better structural stability of the ecosystem) are better adapted to a high and stable industrial biogas production.

Methods

AD samples

AD reactors located at the wastewater treatment plants correspond to Petange (P), Schifflange (S) and Bettembourg (BT) units. U-7 is a MSW-valorising plant located in Mondercange. The remaining studied reactors are private installations. After thorough reactor mixing and careful purging of the sampling port, a representative sludge volume of 2 L was subsampled from each reactor and aliquots were frozen on site in liquid nitrogen. Back to the laboratory, frozen aliquots were stored at $-80\text{ }^{\circ}\text{C}$ prior to the analysis. TS (24 h at $105\text{ }^{\circ}\text{C}$) and VS (6 h at $550\text{ }^{\circ}\text{C}$) were determined in the remaining sludge samples according to the 4630 VDI norm [59]. The pH was measured with a pH 196 Microprocessor pH meter connected to a SenTix[®] 21 pH electrode (WTW, Weilheim, Germany). TIC and $\text{NH}_4\text{-N}$ were measured in conformity with the manufacturer's protocol, using the BiogasPro system (RIMU, Königsbrunn, Germany). The concentration of VFAs was measured following an ether extraction and using a gas chromatography (Agilent technologies, Santa Clara, USA) equipped with a Varian CP-FFAP column and a flame ionization detector (FID). The migration was done with helium (He) as a carrier gas. The total VFAs concentrations (mg kg^{-1}) were expressed as the sum of the individual VFA concentrations measured for acetate, propionate, isobutyrate, butyrate, isovalerate, valerate and caproate. Genomic DNA was extracted from the sludge quantity of 200–500 mg, with the PowerSoil DNA Isolation kit (MoBio) according to the manufacturer's protocol.

Selection of amplification primers and sequencing assay design

Based on the *in silico* results [60], we pre-selected two primer pairs to separately target bacterial and archaeal 16S rRNA gene (Table 2). The Illumina platform-compatible dual-index paired-end approach was designed analogously to the approaches designed elsewhere ([61],

Illumina 16S Metagenomic Sequencing Library Preparation Note Part # 15044223 Rev.A), with additional modifications. Each forward and reverse primer consisted of an Illumina-compatible forward/reverse primer overhang attached to the 5' end. Additionally, a heterogeneity spacer of four degenerate nucleotides (Ns) was added to the forward primer, between the primer overhang and the locus-specific sequence. The Illumina barcodes and sequencing adapters were added during the subsequent cycle-limited amplification step using Nextera XT Index kit (Illumina).

Preparation of small rRNA amplicon libraries and sequencing on Illumina platform

The small rRNA amplicons were generated using the Q5 Hot Start High-Fidelity DNA Polymerase (New England Biolabs Inc.) in triplicate 25- μL reactions, using 1 ng of template DNA, 0.4 μM of each primer and BSA (Sigma) at the final concentration of 1 mg/mL. The reaction mixtures were subjected to an initial denaturation at $98\text{ }^{\circ}\text{C}$ for 30 s, followed by 22–26 cycles at $98\text{ }^{\circ}\text{C}$ for 10 s, annealing for 30 s and elongation at $72\text{ }^{\circ}\text{C}$ for 30 s, and a final elongation at $72\text{ }^{\circ}\text{C}$ for 2 min (Additional file 3: Table S2). Following the amplification, the triplicate amplifications were visualized on 3% agarose gels, pooled together and purified with the AMPure magnetic beads (Agencourt) and quantified with the Qubit dsDNA HS assay kit (Life Technologies). The concentration of the amplicons was adjusted to 1 ng/ μL and 1 μL of each library was used as a template in a second PCR reaction where the Nextera XT barcodes and the Illumina adapters necessary for hybridization to the flow cell were added. PCR amplification was performed with the same enzyme and cycling conditions were as described above, with the total number of cycles reduced to eight and an annealing temperature of $55\text{ }^{\circ}\text{C}$. The resulting amplicons were purified with the AMPure magnetic beads (Agencourt), quantified and pooled in equimolar concentrations (between 96 and 384 samples were mixed and sequenced in a single sequencing run). The detailed protocols are

Table 2 Primers pairs used in this study to specifically amplify 16S rRNA genes from bacteria and archaea

Domain	Primer name	Sequence 5'→3'	Variable region	Average amplicon size (bp)	In silico specificity ^a	Ref.
Bacteria	S-D-Bact-0909-a-S-18	ACTCAAAGGAATWGACGG	V6–V8	484	0–78.8%	[60]
	S*-Univ-1392-a-A-15	ACGGGCGGTGTGTRC			1–89.6%	
Archaea	S-D-Arch-0519-a-S-15	CAGCMGCCCGGTAA	V4–V6	526	0–79.3%	[60]
	S-D-Arch-1041-a-A-18	GGCCATGCACCWCCTCTC			1–93.7%	

^a The *in silico* specificity against the rRNA gene sequences deposited in SILVA database v132; the first number refers to the number of mismatches, while the percentage refers to the proportion of taxa that was targeted specifically

provided as supporting information (Additional file 3). The final concentration of the library pool was determined with a KAPA SYBR FAST Universal qPCR Kit (Kapa Biosystems), according to the manufacturer's instructions. Libraries were mixed with Illumina-generated PhiX control library, denatured with fresh NaOH, diluted to a final concentration of 8 pM, and sequenced with the MiSeq Reagent Kit V3-600 cycles (LIST, Luxembourg), using the sequencing primers for reads 1–4 provided with the kit.

Sequencing data analysis and statistics

The obtained small rRNA amplicon sequence reads were de-multiplexed, quality trimmed and OTUs were constructed with UPARSE version v7.0.1090 [62], and taxonomically annotated using the non-redundant SILVA SSU ribosomal database v.128 [63]. Final nucleotide sequences (OTUs) were deposited in the GenBank database under the accession numbers KU648407–KU659020.

All diversity analyses were performed on per sample subsampled shared files (normalize.shared command) using mothur v.1.34.4 or later [64]. Community richness and diversity were calculated using sobs and invsimpson calculators, respectively. Community evenness was measured via the invsimpson even coefficient on a normalised scale from 0 (uneven; one or several dominant OTUs and many singlets) to 1 (perfectly even). The stability of community structure and membership was calculated using the Bray–Curtis dissimilarity and Jaccard indices. Average species abundance was calculated excluding the samples where OTU was not represented by at least one read. Statistical significance of data was calculated using either Wilcoxon signed rank test or Kruskal–Wallis test, or ANOSIM, and the differences were considered statistically significant at p value ≤ 0.05 . NMDS analyses and heat maps were done using R version 3.4.0 [65]. Partial 16S rRNA gene sequences for the *Cloacimonetes* β -OTUs and the close relatives were aligned using MAFFT algorithm [66] and the neighbor-joining phylogenetic tree was constructed with MEGA6 [67].

The CCA analyses were performed with the CANOCO software version 4.5 [68] and the significance test was done using Monte Carlo permutation (500 times). Spearman correlations were calculated using R version 3.4.0 [65], and the calculated correlations were considered statistically significant at p value ≤ 0.05 . Three metagenome reconstructed genomes of novel *Cloacimonetes* (Cloa 1, 2 and 3)

were taken from Broeksema et al. [28] and correspond to bins 1, 31 and 13, respectively, in that study. The similarity of their protein content towards the sequenced genome of the *Candidatus Cloacamonas acidaminovorans* [24] was visualized with Circos [69].

Additional files

Additional file 1: Figure S1. Characterisation of the studied AD reactors. Typical design of (A) PFR-type (U-7) and (B) CSTR-type reactors (all other units) of the studied biogas plant installations.

Additional file 2: Table S1. Characterisation of the studied AD reactors. Detailed (per sample) characterisation of the studied AD reactors.

Additional file 3: Figure S2. The 16S rRNA gene amplicon assay design. Design of amplification primers used in this study; first- and second-level barcoding strategy, and **Table S2.** The 16S rRNA gene amplification conditions (1st PCR).

Additional file 4: Table S3. The 16S rRNA gene amplicon assay validation. Letter codes for the additional libraries prepared for six samples S1–S6 used to validate the second-level barcoding strategy; **Table S4.** Different preparations of libraries for six samples S1–S6 used to validate the second-level barcoding strategy; **Figure S3.** NMDS of pairwise Bray–Curtis distance comparisons calculated for separate and mix amplicon library preparations (intra-DNA extraction comparison) and for the two DNA extractions tested (inter-DNA extraction comparison) for six selected samples (S1–S6); **Figure S4.** Taxonomical distribution of sequencing reads for separate and mix amplicon library preparations for the two different DNA extractions (Ext1 and Ext2) tested for six selected samples (S1–S6); **Figure S5.** Median richness (sobs index) and diversity (invsimpson index) metrics calculated for separate and mix amplicon library preparations and for the two different DNA extractions (Ext1 and Ext2) tested for six selected samples (S1–S6).

Additional file 5: Table S5. The 16S rRNA gene amplicon sequencing results of 1-year monthly time series of 20 AD reactors located in Luxembourg and Belgium. Relative abundance (%), normalised number of reads) and taxonomic affiliation of the 16S rRNA amplicon sequencing OTUs for bacteria and archaea.

Additional file 6: Figure S6. The 16S rRNA gene amplicon sequencing results of 1-year monthly time series of 20 AD reactors located in Luxembourg and Belgium. Rarefaction curves based on the calculated species richness (Sobs) for bacteria (A) and archaea (B). Rank abundance curve of bacterial OTUs (C); **Figure S7.** Median bacterial richness (A, Sobs), diversity (B, invsimpson) and evenness (C, invsimpson even) per reactor during one-year monitoring survey; **Figure S8.** Cluster-specific average Bray–Curtis dissimilarity in community structures and Jaccard (Jclass) dissimilarity in community membership for bacteria (A, B) and archaea (C, D) for each of the community clusters; **Figure S9.** Median archaeal richness (A, Sobs), diversity (B, invsimpson) and evenness (C, invsimpson even) per reactor during 1-year monitoring survey; **Figure S10.** Calculated average Bray–Curtis dissimilarity in “AD-core” and “AD-transient” community structures and Jaccard dissimilarity in “AD-core” community and “AD-transient” membership for bacteria (A, B) and archaea (C, D) for each of the studied reactors.

Additional file 7: Table S6. The 16S rRNA gene amplicon sequencing results of 1-year monthly time series of 20 AD reactors located in Luxembourg and Belgium. Spearman correlation matrix between operational data and dominant microbial groups/ OTUs.

Additional file 8: Table S7. The 16S rRNA gene amplicon sequencing results of 1-year monthly time series of 20 AD reactors located in Luxembourg and Belgium. Blast search results for β -OTUs analysed in this study against the 8000 metagenome-reconstructed genomes from the study by Parks et al. [54].

Authors' contributions

MC and XG contributed to data acquisition and interpretation and drafted the manuscript. MC and MF performed lab work. EELM and PW contributed to data interpretation and writing of the manuscript. MC, EELM, PW and PD conceived and directed the research. All authors read and approved the final manuscript.

Author details

¹ Environmental Research and Innovation Department, Luxembourg Institute of Science and Technology, 41 rue du Brill, 4422 Belvaux, Luxembourg. ² Eco-Systems Biology Group, Luxembourg Centre for Systems Biomedicine, University of Luxembourg, 7 avenue des Hauts-Fourneaux, 4362 Esch-sur-Alzette, Luxembourg. ³ Department of Microbiology, Genomics and the Environment, Université de Strasbourg, CNRS, GMGM, UMR 7156, Strasbourg, France.

Acknowledgements

We would like to thank Bénédicte De Vos and Anaïs Noo for their technical support. The authors are most thankful to the following people, including members of the Biogas Vereenegung a.s.b.l. and the partnership of the Ecobiogaz project of the INTERREG IV A program, for providing year round access to the biogas plants and for their help in sampling the reactors for sludge: François Delion (MINETT-KOMPOST, Mondercange), Marta Porcel and Nico Godart (NATURGAS KIELEN, Kehlen), Marc and Jean Marc Mangen (BIOMAN, Flaxweiler), Jean and Nicolas Kessler (Ferme du Faascht, Attert), Nico Rasqui (Biogas Biekerich, Hovelange), Gaston Welbes (CAPRISO, Canach), Severin Boonen (Elvange), André Dettaille, Jean-Jacques Schmit and Régine Reinert (STEP, Bettembourg), Gerry Bissen and Gianni Di Pentima (SIVEC, Schifflange), Piero Daresta, Roland Troes, and Raymond Erpelding (SIACH, Pétange).

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

Final nucleotide sequences (OTUs) were deposited in the GenBank database under the accession numbers KU648407–KU659020.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

Funding

This work was supported by the Luxembourg National Research Fund (FNR CORE 2011 project GASPOP; C11/SR/1280949: Influence of the Reactor Design and the Operational Parameters on the Dynamics of the Microbial Consortia Involved in the Biomethanation Process). Additionally, it was co-supported by an ATTRACT programme grant (ATTRACT/A09/03) and a European Union Joint Programming in Neurodegenerative Diseases grant (INTER/JPNP/12/01) both to PW., and an Aide à la Formation Recherche (AFR) grant to E.E.L.M. (PRD-2011-1/SR), all funded by the Luxembourg National Research Fund.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 28 February 2018 Accepted: 6 July 2018

Published online: 19 July 2018

References

- Verstraete W, Wittebolle L, Heylen K, Vanparys B, de Vos P, van de Wiele T, Boon N. Microbial resource management: the road to go for environmental biotechnology. *Eng Life Sci*. 2007;7(2):117–26.
- Read S, Marzorati M, Guimarães BCM, Boon N. Microbial resource management revisited: successful parameters and new concepts. *Appl Microbiol Biotechnol*. 2011;90(3):861–71.
- Sundberg C, Al-Soud WA, Larsson M, Alm E, Yekta SS, Svensson BH, Sorensen SJ, Karlsson A. 454 pyrosequencing analyses of bacterial and archaeal richness in 21 full-scale biogas digesters. *FEMS Microbiol Ecol*. 2013;85(3):612–26.
- Verstraete W. The manufacturing microbe. *Microb Biotechnol*. 2015;8(1):36–7.
- Tiwary A, Williams ID, Pant DC, Kishore VVN. Emerging perspectives on environmental burden minimisation initiatives from anaerobic digestion technologies for community scale biomass valorisation. *Renew Sustain Energy Rev*. 2015;42:883–901.
- Achinas S, Achinas V, Euverink GJW. A technological overview of biogas production from biowaste. *Engineering*. 2017;3(3):299–307.
- Nobu MK, Narihiro T, Rinke C, Kamagata Y, Tringe SG, Woyke T, Liu WT. Microbial dark matter ecogenomics reveals complex synergistic networks in a methanogenic bioreactor. *ISME J*. 2015;9(8):1710–22.
- Kirkegaard RH, McIlroy SJ, Kristensen JM, Nierychlo M, Karst SM, Dueholm MS, Albertsen M, Nielsen PH. Identifying the abundant and active microorganisms common to full scale anaerobic digesters. *bioRxiv*. 2017. <https://doi.org/10.1101/104620>.
- De Vrieze J, Saunders AM, He Y, Fang J, Nielsen PH, Verstraete W, Boon N. Ammonia and temperature determine potential clustering in the anaerobic digestion microbiome. *Water Res*. 2015;75:312–23.
- Abendroth C, Vilanova C, Gunther T, Luschign O, Porcar M. Eubacteria and archaea communities in seven mesophile anaerobic digester plants in Germany. *Biotechnol Biofuels*. 2015;8:87.
- Werner JJ, Knights D, Garcia ML, Scalfone NB, Smith S, Yarasheski K, Cummings TA, Beers AR, Knight R, Angenent LT. Bacterial community structures are unique and resilient in full-scale bioenergy systems. *Proc Natl Acad Sci USA*. 2011;108(10):4158–63.
- Fontana A, Patrone V, Puglisi E, Morelli L, Bassi D, Garuti M, Rossi L, Cappa F. Effects of geographic area, feedstock, temperature, and operating time on microbial communities of six full-scale biogas plants. *Bioresour Technol*. 2016;218:980–90.
- Luo G, Fotidis IA, Angelidaki I. Comparative analysis of taxonomic, functional, and metabolic patterns of microbiomes from 14 full-scale biogas reactors by metagenomic sequencing and radioisotopic analysis. *Biotechnol Biofuels*. 2016;9:51.
- Campanaro S, Treu L, Kougias PG, Luo G, Angelidaki I. Metagenomic binning reveals the functional roles of core abundant microorganisms in twelve full-scale biogas plants. *Water Res*. 2018;140:123–34.
- Koch C, Müller S, Harms H, Harnisch F. Microbiomes in bioenergy production: from analysis to management. *Curr Opin Biotechnol*. 2014;27:65–72.
- Vanwonterghem I, Jensen PD, Ho DP, Batstone DJ, Tyson GW. Linking microbial community structure, interactions and function in anaerobic digesters using new molecular techniques. *Curr Opin Biotechnol*. 2014;27:55–64.
- Carballa M, Regueiro L, Lema JM. Microbial management of anaerobic digestion: exploiting the microbiome-functionality nexus. *Curr Opin Biotechnol*. 2015;33:103–11.
- Wittebolle L, Marzorati M, Clement L, Balloi A, Daffonchio D, Heylen K, De Vos P, Verstraete W, Boon N. Initial community evenness favours functionality under selective stress. *Nature*. 2009;458:623–6.
- Konopka A, Lindemann S, Fredrickson J. Dynamics in microbial communities: unraveling mechanisms to identify principles. *ISME J*. 2015;9(7):1488–95.
- Thingstad TF, Lignell R. Theoretical models for the control of bacterial growth rate, abundance, diversity and carbon demand. *Aquat Microb Ecol*. 1997;13:19–27.
- Saunders AM, Albertsen M, Vollertsen J, Nielsen PH. The activated sludge ecosystem contains a core community of abundant organisms. *ISME J*. 2016;10(1):11–20.
- Goux X, Calusinska M, Fossépré M, Benizri E, Delfosse P. Start-up phase of an anaerobic full-scale farm reactor—appearance of mesophilic anaerobic conditions and establishment of the methanogenic microbial community. *Bioresour Technol*. 2016;212:217–26.
- Nelson MC, Morrison M, Yu Z. A meta-analysis of the microbial diversity observed in anaerobic digesters. *Bioresour Technol*. 2011;102(4):3730–9.
- Pelletier E, Kreimeyer A, Bocs S, Rouy Z, Gyapay G, Chouari R, Rivière D, Ganesan A, Daegelen P, Sghir A, et al. Candidatus Cloacamonas acidaminovorans: genome sequence reconstruction provides a first glimpse of a new bacterial division. *J Bacteriol*. 2008;190(7):2572–9.

25. Lucas R, Kuchenbuch A, Fetzter I, Harms H, Kleinstueber S. Long-term monitoring reveals stable and remarkably similar microbial communities in parallel full-scale biogas reactors digesting energy crops. *FEMS Microbiol Ecol*. 2015;91:3.
26. Sun L, Liu T, Müller B, Schnürer A. The microbial community structure in industrial biogas plants influences the degradation rate of straw and cellulose in batch tests. *Biotechnol Biofuels*. 2016;9:128.
27. Goux X, Calusinska M, Lemaigre S, Marynowska M, Klocke M, Udelhoven T, Benizri E, Delfosse P. Microbial community dynamics in replicate anaerobic digesters exposed sequentially to increasing organic loading rate, acidosis, and process recovery. *Biotechnol Biofuels*. 2015;8(1):1–18.
28. Broeksema B, Calusinska M, McGee F, Winter K, Bongiovanni F, Goux X, Wilmes P, Delfosse P, Ghoniem M. ICoVeR—an interactive visualization tool for verification and refinement of metagenomic bins. *BMC Bioinform*. 2017;18:233.
29. Rago L, Zecchin S, Marzatori S, Goglio A, Cavalca L, Cristiani P, Schievano A. A study of microbial communities on terracotta separator and on biocathode of air breathing microbial fuel cells. *Bioelectrochem*. 2018;120:18–26.
30. Rotaru A-E, Shrestha PM, Liu F, Markovaite B, Chen S, Nevin KP, Lovley DR. Direct interspecies electron transfer between *Geobacter metallireducens* and *Methanosarcina barkeri*. *Appl Environ Microbiol*. 2014;80(15):4599–605.
31. Yachi S, Loreau M. Biodiversity and ecosystem productivity in a fluctuating environment: the insurance hypothesis. *Proc Natl Acad Sci USA*. 1999;96(4):1463–8.
32. Pimm SL. The complexity and stability of ecosystems. *Nature*. 1984;307:321–6.
33. Xiang X, Wang R, Wang H, Gong L, Man B, Xu Y. Distribution of Bathyarchaeota communities across different terrestrial settings and their potential ecological functions. *Sci Rep*. 2017;7:45028.
34. Evans PN, Parks DH, Chadwick GL, Robbins SJ, Orphan VJ, Golding SD, Tyson GW. Methane metabolism in the archaeal phylum Bathyarchaeota revealed by genome-centric metagenomics. *Science*. 2015;350(6259):434–8.
35. Kirkegaard RH, McIlroy SJ, Kristensen JM, Nierychlo M, Karst SM, Dueholm MS, Albertsen M, Nielsen PH. The impact of immigration on microbial community composition in full-scale anaerobic digesters. *Sci Rep*. 2017;7:9343.
36. Rivière D, Desvignes V, Pelletier E, Chaussonnerie S, Guermazi S, Weissenbach J, Li T, Camacho P, Sghir A. Towards the definition of a core of microorganisms involved in anaerobic digestion of sludge. *ISME J*. 2009;3:700–14.
37. Shade A, Handelsman J. Beyond the Venn diagram: the hunt for a core microbiome. *Environ Microbiol*. 2012;14(1):4–12.
38. Mei R, Narihiro T, Nobu MK, Kuroda K, Liu WT. Evaluating digestion efficiency in full-scale anaerobic digesters by identifying active microbial populations through the lens of microbial activity. *Sci Rep*. 2016;6:34090.
39. Verstraete W, Mertens B. The key role of soil microbes. In: Doelman P, Eijsackers HJP, editors. *Vital soil, function, value and properties developments in soil science*, vol. 29. Amsterdam: Elsevier; 2004. p. 127–51.
40. Wang X, Wen X, Yan H, Ding K, Zhao F, Hu M. Bacterial community dynamics in a functionally stable pilot-scale wastewater treatment plant. *Bioresour Technol*. 2011;102:2352–7.
41. Wittebolle L, Vervaeren H, Verstraete W, Boon N. Quantifying community dynamics of nitrifiers in functionally stable reactors. *Appl Environ Microbiol*. 2007;74(1):286–93.
42. De Vrieze J, Verstraete W. Perspectives for microbial community composition in anaerobic digestion: from abundance and activity to connectivity. *Environ Microbiol*. 2016;18(9):2797–809.
43. Campbell BJ, Kirchman DL. Bacterial diversity, community structure and potential growth rates along an estuarine salinity gradient. *ISME J*. 2013;7(1):210–20.
44. Westerholm M, Moestedt J, Schnürer A. Biogas production through syntrophic acetate oxidation and deliberate operating strategies for improved digester performance. *Appl Energy*. 2016;179:124–35.
45. Dridi B, Fardeau ML, Bernard O, Raoult D, Drancourt M. *Methanomassili-coccus luminyensis* gen. nov., sp. nov., a methanogenic archaeon isolated from human faeces. *Int J Syst Evol Microbiol*. 2012;62:1902–7.
46. Lee SH, Kang HJ, Lee YH, Lee TJ, Han K, Choi Y, Park HD. Monitoring bacterial community structure and variability in time scale in full-scale anaerobic digesters. *J Environ Monit*. 2012;14(7):1893–905.
47. Muller EEL, Pinel N, Laczny CC, Hoopmann MR, Narayanasamy S, Lebrun LA, Roume H, Lin J, May P, Hicks ND, et al. Community-integrated omics links dominance of a microbial generalist to fine-tuned resource usage. *Nat Commun*. 2014;5:5603.
48. Fang HHP. Microbial distribution in UASB granules and its resulting effects. *Water Sci Technol*. 2000;42(12):201–8.
49. Chouari R, Le Paslier D, Dauga C, Daegelen P, Weissenbach J, Sghir A. Novel major bacterial candidate division within a municipal anaerobic sludge digester. *Appl Environ Microbiol*. 2005;71:2145–53.
50. Limam RD, Chouari R, Mazéas L, Wu TD, Li T, Grossin-Debattista J, Guerquin-Kern JL, Saidi M, Landoulsi A, Sghir A, et al. Members of the uncultured bacterial candidate division WWE1 are implicated in anaerobic digestion of cellulose. *MicrobiologyOpen*. 2014;9(3):157–67.
51. Kuever J, Visser M, Loeffler C, Boll M, Worm P, Sousa DZ, Plugge CM, Schaap PJ, Muyzer G, Pereira IA, et al. Genome analysis of *Desulfotomaculum gibsoniae* strain Groll(T) a highly versatile Gram-positive sulfate-reducing bacterium. *Stand Genomic Sci*. 2014;9(3):821–39.
52. Calusinska M, Goux X, Lemaigre S, Delfosse P. Biological agent for enrichment of anaerobic digestion reactors, method of preparing such biological agent and bio-augmentation process with said agent. LU93402. Luxembourg; 2017.
53. Nobu MK, Narihiro T, Kuroda K, Mei R, Liu WT. Chasing the elusive Euryarchaeota class WSA2: genomes reveal a uniquely fastidious methyl-reducing methanogen. *ISME J*. 2016;10(10):2478–87.
54. Parks DH, Rinke C, Chuvochina M, Chaumeil P-A, Woodcroft BJ, Evans PN, Hugenholtz P, Tyson GW. Recovery of nearly 8,000 metagenome-assembled genomes substantially expands the tree of life. *Nat Microbiol*. 2017;2(11):1533–42.
55. Tang YQ, Ji P, Hayashi J, Koike Y, Wu XL, Kida K. Characteristic microbial community of a dry thermophilic methanogenic digester: its long-term stability and change with feeding. *Appl Microbiol Biotechnol*. 2011;91(5):1447–61.
56. Klocke M, Mähner P, Mundt K, Souidi K, Linke B. Microbial community analysis of a biogas-producing completely stirred tank reactor fed continuously with fodder beet silage as mono-substrate. *Syst Appl Microbiol*. 2007;30(2):139–51.
57. Leubhn M, Munk B, Effenberger M. Agricultural biogas production in Germany—from practice to microbiology basics. *Energy Sustain Soc*. 2014;4:10.
58. Campanaro S, Treu L, Kougias PG, Francisci D, Valle G, Angelidaki I. Metagenomic analysis and functional characterization of the biogas microbiome using high throughput shotgun sequencing and a novel binning strategy. *Biotechnol Biofuels*. 2016;9(1):1–17.
59. Anonymous. *Vergärung organischer stoffe (german guideline: fermentation of organic materials—characterisation of the substrate, sampling, collection of material data, fermentation tests)*. Berlin: Beuth Verlag GmbH; 2006.
60. Klindworth A, Pruesse E, Schweer T, Peplies J, Quast C, Horn M, Glöckner FO. Evaluation of general 16S ribosomal RNA gene PCR primers for classical and next-generation sequencing-based diversity studies. *Nucleic Acids Res*. 2013;41(1):e1.
61. Kozich JJ, Westcott SL, Baxter NT, Highlander SK, Schloss PD. Development of a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence data on the MiSeq Illumina sequencing platform. *Appl Environ Microbiol*. 2013;79(17):5112–20.
62. Edgar RC. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat Methods*. 2013;10(10):996–8.
63. Pruesse E, Peplies J, Glöckner FO. SINA: accurate high-throughput multiple sequence alignment of ribosomal RNA genes. *Bioinformatics*. 2012;28(14):1823–9.
64. Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson C, et al. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl Environ Microbiol*. 2009;75(23):7537–41.
65. R Core Team. *R: a language and environment for statistical computing*. Vienna: R foundation for statistical computing; 2013.

66. Katoh K, Rozewicki J, Yamada KD. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief Bioinform*. 2017. <https://doi.org/10.1093/bib/bbx108>.
67. Tamura K, Stecher G, Peterson D, Filipowski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol*. 2013;30(12):2725–9.
68. Ter Braak C. The analysis of vegetation–environment relationships by canonical correspondence analysis. *Vegetatio*. 1987;69(1–3):69–77.
69. Krzywinski M, Schein J, Birol I, Connors J, Gascoyne R, Horsman D, Jones SJ, Marra MA. Circos: an information aesthetic for comparative genomics. *Genome Res*. 2009;19(9):1639–45.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

