

Nonequilibrium Thermodynamics of Chemical Reaction Networks: Wisdom from Stochastic Thermodynamics

Riccardo Rao and Massimiliano Esposito

*Complex Systems and Statistical Mechanics, Physics and Materials Science Research Unit,
University of Luxembourg, L-1511 Luxembourg, Luxembourg*

(Received 23 February 2016; revised manuscript received 26 October 2016; published 22 December 2016)

We build a rigorous nonequilibrium thermodynamic description for open chemical reaction networks of elementary reactions. Their dynamics is described by deterministic rate equations with mass action kinetics. Our most general framework considers open networks driven by time-dependent chemostats. The energy and entropy balances are established and a nonequilibrium Gibbs free energy is introduced. The difference between this latter and its equilibrium form represents the minimal work done by the chemostats to bring the network to its nonequilibrium state. It is minimized in nondriven detailed-balanced networks (i.e., networks that relax to equilibrium states) and has an interesting information-theoretic interpretation. We further show that the entropy production of complex-balanced networks (i.e., networks that relax to special kinds of nonequilibrium steady states) splits into two non-negative contributions: one characterizing the dissipation of the nonequilibrium steady state and the other the transients due to relaxation and driving. Our theory lays the path to study time-dependent energy and information transduction in biochemical networks.

DOI: [10.1103/PhysRevX.6.041064](https://doi.org/10.1103/PhysRevX.6.041064)

Subject Areas: Chemical Physics, Nonlinear Dynamics, Statistical Physics

I. INTRODUCTION

Thermodynamics of chemical reactions has a long history. The second half of the 19th century witnessed the dawn of the modern studies on thermodynamics of chemical mixtures. It is indeed at that time that Gibbs introduced the concept of *chemical potential* and used it to define the thermodynamic potentials of noninteracting mixtures [1]. Several decades later, this enabled de Donder to approach the study of chemical reacting mixtures from a thermodynamic standpoint. He proposed the concept of *affinity* to characterize the chemical force irreversibly driving chemical reactions and related it to the thermodynamic properties of mixtures established by Gibbs [2]. Prigogine, who perpetuated the Brussels School founded by de Donder, introduced the assumption of *local equilibrium* to describe irreversible processes in terms of equilibrium quantities [3,4]. In doing so, he pioneered the connections between thermodynamics and kinetics of chemical reacting mixtures [5].

During the second half of the 20th century, part of the attention moved to systems with small particle numbers which are ill described by “deterministic” rate equations. The Brussels School, as well as other groups, produced various studies on the nonequilibrium thermodynamics of

chemical systems [6–11] using a stochastic description based on the (chemical) master equation [12,13]. These studies played an important role during the first decade of the 21st century for the development of *stochastic thermodynamics*, a theory that systematically establishes a nonequilibrium thermodynamic description for systems obeying stochastic dynamics [14–17], including chemical reaction networks (CRNs) [18–22].

Another significant part of the attention moved to the thermodynamic description of biochemical reactions in terms of deterministic rate equations [23,24]. This is not so surprising since living systems are the paramount example of nonequilibrium processes and they are powered by chemical reactions. The fact that metabolic processes involve thousands of coupled reactions also emphasized the importance of a network description [25–27]. While complex dynamical behaviors such as oscillations were analyzed in small CRNs [28,29], most studies on large biochemical networks focused on the steady-state dynamics. Very few studies considered the thermodynamic properties of CRNs [30–33]. One of the first nonequilibrium thermodynamic descriptions of open biochemical networks was proposed in Ref. [34]. However, it did not take advantage of chemical reaction network theory, which connects the network topology to its dynamical behavior and which was extensively studied by mathematicians during the 1970s [35–37] (this theory was also later extended to stochastic dynamics [38–41]). As far as we know, the first and single study that related the nonequilibrium thermodynamics of CRNs to their topology is Ref. [22], still restricting itself to steady states.

Published by the American Physical Society under the terms of the Creative Commons Attribution 3.0 License. Further distribution of this work must maintain attribution to the author(s) and the published article's title, journal citation, and DOI.

In this paper, we consider the most general setting for the study of CRNs, namely, open networks driven by chemostatted concentrations that may change over time. To the best of our knowledge, this was never considered before. In this way, steady-state properties as well as transient ones are captured. Hence, in the same way that stochastic thermodynamics is built on top of stochastic dynamics, we systematically build a nonequilibrium thermodynamic description of CRNs on top of deterministic chemical rate equations. In doing so, we establish the energy and entropy balance and introduce the nonequilibrium entropy of the CRN as well as its nonequilibrium Gibbs free energy. We show the latter to bear an information-theoretical interpretation similar to that of stochastic thermodynamics [42–45] and to be related to the dynamical potentials derived by mathematicians. We also show the relation between the minimal chemical work necessary to manipulate the CRNs far from equilibrium and the nonequilibrium Gibbs free energy. Our theory embeds both the Prigoginian approach to thermodynamics of irreversible processes [5] and the thermodynamics of biochemical reactions [23]. Making full use of the mathematical chemical reaction network theory, we further analyze the thermodynamic behavior of two important classes of CRNs: detailed-balanced networks and complex-balanced networks. In the absence of time-dependent driving, the former converges to thermodynamic equilibrium by minimizing their nonequilibrium Gibbs free energy. In contrast, the latter converges to a specific class of nonequilibrium steady states and always allows for an adiabatic–nonadiabatic separation of their entropy production, which is analogous to that found in stochastic thermodynamics [46–50]. Recently, a result similar to the latter was independently found in Ref. [51].

A. Outline and notation

The paper is organized as follows. After introducing the necessary concepts in chemical kinetics and chemical reaction network theory, Sec. II, the nonequilibrium thermodynamic description is established in Sec. III. As in stochastic thermodynamics, we build it on top of the dynamics and formulate the entropy and energy balance, Secs. III D and III E. Chemical work and nonequilibrium Gibbs free energy are also defined, and the information-theoretic content of the latter is discussed. The special properties of detailed-balanced and of complex-balanced networks are considered in Secs. V and IV, respectively. Conclusions and perspectives are drawn in Sec. VI, while some technical derivations are detailed in the appendixes.

We now proceed by fixing the notation. We consider a system composed of reacting *chemical species* X_σ , each of which is identified by an index $\sigma \in \mathcal{S}$, where \mathcal{S} is the set of all indices or species. The species populations change due to *elementary reactions*, i.e., all reacting species and reactions must be resolved (none can be

hidden), and all reactions must be *reversible*, i.e., each forward reaction $+\rho$ has a corresponding backward reaction $-\rho$. Each pair of forward-backward reactions is a *reaction pathway* denoted by $\rho \in \mathcal{R}$. The orientation of the set of reaction pathways \mathcal{R} is arbitrary. Hence, a generic CRN is represented as

$$\sum_{\sigma} \nabla_{+\rho}^{\sigma} X_{\sigma} \xrightleftharpoons[k^{-\rho}]{k^{+\rho}} \sum_{\sigma} \nabla_{-\rho}^{\sigma} X_{\sigma}. \quad (1)$$

The constants $k^{+\rho}$ ($k^{-\rho}$) are the *rate constants* of the forward (backward) reactions. The stoichiometric coefficients $-\nabla_{+\rho}^{\sigma}$ and $\nabla_{-\rho}^{\sigma}$ identify the number of molecules of X_{σ} involved in each forward reaction $+\rho$ (the stoichiometric coefficients of the backward reactions have opposite signs). Once stacked into two non-negative matrices, $\nabla_{+} = \{\nabla_{+\rho}^{\sigma}\}$ and $\nabla_{-} = \{\nabla_{-\rho}^{\sigma}\}$, they define the integer-valued stoichiometric matrix

$$\nabla \equiv \nabla_{-} - \nabla_{+}. \quad (2)$$

The reason for the choice of the symbol “ ∇ ” will become clear later.

Example 1.—The stoichiometric matrix of the CRN depicted in Fig. 1 is

$$\nabla = \begin{pmatrix} -1 & 0 \\ 2 & 0 \\ 1 & -1 \\ 0 & -1 \\ 0 & 1 \end{pmatrix}. \quad (3)$$

□

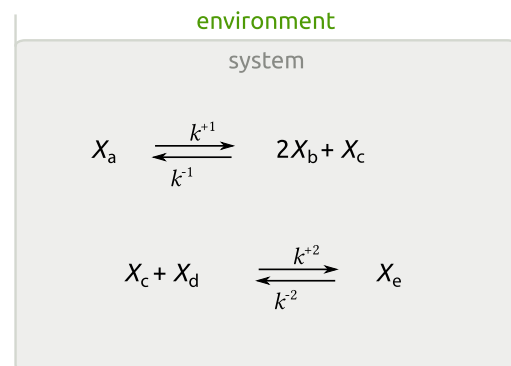


FIG. 1. Representation of a closed CRN. The chemical species are $\{X_a, \dots, X_e\}$. The two reaction pathways are labeled by 1 and 2. The nonzero stoichiometric coefficients are $-\nabla_{+1}^a = -1$, $\nabla_{-1}^b = 2$, and $\nabla_{-1}^c = 1$ for the first forward reaction and $-\nabla_{+2}^c = -1$, $-\nabla_{+2}^d = -1$, and $\nabla_{-2}^e = 1$ for the second one. Since the network is closed, no chemical species is exchanged with the environment.

Physical quantities associated with species and reactions are represented in upper-lower indices vectorial notation. Upper and lower indexed quantities have the same physical values, e.g., $Z^i = Z_i$, $\forall i$. We use the Einstein summation notation: repeated upper-lower indices implies the summation over all the allowed values for those indices—e.g., $\sigma \in \mathcal{S}$ for species and $\rho \in \mathcal{R}$ for reactions. Given two arbitrary vectorial quantities $\mathbf{a} = \{a^i\}$ and $\mathbf{b} = \{b^i\}$, the following notation is used:

$$a^{ib_i} \equiv \prod_i a^{ib_i}.$$

Finally, given the matrix C , whose elements are $\{C_{ij}^i\}$, the elements of the transposed matrix C^T are $\{C_i^j\}$.

The time derivative of a physical quantity A is denoted by $d_t A$, its steady state value by an overbar \bar{A} , and its equilibrium value by A_{eq} or A^{eq} . We reserve the overdot \dot{A} to denote the rate of change of quantities that are *not* exact time derivatives.

II. DYNAMICS OF CRNS

In this section, we formulate the mathematical description of CRNs [52,53] in a suitable way for a thermodynamic analysis. We introduce closed and open CRNs and show how to drive these latter in a time-dependent way. We then define conservation laws and cycles and review the dynamical properties of two important classes of CRNs: detailed-balanced networks and complex-balanced networks.

We consider a chemical system in which the reacting species $\{X_\sigma\}$ are part of a homogeneous and ideal dilute solution: the reactions proceed slowly compared to diffusion and the solvent is much more abundant than the reacting species. Temperature T and pressure p are kept constant. Since the volume of the solution V is overwhelmingly dominated by the solvent, it is assumed constant. The species abundances are large enough so that the molecule's discreteness can be neglected. Thus, at any time t , the system state is well described by the molar concentration distribution $\{Z^\sigma \equiv N^\sigma/V\}$, where N^σ is the molarity of the species X_σ .

The reaction kinetics is controlled by the reaction rate functions $J^{\pm\rho}(\{Z^\sigma\})$, which measure the rate of occurrence of reactions and satisfy the mass action kinetics [52,54,55]:

$$J^{\pm\rho} \equiv J^{\pm\rho}(\{Z^\sigma\}) = k^{\pm\rho} Z^{\sigma \nabla_\sigma^{\pm\rho}}. \quad (4)$$

The net concentration current along a reaction pathway ρ is thus given by

$$J^\rho \equiv J^{+\rho} - J^{-\rho} = k^{+\rho} Z^{\sigma \nabla_\sigma^{+\rho}} - k^{-\rho} Z^{\sigma \nabla_\sigma^{-\rho}}. \quad (5)$$

Example 2.—For the CRN in Fig. 1 the currents are

$$\begin{aligned} J^1 &= k^{+1} Z^a - k^{-1} (Z^b)^2 Z^c, \\ J^2 &= k^{+2} Z^c Z^d - k^{-2} Z^e. \end{aligned} \quad (6)$$

□

A. Closed CRNs

A closed CRN does not exchange any chemical species with the environment. Hence, the species concentrations vary solely due to chemical reactions and satisfy the rate equations

$$d_t Z^\sigma = \nabla_\rho^\sigma J^\rho, \quad \forall \sigma \in \mathcal{S}. \quad (7)$$

Since rate equations are nonlinear, complex dynamical behaviors may emerge [29]. The fact that the rate equations [Eq. (7)] can be thought of as a continuity equation for the concentration, where the stoichiometric matrix ∇ [Eq. (2)] acts as a discrete differential operator, explains the choice of the symbol “ ∇ ” for the stoichiometric matrix [56].

B. Driven CRNs

In open CRNs, matter is exchanged with the environment via reservoirs that control the concentrations of some specific species, Fig. 2. These externally controlled species are said to be *chemostatted*, while the reservoirs controlling them are called chemostats. The chemostating procedure may mimic various types of controls by the environment. For instance, a direct control could be implemented via external reactions (not belonging to the CRN) or via abundant species whose concentrations are negligibly affected by the CRN reactions within relevant time scales. An indirect control may be achieved via semipermeable membranes or by controlled injection of chemicals in continuous stirred-tank reactors.

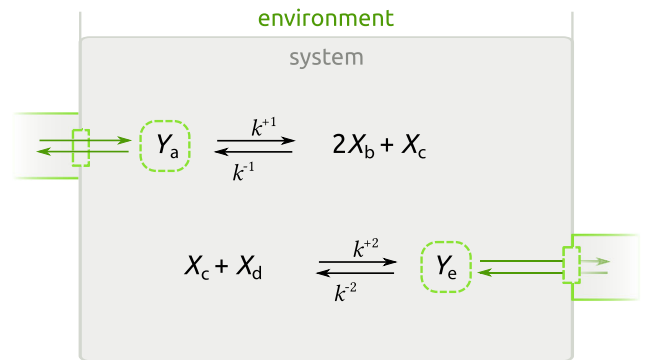


FIG. 2. Representation of an open CRN. With respect to the CRN in Fig. 1, the species X_a and X_e are chemostatted, hence, represented as Y_a and Y_e . The green boxes on the sides represent the reservoirs of chemostatted species.

Among the chemical species, the chemostatted ones are denoted by the indices $\sigma_y \in \mathcal{S}_y$, and the internal ones by $\sigma_x \in \mathcal{S}_x$ ($\mathcal{S} \equiv \mathcal{S}_x \cup \mathcal{S}_y$). Also, the part of the stoichiometric matrix related to the internal (chemostatted) species is denoted by $\nabla^X = \{\nabla_\rho^{\sigma_x}\}$ ($\nabla^Y = \{\nabla_\rho^{\sigma_y}\}$).

Example 3.—When chemostating the CRN in Fig. 1 as in Fig. 2 the stoichiometric matrix Eq. (3) splits into

$$\nabla^X = \begin{pmatrix} 2 & 0 \\ 1 & -1 \\ 0 & -1 \end{pmatrix}, \quad \nabla^Y = \begin{pmatrix} -1 & 0 \\ 0 & 1 \end{pmatrix}. \quad (8)$$

□

In nondriven open CRNs, the chemostatted species have constant concentrations, i.e., $\{d_t Z^{\sigma_y} = 0\}$. In driven open CRNs, the chemostatted concentrations change over time according to some time-dependent protocol $\pi(t)$: $\{Z^{\sigma_y} \equiv Z^{\sigma_y}(\pi(t))\}$. The changes of the internal species are solely due to reactions and satisfy the rate equations

$$d_t Z^{\sigma_x} = \nabla_\rho^{\sigma_x} J^\rho, \quad \forall \sigma_x \in \mathcal{S}_x. \quad (9)$$

Instead, the changes of chemostatted species $\{d_t Z^{\sigma_y}\}$ are not only given by the species formation rates $\{\nabla_\rho^{\sigma_y} J^\rho\}$ but must in addition contain the external currents $\{I^{\sigma_y}\}$, which quantify the rate at which chemostatted species enter into the CRN (negative if chemostatted species leave the CRN),

$$d_t Z^{\sigma_y} = \nabla_\rho^{\sigma_y} J^\rho + I^{\sigma_y}, \quad \forall \sigma_y \in \mathcal{S}_y. \quad (10)$$

This latter equation is not a differential equation since the chemostatted concentrations $\{Z^{\sigma_y}\}$ are not dynamical variables. It shows that the external control of the chemostatted concentration is not necessarily direct, via the chemostatted concentrations, but can also be indirectly controlled via the external currents. We note that Eq. (10) is the dynamical expression of the decomposition of changes of species populations in internal-external introduced by de Donder (see Secs. 4.1 and 15.2 of Ref. [57]).

A steady-state distribution $\{\bar{Z}^{\sigma_x}\}$, if it exists, must satisfy

$$\nabla_\rho^{\sigma_x} \bar{J}^\rho = 0, \quad \forall \sigma_x \in \mathcal{S}_x, \quad (11a)$$

$$\nabla_\rho^{\sigma_y} \bar{J}^\rho + \bar{I}^{\sigma_y} = 0, \quad \forall \sigma_y \in \mathcal{S}_y, \quad (11b)$$

for given chemostatted concentrations $\{Z^{\sigma_y}\}$.

C. Conservation laws

In a closed CRN, a conservation law $\ell = \{\ell_\sigma\}$ is a left null eigenvector of the stoichiometric matrix ∇ [23,25]:

$$\ell_\sigma \nabla_\rho^\sigma = 0, \quad \forall \rho \in \mathcal{R}. \quad (12)$$

Conservation laws identify conserved quantities $L \equiv \ell_\sigma Z^\sigma$, called components [23,25], which satisfy

$$d_t L = \ell_\sigma d_t Z^\sigma = 0. \quad (13)$$

We denote a set of independent conservation laws of the closed network by $\{\ell^\lambda\}$ and the corresponding components by $\{L^\lambda \equiv \ell^\lambda_\sigma Z^\sigma\}$. The choice of this set is not unique, and different choices have different physical meanings. This set is never empty since the total mass is always conserved. Physically, conservation laws are often related to parts of molecules, called moieties [58], which are exchanged between different species and/or subject to isomerization (see Example 4).

In an open CRN, since only $\{Z^{\sigma_x}\}$ are dynamical variables, the conservation laws become the left null eigenvectors of the stoichiometric matrix of the internal species ∇^X . Stated differently, when starting from the closed CRN, the chemostating procedure may break a subset of the conservation laws of the closed network $\{\ell^\lambda\}$ [56]. For example, when the first chemostat is introduced the total mass conservation law is always broken. Within the set $\{\ell^\lambda\}$, we label the broken ones by λ_b and the unbroken ones by λ_u . The broken conservation laws are characterized by

$$\underbrace{\ell_{\sigma_x}^{\lambda_b} \nabla_\rho^{\sigma_x}}_{\neq 0} + \ell_{\sigma_y}^{\lambda_b} \nabla_\rho^{\sigma_y} = 0, \quad \forall \rho \in \mathcal{R}, \quad (14)$$

where the first term is nonvanishing for at least one $\rho \in \mathcal{R}$. The broken components $\{L^{\lambda_b} \equiv \ell_{\sigma}^{\lambda_b} Z^\sigma\}$ are no longer constant over time. On the other hand, the unbroken conservation laws are characterized by

$$\underbrace{\ell_{\sigma_x}^{\lambda_u} \nabla_\rho^{\sigma_x}}_{=0} + \ell_{\sigma_y}^{\lambda_u} \nabla_\rho^{\sigma_y} = 0, \quad \forall \rho \in \mathcal{R}, \quad (15)$$

where the first term vanishes for all $\rho \in \mathcal{R}$. Therefore, the unbroken components $\{L^{\lambda_u} \equiv \ell_{\sigma}^{\lambda_u} Z^\sigma\}$ remain constant over time. Without loss of generality, we choose the set $\{\ell^\lambda\}$ such that the entries related to the chemostatted species vanish, $\ell_{\sigma_y}^{\lambda_u} = 0, \forall \lambda_u, \sigma_y$.

Example 4.—For the CRN in Fig. 1, an independent set of conservation laws is

$$\begin{aligned} \ell^1 &= (2 \quad 1 \quad 0 \quad 0 \quad 0), \\ \ell^2 &= (0 \quad 0 \quad 0 \quad 1 \quad 1), \\ \ell^3 &= (0 \quad \frac{1}{2} \quad -1 \quad 1 \quad 0). \end{aligned} \quad (16)$$

When chemostating as in Fig. 2, the first two conservation laws break while the last one remains unbroken. We also

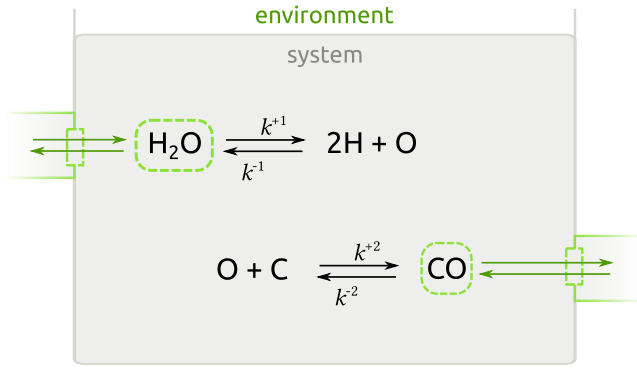


FIG. 3. Specific implementation of the CRN in Fig. 2.

note that this set is chosen so that the unbroken conservation law satisfies $\ell_a^3 = \ell_c^3 = 0$. When considering the specific implementation in Fig. 3 of the CRN in Fig. 2, we see that the first two conservation laws in Eq. (16) represent the conservation of the concentrations of the moiety H and C, respectively. Instead, the third conservation law in Eq. (16) does not have a straightforward interpretation. It is related to the fact that when the species H or C are produced, also O must be produced and vice versa. \square

D. Detailed-balanced networks

A steady state [Eq. (11)] is said to be an equilibrium state $\{Z_{\text{eq}}^\sigma\}$ if it satisfies the detailed-balance property [[57], § 9.4], i.e., all concentration currents Eq. (5) vanish:

$$J_{\text{eq}}^\rho \equiv J^\rho(\{Z_{\text{eq}}^\sigma\}) = 0, \quad \forall \rho \in \mathcal{R}. \quad (17)$$

For open networks, this means that the external currents, Eq. (11b), must also vanish, $\{I_{\text{eq}}^{\sigma_y} = 0\}$. By virtue of mass action kinetics, Eq. (4), the detailed-balance property Eq. (17) can be rewritten as

$$\frac{k^{+\rho}}{k^{-\rho}} = Z_{\text{eq}}^\sigma \nabla_\sigma^\rho, \quad \forall \rho \in \mathcal{R}. \quad (18)$$

A CRN is said to be detailed balanced if, for given kinetics $\{k^{\pm\rho}\}$ and chemostating $\{Z^{\sigma_y}\}$, its dynamics exhibits an equilibrium steady state, Eq. (17). For each set of unbroken components $\{L^{\lambda_u}\}$ —which are given by the initial condition and constrain the space where the dynamics dwells—the equilibrium distribution is globally stable [59]. Equivalently, detailed-balanced networks always relax to an equilibrium state, which for a given kinetics and chemostating is unique and depends on the unbroken components only; see also Sec. V.

Closed CRNs must be detailed balanced. This statement can be seen as the zeroth law for CRNs. Consequently, rather than considering Eq. (18) as a property of the equilibrium distribution, we impose it as a property that

the rate constants must satisfy and call it a local detailed-balance property. It is a universal property of elementary reactions that holds regardless of the network state. Indeed, while the equilibrium distribution depends on the components, the rhs of Eq. (18) does not. This point will become explicit after introducing the thermodynamic structure, Eq. (88) in Sec. V. The local detailed-balance property will be rewritten in a thermodynamic form in Sec. III B, Eq. (50).

In open nondriven CRNs, the chemostating procedure may prevent the system from reaching an equilibrium state. To express this scenario algebraically, we now introduce the concepts of emergent cycle and cycle affinity.

A cycle $\tilde{\mathbf{c}} = \{\tilde{c}^\rho\}$ is a right null eigenvector of the stoichiometric matrix [56], namely,

$$\nabla_\rho^\sigma \tilde{c}^\rho = 0, \quad \forall \sigma \in \mathcal{S}. \quad (19)$$

Since ∇ is integer valued, $\tilde{\mathbf{c}}$ can always be rescaled to only contain integer coefficients. In this representation, its entries denote the number of times each reaction occurs (negative signs identify reactions occurring in backward direction) along a transformation that overall leaves the concentration distributions $\{Z^\sigma\}$ unchanged; see Example 5. We denote by $\{\tilde{\mathbf{c}}_\alpha\}$ a set of linearly independent cycles. An emergent cycle $\mathbf{c} = \{c^\rho\}$ is defined algebraically as [56]

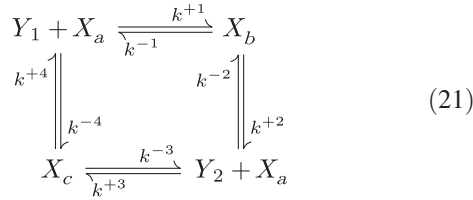
$$\begin{aligned} \nabla_\rho^{\sigma_x} c^\rho &= 0, \quad \forall \sigma_x \in \mathcal{S}_x, \\ \nabla_\rho^{\sigma_y} c^\rho &\neq 0, \quad \text{for at least one } \sigma_y \in \mathcal{S}_y. \end{aligned} \quad (20)$$

In its integer-valued representation, the entries of \mathbf{c} denote the number of times each reaction occurs along a transformation that overall leaves the concentrations of the internal species $\{Z^{\sigma_x}\}$ unchanged while changing the concentrations of the chemostatted species by an amount $\nabla_\rho^{\sigma_y} c^\rho$. These latter are, however, immediately restored to their prior values due to the injection of $-\nabla_\rho^{\sigma_y} c^\rho$ molecules of X_{σ_y} performed by the chemostats. Emergent cycles are, thus, pathways transferring chemicals across chemostats while leaving the internal state of the CRN unchanged. We denote by $\{\mathbf{c}_\varepsilon\}$ a set of linearly independent emergent cycles.

When chemostating an initially closed CRN, for each species that is chemostatted, either a conservation law breaks—as mentioned in Sec. II C—or an independent emergent cycle arises [56]. This follows from the rank nullity theorem for the stoichiometric matrices ∇ and ∇^x , which ensures that the number of chemostatted species $|\mathcal{S}_y|$ equals the number of broken conservation laws $|\lambda_b|$ plus the number of independent emergent cycles $|\varepsilon|$: $|\mathcal{S}_y| = |\lambda_b| + |\varepsilon|$. Importantly, the rise of emergent cycles is a topological feature: it depends on the species that are chemostatted, but not on the chemostatted concentrations.

We also note that emergent cycles are modeled as “flux modes” in the context of metabolic networks [60–62].

Example 5.—To illustrate the concepts of cycles and emergent cycles, we use the following CRN [56]:



whose Y_1 and Y_2 species are chemostatted. The stoichiometric matrix decomposes as

$$\begin{aligned}
 \nabla^X &= \begin{pmatrix} -1 & 1 & -1 & 1 \\ 1 & -1 & 0 & 0 \\ 0 & 0 & 1 & -1 \end{pmatrix}, \\
 \nabla^Y &= \begin{pmatrix} -1 & 0 & 0 & 1 \\ 0 & 1 & -1 & 0 \end{pmatrix}.
 \end{aligned} \quad (22)$$

The set of linearly independent cycles, Eq. (19), consists of only one cycle, which can be written as

$$\tilde{\mathbf{c}} = (1 \ 1 \ 1 \ 1)^T. \quad (23)$$

As the CRN is chemostatted, one linearly independent emergent cycle Eq. (20) arises:

$$\mathbf{c} = (1 \ 1 \ -1 \ -1)^T. \quad (24)$$

We now see that if each reaction occurs a number of times given by the entry of the cycle Eq. (23), the CRN goes back to the initial state, no matter which one it is. On the other hand, when the emergent cycle Eq. (24) is performed, the state of the internal species does not change, while two molecules of Y_1 are annihilated and two of Y_2 are created. However, since the chemostats restore their initial values, the overall result of \mathbf{c} is to transfer two Y_1 , transformed in Y_2 , from the first to the second chemostat.

The closed version of this CRN has two independent conservation laws,

$$\begin{aligned}
 \ell^1 &= (0 \ 1 \ 1 \ 1 \ 1), \\
 \ell^2 &= (1 \ 1 \ 1 \ 0 \ 0),
 \end{aligned} \quad (25)$$

the first of which, ℓ^1 , is broken following the chemostatting of any of the two species Y_1 or Y_2 . The other chemostatted species, instead, gives rise to the emergent cycle Eq. (24), so that the relationship $|\mathcal{S}_y| = |\lambda_b| + |\varepsilon|$ is satisfied. \square

Any cycle $\tilde{\mathbf{c}}_\alpha$ and emergent cycle \mathbf{c}_ε bears a cycle affinity [56],

$$\tilde{\mathcal{A}}_\alpha = \tilde{c}_\alpha^\rho RT \ln \frac{J_{+\rho}}{J_{-\rho}}, \quad (26)$$

$$\mathcal{A}_\varepsilon = c_\varepsilon^\rho RT \ln \frac{J_{+\rho}}{J_{-\rho}}. \quad (27)$$

From the definition of cycle, Eq. (19), and current, Eq. (5), and the local detailed balance, Eq. (18), it follows that the cycle affinities along the cycles Eq. (19) vanish, $\{\tilde{\mathcal{A}}_\alpha = 0\}$, and that the cycle affinities along the emergent cycles depend on only the chemostatted concentrations

$$\mathcal{A}_\varepsilon = c_\varepsilon^\rho RT \ln \frac{k_{+\rho}}{k_{-\rho}} Z_{\sigma_y}^{-\nabla_y^\rho}. \quad (28)$$

Since emergent cycles are pathways connecting different chemostats, the emergent affinities quantify the chemical forces acting along the cycles. This point will become clearer later, when the thermodynamic expressions of the emergent cycle affinities $\{\mathcal{A}_\varepsilon\}$ is given, Eq. (49).

A CRN is detailed balanced if and only if all the emergent cycle affinities $\{\mathcal{A}_\varepsilon\}$ vanish. This condition is equivalent to the Wegscheider condition [59]. This happens when the chemostatted concentrations fit an equilibrium distribution. As a special case, unconditionally detailed-balanced networks are open CRNs with no emergent cycle. Therefore, they are detailed balanced for any choice of the chemostatted concentrations. Consequently, even when a time-dependent driving acts on such a CRN and prevents it from reaching an equilibrium state, a well-defined equilibrium state exists at any time: the equilibrium state to which the CRN would relax if the time-dependent driving were stopped.

Example 6.—Any CRN with one chemostatted species only ($|\mathcal{S}_y| = 1$) is unconditionally detailed balanced. Indeed, as mentioned in Sec. II C, the first chemostatted species always breaks the mass conservation law $|\lambda_b| = 1$, and, thus, no emergent cycle arises, $|\varepsilon| = |\mathcal{S}_y| - |\lambda_b| = 0$.

The open CRN in Fig. 2 is an example of an unconditionally detailed-balanced network with two chemostatted species, since the chemostatting breaks two conservation laws; see Example 4. Indeed, a nonequilibrium steady state would require a continuous injection of Y_a and ejection of Y_e (or vice versa). But this would necessarily result in a continuous production of X_b and consumption of X_d , which is in contradiction with the steady-state assumption. \square

Finally, a tacit assumption in the above discussion is that the network involves a finite number of species and reactions, i.e., the CRN is finite dimensional. Infinite-dimensional CRNs can exhibit long-time behaviors

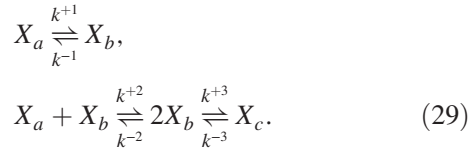
different from equilibrium even in the absence of emergent cycles [63].

E. Complex-balanced networks

To discuss complex-balanced networks and complex-balanced distributions, we first introduce the notion of complex in open CRNs.

A *complex* is a group of species that combines in a reaction as products or as reactants. Each side of Eq. (1) defines a complex, but different reactions might involve the same complex. We label complexes by $\gamma \in \mathcal{C}$, where \mathcal{C} is the set of complexes.

Example 7.—Let us consider the following CRN [64]



The set of complexes is $\mathcal{C} = \{X_a, X_b, X_a + X_b, 2X_b, X_c\}$, and the complex $2X_b$ is involved in both the second and third reaction. \square

The notion of complex allows us to decompose the stoichiometric matrix ∇ as

$$\nabla_\rho^\sigma = \Gamma_\gamma^\sigma \partial_\rho^\gamma. \quad (30)$$

We call $\Gamma = \{\Gamma_\gamma^\sigma\}$ the composition matrix [35,37]. Its entries Γ_γ^σ are the stoichiometric number of species X_σ in the complex γ . The composition matrix encodes the structure of each complex in terms of species; see Example 8. The matrix $\partial = \{\partial_\rho^\gamma\}$ denotes the incidence matrix of the CRN, whose entries are given by

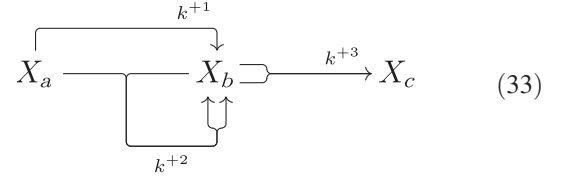
$$\partial_\rho^\gamma = \begin{cases} 1 & \text{if } \gamma \text{ is the product complex of } +\rho \\ -1 & \text{if } \gamma \text{ is the reactant complex of } +\rho \\ 0 & \text{otherwise.} \end{cases} \quad (31)$$

The incidence matrix encodes the structure of the network at the level of complexes, i.e., how complexes are connected by reactions. If we think of complexes as network nodes, the incidence matrix associates an edge to each reaction pathway and the resulting topological structure is a reaction graph, see, e.g., Fig. 1 and Eqs. (21) and (29). The stoichiometric matrix instead encodes the structure of the network at the level of species. If we think of species as the network nodes, the stoichiometric matrix does not define a graph, since reaction connects more than a pair of species, in general. The structure originating is rather a hypergraph [56,65] or, equivalently, a Petri net [66,67].

Example 8.—The composition matrix and the incidence matrix of the CRN in Eq. (29) are

$$\Gamma = \begin{pmatrix} 1 & 0 & 1 & 0 & 0 \\ 0 & 1 & 1 & 2 & 0 \\ 0 & 0 & 0 & 0 & 1 \end{pmatrix}, \quad \partial = \begin{pmatrix} -1 & 0 & 0 \\ 1 & 0 & 0 \\ 0 & -1 & 0 \\ 0 & 1 & -1 \\ 0 & 0 & 1 \end{pmatrix}, \quad (32)$$

where the complexes are ordered as in Example 7. The corresponding reaction hypergraph is



where only the forward reactions are depicted. \square

In an open CRN, we regroup all complexes $\gamma \in \mathcal{C}$ of the closed CRN that have the same stoichiometry for the internal species (i.e., all complexes with the same internal part of the composition matrix Γ_γ^X regardless of the chemostatted part Γ_γ^Y) in sets denoted by \mathcal{C}_j , for $j = 1, 2, \dots$. Complexes of the closed network made solely of chemostatted species in the open CRN are all regrouped in the same complex \mathcal{C}_0 . This allows one to decompose the internal species stoichiometric matrix as

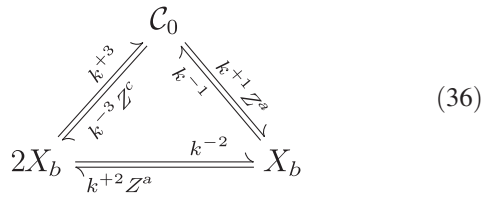
$$\nabla_\rho^{\sigma_x} = \Gamma_j^{\sigma_x} \partial_\rho^j. \quad (34)$$

where $\{\Gamma_j^{\sigma_x} \equiv \Gamma_\gamma^{\sigma_x}, \text{ for } \gamma \in \mathcal{C}_j\}$ are the entries of the composition matrix corresponding to the internal species, and $\{\partial_\rho^j \equiv \sum_{\gamma \in \mathcal{C}_j} \partial_\rho^\gamma\}$ are the entries of the incidence matrix describing the network of regrouped complexes. This regrouping corresponds to the—equivalent—CRN made of only internal species with the effective rate constant $\{k^{\pm\rho} Z^{\sigma_y} \nabla_{\sigma_y}^{\pm\rho}\}$ ruling each reaction.

Example 9.—Let us consider the CRN Eq. (29) where the species X_a and X_c are chemostatted. The five complexes of the closed network, see Example 7, are regrouped as $\mathcal{C}_0 = \{X_a, X_c\}$, $\mathcal{C}_1 = \{X_b, X_b + X_a\}$, and $\mathcal{C}_2 = \{2X_b\}$. In terms of these groups of complexes, the composition matrix and incidence matrix are

$$\Gamma^X = \begin{pmatrix} -1 & 0 & 1 \\ 0 & 1 & 2 \\ 0 & 1 & -1 \end{pmatrix}, \quad \partial^C = \begin{pmatrix} -1 & 0 & 1 \\ 1 & -1 & 0 \\ 0 & 1 & -1 \end{pmatrix}, \quad (35)$$

which corresponds to the effective representation



□

A steady-state distribution $\{\bar{Z}^{\sigma_x}\}$ (11) is said to be complex balanced if the net current flowing in each group of complexes C_j vanishes, i.e., if the currents $\{\bar{J}^\rho\}$ satisfy

$$\partial_\rho^j \bar{J}^\rho \equiv \sum_{\gamma \in C_j} \partial_\rho^\gamma \bar{J}^\rho = 0, \quad \forall j. \quad (37)$$

Complex-balanced steady states are, therefore, a subclass of steady states Eq. (11a) that include equilibrium ones, Eq. (17), as a special case:

$$\underbrace{\Gamma_j^{\sigma_x} \partial_\rho^j \bar{J}^\rho}_{\substack{=0 \text{ iff Detailed-Balanced Steady State} \\ =0 \text{ iff Complex-Balanced Steady State} \\ =0 \text{ for generic steady states}}} = 0. \quad (38)$$

While for generic steady states only the internal species formation rates vanish, for complex-balanced ones the complex formation rates also vanish.

For a fixed kinetics ($\{k^{\pm\rho}\}$) and chemostatting (\mathcal{S}_y and $\{Z^{\sigma_y}\}$), a CRN is complex balanced if its dynamics exhibits a complex-balanced steady state, Eq. (37) [35,36]. The complex-balanced distribution Eq. (37) depends on the unbroken components $\{L^{\lambda_u}\}$, which can be inferred from the initial conditions, and is always globally stable [68]. Hence, complex-balanced networks always relax to a—complex-balanced—steady state. Detailed-balanced networks are a subclass of complex-balanced networks.

Whether or not a CRN is complex balanced depends on the network topology (∇), the kinetics ($\{k^{\pm\rho}\}$), and the chemostatting (\mathcal{S}_y and $\{Z^{\sigma_y}\}$). For any given network topology and set of chemostatted species \mathcal{S}_y , one can always find a set of effective rate constants $\{k^{\pm\rho} Z^{\sigma_y \nabla \sigma_y^{\pm\rho}}\}$ that makes that CRN complex balanced [37]. However, for some CRNs, this set coincides with the one that makes the CRN detailed balanced [69]. A characterization of the set of effective rate constants that make a CRN complex balanced is reported in Refs. [37,69].

Deficiency-zero CRNs are a class of CRNs that are complex balanced irrespective of the effective kinetics $\{k^{\pm\rho} Z^{\sigma_y \nabla \sigma_y^{\pm\rho}}\}$ [35–37]. The network deficiency is a topological property of the CRN, which we briefly discuss in Appendix D; see Refs. [22,52,53] for more details. Consequently, regardless of the way in which a deficiency-zero CRN is driven in time, it will always remain

complex balanced. Throughout this paper, we refer to these CRNs as unconditionally complex balanced, as in the seminal work [35].

Example 10.—The open CRN, Eq. (36), has a single steady state \bar{Z}^b for any given set of rate constants and chemostatted concentrations Z^a and Z^c [64], defined by Eq. (11a):

$$\begin{aligned}
 d_i \bar{Z}^b &= \bar{J}^1 + \bar{J}^2 - 2\bar{J}^3 \\
 &= k^{+1} Z^a - k^{-1} \bar{Z}^b + k^{+2} Z^a \bar{Z}^b - k^{-2} (\bar{Z}^b)^2 \\
 &\quad + 2k^{-3} Z^c - 2k^{+3} (\bar{Z}^b)^2 = 0.
 \end{aligned} \quad (39)$$

If the stronger condition Eq. (37) holds,

$$\begin{aligned}
 \bar{J}^3 - \bar{J}^1 &= 0 \quad (\text{group } C_0), \\
 \bar{J}^1 - \bar{J}^2 &= 0 \quad (\text{group } C_1), \\
 \bar{J}^2 - \bar{J}^3 &= 0 \quad (\text{group } C_2),
 \end{aligned} \quad (40)$$

which is equivalent to

$$\begin{aligned}
 k^{+1} Z^a - k^{-1} \bar{Z}^b &= k^{+2} Z^a \bar{Z}^b - k^{-2} (\bar{Z}^b)^2 \\
 &= k^{+3} (\bar{Z}^b)^2 - k^{-3} Z^c,
 \end{aligned} \quad (41)$$

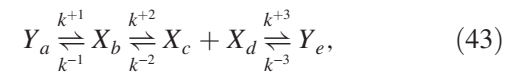
the steady state is complex balanced. Yet, if the steady-state currents are all independently vanishing,

$$\bar{J}^1 = \bar{J}^2 = \bar{J}^3 = 0, \quad (42)$$

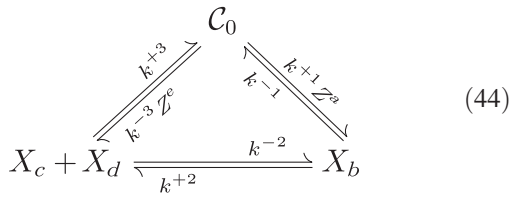
i.e., Eq. (41) is equal to zero, then the steady state is detailed balanced.

When, for simplicity, all rate constants are taken as 1, the complex-balanced set of quadratic equations, Eq. (41), admits a positive solution \bar{Z}^b *only if* $Z^a = 2 - Z^c$ ($0 < Z^c < 2$) or $Z^a = \sqrt{Z^c}$. The former case corresponds to a genuine complex-balanced state, $\bar{Z}^b = 1$ with currents $\bar{J}^1 = \bar{J}^2 = \bar{J}^3 = 1 - Z^c$, while the second corresponds to a detailed-balance state, $\bar{Z}^b = \sqrt{Z^c}$ with vanishing currents. When, for example, $Z^a = 1$ and $Z^c = 4$, neither of the two previous conditions holds: the nonequilibrium steady state is $\bar{Z}^b = \sqrt{3}$ with currents $\bar{J}^1 = 1 - \sqrt{3}$, $\bar{J}^2 = -3 + \sqrt{3}$, and $\bar{J}^3 = -1$. □

Example 11.—Let us now consider the following open CRN [22]:



where the species Y_a and Y_e are chemostatted. Out of the four complexes of the closed network, $\{Y_a, X_b, X_c + X_d, Y_e\}$, two are grouped into $C_0 = \{Y_a, Y_e\}$ and the other two remain $C_1 = \{X_b\}$ and $C_2 = \{X_c + X_d\}$. The effective representation of this open CRN is



This network is deficiency zero and, hence, unconditionally complex balanced [22]. Therefore, given any set of rate constants $k^{\pm 1}$, $k^{\pm 2}$, and $k^{\pm 3}$, and the chemostatted concentrations Z^a and Z^e , the steady state of this CRN is complex balanced, i.e., the steady state always satisfies a set of condition like those in Eq. (40). Indeed, contrary to Example 10, steady-state currents $\{\bar{J}^1, \bar{J}^2, \bar{J}^3\}$ different from each other cannot exist since they would induce a growth or decrease of some concentrations. \square

III. THERMODYNAMICS OF CHEMICAL NETWORKS

Using local equilibrium, here we build the connection between the dynamics and the nonequilibrium thermodynamics for arbitrary CRNs. In the spirit of stochastic thermodynamics, we derive an energy and entropy balance, and express the dissipation of the CRN as the difference between the chemical work done by the reservoirs on the CRN and its change in nonequilibrium free energy. We, finally, discuss the information-theoretical content of the nonequilibrium free energy and its relation to the dynamical potentials used in chemical reaction network theory.

A. Local equilibrium

Since we consider homogeneous reaction mixtures in ideal dilute solutions, the assumption of local equilibrium (Ref. [57], Sec. 15.1, and Ref. [70]) means that the equilibration following any reaction event is much faster than any reaction time scale. Thus, what is assumed is that the nonequilibrium nature of the thermodynamic description is solely due to the reaction mechanisms. If all reactions could be instantaneously shut down, the state of the whole CRN would immediately become an equilibrated ideal mixture of species. As a result, all the intensive thermodynamic variables are well defined and equal everywhere in the system. The temperature T is set by the solvent, which acts as a thermal bath, while the pressure p is set by the environment the solution is exposed to. As a result, each chemical species is characterized by a chemical potential (Ref. [23], Sec. III. 1),

$$\mu_\sigma = \mu_\sigma^\circ + RT \ln \frac{Z_\sigma}{Z_{\text{tot}}}, \quad \forall \sigma \in \mathcal{S}, \quad (45)$$

where R denotes the gas constant and $\{\mu_\sigma^\circ \equiv \mu_\sigma^\circ(T)\}$ are the standard-state chemical potentials, which depend on the temperature and on the nature of the solvent. The total

concentration of the solution is denoted by $Z_{\text{tot}} = \sum_\sigma Z_\sigma + Z^0$, where Z^0 is the concentration of the solvent. We assume for simplicity that the solvent does not react with the solutes. In case it does, our results still hold provided one treats the solvent as a nondriven chemostatted species, as discussed in Appendix A. Since the solvent is much more abundant than the solutes, the total concentration is almost equal to that of the solvent which is a constant, $Z_{\text{tot}} \approx Z_0$. Without loss of generality, the constant term $-RT \ln Z_{\text{tot}} \approx -RT \ln Z^0$ in Eq. (45) is absorbed in the standard-state chemical potentials. Consequently, many equations appear with nonmatching dimensions. We also emphasize that standard-state quantities, denoted with “ \circ ”, are defined as those measured in ideal conditions, at standard pressure ($p^\circ = 100$ kPa) and molar concentration ($Z_\sigma^\circ = 1$ mol/dm³), but not at a standard temperature (Ref. [71], p. 61).

Because of the assumption of local equilibrium and homogeneous reaction mixture, the densities of all extensive thermodynamic quantities are well defined and equal everywhere in the system. With a slight abuse of notation, we use the same symbol and name for densities as for their corresponding extensive quantity. For example, S is the molar entropy divided by the volume of the solution, but we denote it as *entropy*. We apply the same logic to rates of change. For example, we call entropy production rate the molar entropy production density rate.

B. Affinities, emergent affinities, and local detailed balance

The thermodynamic forces driving reactions are given by differences of chemical potential [Eq. (45)],

$$\Delta_r G_\rho \equiv \nabla_\rho^\sigma \mu_\sigma, \quad (46)$$

also called Gibbs free energies of reaction (Ref. [23], Sec. III. 2, and Ref. [57], Sec. IX.3). Since these must all vanish at equilibrium, $\nabla_\rho^\sigma \mu_\sigma^{\text{eq}} = 0$, $\forall \rho$, we have

$$\Delta_r G_\rho = -RT \nabla_\rho^\sigma \ln \frac{Z_\sigma}{Z_\sigma^{\text{eq}}}. \quad (47)$$

The local detailed balance, Eq. (18)] allows us to express these thermodynamic forces in terms of reaction affinities,

$$A_\rho \equiv RT \ln \frac{J_{+\rho}}{J_{-\rho}} = -\Delta_r G_\rho, \quad (48)$$

which quantify the kinetic force acting along each reaction pathway (Ref. [57], Sec. IV.1.3).

The change of Gibbs free energy along emergent cycles,

$$A_\varepsilon = -c_\varepsilon^\rho \Delta_r G_\rho = -c_\varepsilon^\rho \nabla_\rho^\sigma \mu_\sigma, \quad (49)$$

gives the external thermodynamic forces the network is coupled to, as we see in Eq. (61), and thus provides a thermodynamic meaning to the cycle affinities Eq. (28).

Combining the detailed-balance property Eq. (18) and the equilibrium condition on the affinities $A_\rho^{\text{eq}} = 0$ [Eq. (46)], we can relate the Gibbs free energies of reaction to the rate constants

$$\frac{k^{+\rho}}{k^{-\rho}} = \exp \left\{ -\frac{\Delta_r G_\rho^\circ}{RT} \right\}, \quad (50)$$

where $\Delta_r G_\rho^\circ \equiv \nabla_\rho^\sigma \mu_\sigma^\circ$. This relation is the thermodynamic counterpart of the local detailed balance Eq. (18). It plays the same role as in stochastic thermodynamics, namely, connecting the thermodynamic description to the stochastic dynamics. We emphasize that the local detailed-balance property as well as the local equilibrium assumption by no mean imply that the CRN operates close to equilibrium. Their importance is to assign well-defined equilibrium potentials to the states of the CRN, which are then connected by the nonequilibrium mechanisms, i.e., reactions.

C. Enthalpies and entropies of reaction

To identify the heat produced by the CRN, we need to distinguish the enthalpic change produced by each reaction from the entropic one. We consider the decomposition of the standard-state chemical potentials (Ref. [23], Sec. III. 2):

$$\mu_\sigma^\circ = h_\sigma^\circ - T s_\sigma^\circ. \quad (51)$$

The standard enthalpies of formation $\{h_\sigma^\circ\}$ take into account the enthalpic contributions carried by each species (Ref. [23], Sec. III. 2, and Ref. [72], Sec. X.4.2). Enthalpy changes caused by reactions give the enthalpies of reaction (Ref. [23], Sec. III. 2, and Ref. [57], Sec. II. 4),

$$\Delta_r H_\rho = \nabla_\rho^\sigma h_\sigma^\circ, \quad (52)$$

which at constant pressure measure the heat of reaction. This is the content of the Hess law (see, e.g., Ref. [72], Sec. X.4.1). The standard entropies of formation $\{s_\sigma^\circ\}$ take into account the internal entropic contribution carried by each species under standard-state conditions (Ref. [23], Sec. III. 2). Using Eq. (51), the chemical potentials Eq. (45) can be rewritten as

$$\mu_\sigma = h_\sigma^\circ - T \underbrace{(s_\sigma^\circ - R \ln Z_\sigma)}_{\equiv s_\sigma}. \quad (53)$$

The entropies of formation $\{s_\sigma \equiv s_\sigma^\circ - R \ln Z_\sigma\}$ account for the entropic contribution of each species in the CRN (Ref. [23], Sec. III. 2). Entropy changes along reactions are given by

$$\Delta_r S_\rho = \nabla_\rho^\sigma s_\sigma, \quad (54)$$

called entropies of reaction [[23] § 3.2].

D. Entropy balance

1. Entropy production rate

The entropy production rate is a non-negative measure of the break of detailed balance in each chemical reaction. Its typical form is given by (Ref. [8] and Ref. [57], Sec. IX.5)

$$T \dot{S}_i \equiv RT (J_{+\rho} - J_{-\rho}) \ln \frac{J_{+\rho}}{J_{-\rho}} \geq 0, \quad (55)$$

because (1) it is non-negative and vanishes only at equilibrium, i.e., when the detailed-balance property Eq. (17) is satisfied, and (2) it vanishes to first order around equilibrium, thus allowing for quasistatic reversible transformations. Indeed, defining

$$\frac{Z^\sigma - Z_{\text{eq}}^\sigma}{Z_{\text{eq}}^\sigma} = \epsilon^\sigma, \quad |\epsilon^\sigma| \ll 1, \quad \forall \sigma \in \mathcal{S}, \quad (56)$$

we find that

$$\dot{S}_i = E_\sigma^\sigma \epsilon^\sigma \epsilon_\sigma + \mathcal{O}(\epsilon^3), \quad (57)$$

where $E \equiv \{E_\sigma^\sigma\}$ is a positive semidefinite symmetric matrix.

Furthermore, it can be rewritten in a thermodynamically appealing way using [Eq. (48)]

$$T \dot{S}_i = -J^\rho \Delta_r G_\rho. \quad (58)$$

It can be further expressed as the sum of two distinct contributions [56]:

$$T \dot{S}_i = \underbrace{-\mu_{\sigma_x} d_t Z^{\sigma_x}}_{\equiv T \dot{S}_x} - \underbrace{\mu_{\sigma_y} (d_t Z^{\sigma_y} - I^{\sigma_y})}_{\equiv T \dot{S}_y}. \quad (59)$$

The first term is due to changes in the internal species and thus vanishes at steady state. The second term is due to the chemostats. It takes into account both the exchange of chemostatted species and the time-dependent driving of their concentration. If the system reaches a nonequilibrium steady state, the external currents $\{\bar{I}^{\sigma_y}\}$ do not vanish and the entropy production reads

$$T \bar{\dot{S}}_i = \bar{I}^{\sigma_y} \mu_{\sigma_y}. \quad (60)$$

This expression can be rewritten as a bilinear form of emergent cycle affinities $\{\mathcal{A}_\epsilon\}$ Eq. (49) and currents along the emergent cycle $\{\bar{\mathcal{J}}^\epsilon \equiv c_\rho^\epsilon \bar{J}^\rho\}$ [56]

$$T\dot{\bar{S}}_i = \bar{\mathcal{J}}^\varepsilon \mathcal{A}_\varepsilon, \quad (61)$$

which clearly emphasizes the crucial role of emergent cycles in steady-state dissipation.

2. Entropy flow rate

The entropy flow rate measures the reversible entropy changes in the environment due to exchange processes with the system [57]. Using the expressions for the enthalpy of reaction Eq. (52) and entropy of formation Eq. (53), we express the entropy flow rate as

$$T\dot{S}_e \equiv \underbrace{J^\rho \Delta_r H_\rho}_{\equiv \dot{Q}} + I^{\sigma_y} T s_{\sigma_y}. \quad (62)$$

The first contribution is the heat flow rate (positive if heat is absorbed by the system). When divided by temperature, it measures minus the entropy changes in the thermal bath. The second contribution accounts for minus the entropy change in the chemostats.

3. System entropy

The entropy of the ideal dilute solution constituting the CRN is given by (see Appendix A)

$$S = Z^\sigma s_\sigma + RZ^S + S_0. \quad (63)$$

The total concentration term,

$$Z^S \equiv \sum_{\sigma \in \mathcal{S}} Z^\sigma, \quad (64)$$

and the constant S_0 together represent the entropic contribution of the solvent. S_0 may also account for the entropy of chemical species not involved in the reactions. We also prove in Appendix B that the entropy [Eq. (63)] can be obtained as a large particle limit of the stochastic entropy of CRNs.

S would be an equilibrium entropy if the reactions could all be shut down. But in the presence of reactions, it becomes the nonequilibrium entropy of the CRN. Indeed, Using eqs. (53), (58), and (62), we find that its change can be expressed as

$$\begin{aligned} d_t S &= s_\sigma d_t Z^\sigma + Z^\sigma d_t s_\sigma + R d_t Z^S \\ &= s_\sigma d_t Z^\sigma \\ &= J^\rho \Delta_r S_\rho + I^{\sigma_y} s_{\sigma_y} \\ &= \dot{S}_i + \dot{S}_e. \end{aligned} \quad (65)$$

This relation is the nonequilibrium formulation of the second law of thermodynamics for CRNs. It demonstrates that the non-negative entropy production Eq. (55) measures

the entropy changes in the system plus those in the reservoirs (thermal and chemostats) [57].

E. Energy balance

1. First law of thermodynamics

Since the CRN is kept at constant pressure p , its enthalpy

$$H = Z^\sigma h_\sigma^\circ + H_0 \quad (66)$$

is equal to the CRN internal energy, up to a constant. Indeed, the enthalpy H is a density which, when written in terms of the internal energy (density) U , reads $H = U + p$.

Using the rate equations (9) and (10), the enthalpy rate of change can be expressed as the sum of the heat flow rate, defined in Eq. (62), and the enthalpy of formation exchange rate:

$$d_t H = h_\sigma^\circ d_t Z^\sigma = \dot{Q} + I^{\sigma_y} h_{\sigma_y}^\circ. \quad (67)$$

Equivalently, it can be rewritten in terms of the entropy flow rate Eq. (62) as (Ref. [57], Sec. IV.1.2)

$$d_t H = T\dot{S}_e + I^{\sigma_y} \mu_{\sigma_y}. \quad (68)$$

The last term on the rhs of Eq. (68) is the free energy exchanged with the chemostats. It represents the chemical work rate performed by the chemostats on the CRN [21,23]:

$$\dot{W}_c \equiv I^{\sigma_y} \mu_{\sigma_y}. \quad (69)$$

Either Eq. (67) or (68) may be considered as the non-equilibrium formulation of the first law of thermodynamics for CRNs. The former has the advantage to solely focus on energy exchanges. The latter contains entropic contributions but is appealing because it involves the chemical work Eq. (69).

2. Nonequilibrium Gibbs free energy

We are now in the position to introduce the thermodynamic potential regulating CRNs. The Gibbs free energy of ideal dilute solutions reads

$$G \equiv H - TS = Z^\sigma \mu_\sigma - RTZ^S + G_0. \quad (70)$$

As for entropy, the total concentration term $-RTZ^S$ and the constant G_0 represent the contribution of the solvent (see Appendix A). Furthermore, in the presence of reactions, G becomes the nonequilibrium Gibbs free energy of CRNs.

We now show that the nonequilibrium Gibbs free energy of a closed CRN is always greater than or equal to its corresponding equilibrium form. A generic non-equilibrium concentration distribution $\{Z^\sigma\}$ is characterized by the set of components $\{L^\lambda = \ell_\sigma^\lambda Z^\sigma\}$. Let $\{Z_{\text{eq}}^\sigma\}$ be

the corresponding equilibrium distribution defined by the detailed-balance property Eq. (18) and characterized by the same set of components $\{L^\lambda\}$ [a formal expression for the equilibrium distribution is given in Eq. (88)]. At equilibrium, the Gibbs free energy Eq. (70) reads

$$G_{\text{eq}} = Z_{\text{eq}}^\sigma \mu_\sigma^{\text{eq}} - RT Z_{\text{eq}}^S + G_0. \quad (71)$$

As we discuss in Sec. III B, the equilibrium chemical potentials must satisfy $\nabla_\rho^\sigma \mu_\sigma^{\text{eq}} = 0$. We deduce that μ_σ^{eq} must be a linear combination of the closed system conservation laws Eq. (12),

$$\mu_\sigma^{\text{eq}} = f_\lambda \ell_\sigma^\lambda, \quad (72)$$

where $\{f_\lambda\}$ are real coefficients. Thus, we can write the equilibrium Gibbs free energy as

$$G_{\text{eq}} = f_\lambda L^\lambda - RT Z_{\text{eq}}^S + G_0. \quad (73)$$

In this form, the first term of the Gibbs free energy appears as a bilinear form of components $\{L^\lambda\}$ and conjugated generalized forces $\{f_\lambda\}$ (Ref. [23], Sec. III. 3), which can be thought of as chemical potentials of the components. From Eq. (72) and the properties of components Eq. (13), the equality $Z_{\text{eq}}^\sigma \mu_\sigma^{\text{eq}} = Z^\sigma \mu_\sigma^{\text{eq}}$ follows. Hence, using the definition of chemical potential Eq. (45), the nonequilibrium Gibbs free energy G of the generic distribution $\{Z^\sigma\}$ defined above is related to G_{eq} [Eq. (73)] by

$$G = G_{\text{eq}} + RT \mathcal{L}(\{Z^\sigma\}|\{Z_{\text{eq}}^\sigma\}), \quad (74)$$

where we introduce the relative entropy for non-normalized concentration distributions, also called the Shear Lyapunov function or the pseudo-Helmholtz function [35,73,74]:

$$\mathcal{L}(\{Z^\sigma\}|\{Z'^\sigma\}) \equiv Z^\sigma \ln \frac{Z^\sigma}{Z'^\sigma} - (Z^S - Z'^S) \geq 0. \quad (75)$$

This quantity is a natural generalization of the relative entropy, or Kullback-Leibler divergence, used to compare two normalized probability distributions [75]. For simplicity, we still refer to it as relative entropy. It quantifies the distance between two distributions: it is always positive and vanishes only if the two distributions are identical: $\{Z^\sigma\} = \{Z'^\sigma\}$. Hence, Eq. (74) proves that the nonequilibrium Gibbs free energy of a closed CRN is always greater than or equal to its corresponding equilibrium form, $G \geq G_{\text{eq}}$.

We now proceed to show that the nonequilibrium Gibbs free energy is minimized by the dynamics in closed CRNs; viz., G —or, equivalently, $\mathcal{L}(\{Z^\sigma\}|\{Z_{\text{eq}}^\sigma\})$ [59,76]—acts as a Lyapunov function in closed CRNs. Indeed, the time derivative of G Eq. (70) always reads

$$\begin{aligned} d_t G &= \mu_\sigma d_t Z^\sigma + Z^\sigma d_t \mu_\sigma + R d_t Z^S \\ &= \mu_\sigma d_t Z^\sigma. \end{aligned} \quad (76)$$

When using the rate equation for closed CRNs Eq. (7), we find that $d_t G = -J^\rho \nabla_\rho^\sigma \mu_\sigma$. Using Eq. (74) together with Eqs. (46) and (58), we get

$$d_t G = RT d_t \mathcal{L}(\{Z^\sigma\}|\{Z_{\text{eq}}^\sigma\}) = -T \dot{S}_i \leq 0, \quad (77)$$

which proves the aforementioned result.

3. Chemical work

In arbitrary CRNs, the rate of change of nonequilibrium Gibbs free energy, Eq. (76), can be related to the entropy production rate, Eq. (59), using the rate equations of open CRN, Eqs. (9) and (10), and the chemical work rate, Eq. (69):

$$T \dot{S}_i = \dot{W}_c - d_t G \geq 0. \quad (78)$$

This important result shows that the positivity of the entropy production sets an intrinsic limit on the chemical work that the chemostats must perform on the CRN to change its concentration distribution. The equality sign is achieved for quasistatic transformations ($\dot{S}_i \approx 0$).

If we now integrate Eq. (78) along a transformation generated by an arbitrary time-dependent protocol $\pi(t)$, which drives the CRN from an initial concentration distribution $\{Z_i^\sigma\}$ to a final one $\{Z_f^\sigma\}$, we find

$$T \Delta_i S = W_c - \Delta G \geq 0, \quad (79)$$

where $\Delta G = G_f - G_i$ is the difference of nonequilibrium Gibbs free energies between the final and the initial state. Let us also consider the equilibrium state $\{Z_{\text{eq}i}^\sigma\}$ ($\{Z_{\text{eq}f}^\sigma\}$) obtained from $\{Z_i^\sigma\}$ ($\{Z_f^\sigma\}$) if one closes the network (i.e., interrupts the chemostatting procedure) and lets it relax to equilibrium, as illustrated in Fig. 4. The Gibbs free energy difference between these two equilibrium distributions, $\Delta G_{\text{eq}} = G_{\text{eq}f} - G_{\text{eq}i}$, is related to ΔG via the difference of relative entropies, Eq. (74):

$$\Delta G = \Delta G_{\text{eq}} + RT \Delta \mathcal{L}, \quad (80)$$

where

$$\Delta \mathcal{L} \equiv \mathcal{L}(\{Z_f^\sigma\}|\{Z_{\text{eq}f}^\sigma\}) - \mathcal{L}(\{Z_i^\sigma\}|\{Z_{\text{eq}i}^\sigma\}). \quad (81)$$

Thus, the chemical work Eq. (79) can be rewritten as

$$W_c - \Delta G_{\text{eq}} = RT \Delta \mathcal{L} + T \Delta_i S, \quad (82)$$

which is a key result of our paper. ΔG_{eq} represents the reversible work needed to reversibly transform the CRN

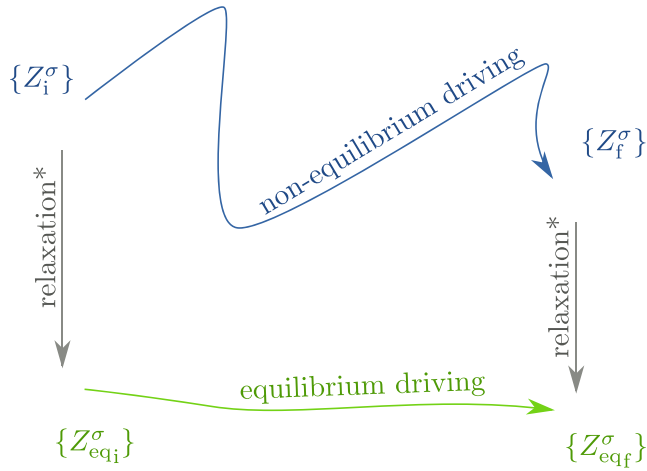


FIG. 4. Pictorial representation of the transformation between two nonequilibrium concentration distributions. The nonequilibrium transformation (blue line) is compared with the equilibrium one (green line). The equilibrium transformation depends on the equilibrium states corresponding to the initial and final concentration distributions. In Sec. III E 3, for an arbitrary CRN, these equilibrium states are obtained by first closing the network and then letting it relax to equilibrium. Instead, in Sec. V, for a detailed balance CRN, the equilibrium states are obtained by simply stopping the time-dependent driving and letting the system spontaneously relax to equilibrium.

from $\{Z_{\text{eq}i}^{\sigma}\}$ to $\{Z_{\text{eq}f}^{\sigma}\}$. Implementing such a reversible transformation may be difficult to achieve in practice. However, it allows us to interpret the difference $W_c^{\text{irr}} \equiv W_c - \Delta G_{\text{eq}}$ in Eq. (82) as the chemical work dissipated during the nonequilibrium transformation, i.e., the irreversible chemical work. The positivity of the entropy production implies that

$$W_c^{\text{irr}} \geq RT\Delta\mathcal{L}. \quad (83)$$

This relation sets limits on the irreversible chemical work involved in arbitrary far-from-equilibrium transformations. For transformations connecting two equilibrium distributions, we get the expected inequality $W_c^{\text{irr}} \geq 0$. More interestingly, Eq. (83) tells us how much chemical work the chemostat needs to provide to create a nonequilibrium distribution from an equilibrium one. It also tells us how much chemical work can be extracted from a CRN relaxing to equilibrium.

The conceptual analogue of Eq. (82) in stochastic thermodynamics (where probability distributions replace non-normalized concentration distributions) is called the nonequilibrium Landauer principle [42,43] (see also Refs. [77–79]). It has been shown to play a crucial role in analyzing the thermodynamic cost of information processing (e.g., for Maxwell demons, feedback control, or proofreading). The inequality Eq. (83) is, therefore, a nonequilibrium Landauer principle for CRN.

IV. THERMODYNAMICS OF COMPLEX-BALANCED NETWORKS

In this section, we focus on unconditionally complex-balanced networks. We see that the thermodynamics of these networks bears remarkable similarities to stochastic thermodynamics.

Let us first observe that whenever a CRN displays a well-defined steady-state distribution $\{\bar{Z}^{\sigma_x}\}$, the entropy production rate Eq. (55) can be formally decomposed as the sum of an adiabatic and a nonadiabatic contribution,

$$T\dot{S}_i = \underbrace{J^{\rho} RT \ln \frac{\bar{J}_{+\rho}}{\bar{J}_{-\rho}}}_{\equiv T\dot{S}_a} - d_t Z^{\sigma_x} RT \ln \frac{Z^{\sigma_x}}{\bar{Z}^{\sigma_x}}, \quad (84)$$

in analogy with what was done in stochastic thermodynamics [46–50]. As we discuss in Sec. II E, unconditionally complex-balanced networks have a unique steady-state distribution, $\{\bar{Z}^{\sigma_x} \equiv \bar{Z}^{\sigma_x}(\pi(t))\}$, Eq. (37), for any value of the chemostatted concentrations, $\{Z^{\sigma_y} \equiv Z^{\sigma_y}(\pi(t))\}$, and of the fixed unbroken components $\{L^{\lambda_u}\}$. The decomposition Eq. (84) is thus well defined at any time, for any protocol $\pi(t)$. As a central result, we prove in Appendix C that the adiabatic and nonadiabatic contribution are non-negative for unconditionally complex-balanced networks as well as for complex-balanced networks without time-dependent driving.

The adiabatic entropy production rate encodes the dissipation of the steady state $\{\bar{Z}^{\sigma_x}\}$. It can be rewritten in terms of the steady-state Gibbs free energy of reaction $\{\Delta_r \bar{G}_{\rho}\}$ Eq. (48) as

$$T\dot{S}_a = -J^{\rho} \Delta_r \bar{G}_{\rho} \geq 0. \quad (85)$$

This inequality highlights the fact that the transient dynamics—generating the currents $\{J^{\rho}\}$ —is constrained by the thermodynamics of the complex-balanced steady state, i.e., by $\{\Delta_r \bar{G}_{\rho}\}$.

The nonadiabatic entropy production rate characterizes the dissipation of the transient dynamics. It can be decomposed as

$$T\dot{S}_{\text{na}} = -RT d_t \mathcal{L}(\{Z^{\sigma_x}\} | \{\bar{Z}^{\sigma_x}\}) + \underbrace{RT d_t \bar{Z}^{\sigma_x} - Z^{\sigma_x} d_t \bar{\mu}_{\sigma_x}}_{\equiv T\dot{S}_d} \geq 0, \quad (86)$$

where $\bar{Z}^{\sigma_x} = \sum_{\sigma_x \in \mathcal{S}_x} \bar{Z}^{\sigma_x}$ (see Refs. [46,48] for the analogous decomposition in the stochastic context). The first term is proportional to the time derivative of the relative entropy Eq. (75) between the nonequilibrium concentration distribution at time t and the corresponding complex-balanced steady-state distribution. Hence, it describes the dissipation of the relaxation towards the steady state. The

second term, $T\dot{S}_d$, is related to the time-dependent driving performed via the chemostatted species and thus denoted the driving entropy production rate [46]. It vanishes in nondriven networks where we obtain

$$\dot{S}_{na} = -Rd_t \mathcal{L}(\{Z^{\sigma_x}\}|\{\bar{Z}^{\sigma_x}\}) \geq 0. \quad (87)$$

This result shows the role of the relative entropy $\mathcal{L}(\{Z^{\sigma_x}\}|\{\bar{Z}^{\sigma_x}\})$ as a Lyapunov function in nondriven complex-balanced networks with mass action kinetics. It was known in the mathematical literature [35,80], but we provide a clear thermodynamic interpretation to this result by demonstrating that it derives from the nonadiabatic entropy production rate.

We mention that an alternative derivation of the adiabatic–nonadiabatic decomposition for nondriven complex-balanced networks with mass action kinetics was very recently found in Ref. [51].

V. THERMODYNAMICS OF OPEN DETAILED-BALANCED NETWORKS

We finish our study by considering detailed-balanced networks. We discuss the equilibrium distribution, introduce a new class of nonequilibrium potentials, and derive a new work inequality.

Let us also emphasize that open detailed-balanced CRNs are a special class of open complex-balanced CRNs for which the adiabatic entropy production rate vanishes (since the steady state is detailed balanced) and thus the nonadiabatic entropy production characterizes the entire dissipation.

A. Equilibrium distribution

As we discuss in Sec. II D, for given kinetics $\{k^{\pm\rho}\}$, chemostatting $\{Z^{\sigma_y}\}$ and unbroken components $\{L^{\lambda_u}\}$, detailed-balanced networks always relax to a unique equilibrium distribution. Since the equilibrium chemical potentials can be expressed as a linear combination of conservation laws, Eq. (72), we can express the equilibrium distribution as

$$Z_{\sigma}^{\text{eq}} = \exp\left\{-\frac{\mu_{\sigma}^{\text{eq}} - f_{\lambda} \ell_{\sigma}^{\lambda}}{RT}\right\}, \quad (88)$$

inverting the expression for the chemical potentials Eq. (45). Since the independent set of unbroken conservation laws $\{\ell^{\lambda_u}\}$ is such that $\ell_{\sigma_y}^{\lambda_u} = 0, \forall \lambda_u, \sigma_y$, see Sec. II C, we have that

$$\mu_{\sigma_y}^{\text{eq}} = f_{\lambda_b} \ell_{\sigma_y}^{\lambda_b}, \quad \forall \sigma_y \in \mathcal{S}_y. \quad (89)$$

We thus conclude that the $|\lambda_b|$ broken generalized forces $\{f_{\lambda_b}\}$ depend on only the chemostatted concentrations $\{Z^{\sigma_y}\}$. Instead, the remaining $|\lambda_u|$ unbroken generalized

forces f_{λ_u} can be determined by inverting the nonlinear set of equations $L^{\lambda_u} = \ell_{\sigma_x}^{\lambda_u} Z_{\text{eq}}^{\sigma_x}$. They, therefore, depend on both $\{Z^{\sigma_y}\}$ and $\{L^{\lambda_u}\}$.

One can easily recover the local detailed-balanced property [Eqs. (50) and (18)] using Eq. (88).

B. Open nondriven networks

As a consequence of the break of conservation laws, the nonequilibrium Gibbs free energy G Eq. (70) is no longer minimized at equilibrium in open detailed-balanced networks. In analogy to equilibrium thermodynamics [23], the proper thermodynamic potential is obtained from G by subtracting the energetic contribution of the broken conservation laws. This transformed nonequilibrium Gibbs free energy reads

$$\begin{aligned} \mathcal{G} &\equiv G - f_{\lambda_b} L^{\lambda_b} \\ &= Z^{\sigma} (\mu_{\sigma} - f_{\lambda_b} \ell_{\sigma}^{\lambda_b}) - RTZ^S + G_0. \end{aligned} \quad (90)$$

We proceed to show that \mathcal{G} is minimized by the dynamics in nondriven open detailed-balanced networks. Let $\{Z^{\sigma_x}\}$ be a generic concentration distribution in a detailed-balanced network characterized by $\{L^{\lambda_u}\}$ and $\{Z^{\sigma_y}\}$, and let $\{Z_{\text{eq}}^{\sigma_x}\}$ be its corresponding equilibrium. Using the relation between equilibrium chemical potentials and conservation laws Eq. (72), the transformed Gibbs free energy Eq. (90) at equilibrium reads

$$\mathcal{G}_{\text{eq}} = f_{\lambda_u} L^{\lambda_u} - RTZ_{\text{eq}}^S + G_0. \quad (91)$$

Yet, combining Eq. (72) and the properties of unbroken components, one can readily show that $Z_{\text{eq}}^{\sigma} (\mu_{\sigma}^{\text{eq}} - f_{\lambda_b} \ell_{\sigma}^{\lambda_b}) = Z^{\sigma} (\mu_{\sigma}^{\text{eq}} - f_{\lambda_b} \ell_{\sigma}^{\lambda_b})$. The relation between the nonequilibrium \mathcal{G} and the corresponding equilibrium value thus follows

$$\mathcal{G} = \mathcal{G}_{\text{eq}} + RT\mathcal{L}(\{Z^{\sigma_x}\}|\{Z_{\text{eq}}^{\sigma_x}\}) \quad (92)$$

(we show in Appendix A the derivation of the latter in the presence of a reacting solvent). The non-negativity of the relative entropy for concentration distributions $\mathcal{L}(\{Z^{\sigma_x}\}|\{Z_{\text{eq}}^{\sigma_x}\})$ ensures that the nonequilibrium transformed Gibbs free energy is always greater than or equal to its equilibrium value, $\mathcal{G} \geq \mathcal{G}_{\text{eq}}$. Since entropy production and nonadiabatic entropy production coincide, using Eqs. (87) and (92), we obtain

$$d_t \mathcal{G} = RTd_t \mathcal{L}(\{Z^{\sigma_x}\}|\{Z_{\text{eq}}^{\sigma_x}\}) = -T\dot{S}_i \leq 0, \quad (93)$$

which demonstrates the role of \mathcal{G} as a Lyapunov function. The relative entropy $\mathcal{L}(\{Z^{\sigma_x}\}|\{Z_{\text{eq}}^{\sigma_x}\})$ was known to be a Lyapunov function for detailed-balanced networks [76,81], but we provide its clear connection to the transformed nonequilibrium Gibbs free energy. To summarize, instead

of minimizing the nonequilibrium Gibbs free energy G Eq. (70) as in closed CRNs, the dynamics minimizes the transformed nonequilibrium Gibbs free energy \mathcal{G} in open nondriven detailed-balanced CRNs.

C. Open driven networks

We now consider unconditionally detailed-balanced CRNs. As we discuss in Sec. IID, they are characterized by a unique equilibrium distribution $\{Z_{\text{eq}}^{\sigma_x} \equiv Z_{\text{eq}}^{\sigma_x}(\pi(t))\}$, defined by Eq. (18), for any value of the chemostatted concentrations $\{Z^{\sigma_y} = Z^{\sigma_y}(\pi(t))\}$.

We start by showing that the external fluxes $\{I^{\sigma_y}\}$ can be expressed as the influx rate of moieties. Since the CRN is open and unconditionally detailed balanced, each chemostatted species breaks a conservation law (no emergent cycle is created, Sec. IID). Therefore, the matrix whose entries are $\{\ell_{\sigma_y}^{\lambda_b}\}$ in Eq. (89) is square and also nonsingular [82]. We can thus invert Eq. (89) to get

$$f_{\lambda_b} = \mu_{\sigma_y}^{\text{eq}} \hat{\ell}_{\lambda_b}^{\sigma_y}, \quad (94)$$

where $\{\hat{\ell}_{\lambda_b}^{\sigma_y}\}$ denote the entries of the inverse matrix of that with entries $\{\ell_{\sigma_y}^{\lambda_b}\}$. Hence, using the definition of a broken component, $\{L^{\lambda_b} \equiv \ell_{\sigma}^{\lambda_b} Z^{\sigma}\}$, we obtain that

$$f_{\lambda_b} L^{\lambda_b} = \mu_{\sigma_y}^{\text{eq}} \underbrace{\hat{\ell}_{\lambda_b}^{\sigma_y} \ell_{\sigma}^{\lambda_b} Z^{\sigma}}_{\equiv M^{\sigma_y}}. \quad (95)$$

From the rate equations for the chemostatted concentrations Eq. (10), we find that

$$d_t M^{\sigma_y} = I^{\sigma_y}, \quad \forall \sigma_y \in \mathcal{S}_y. \quad (96)$$

We can thus interpret M^{σ_y} as the concentration of a moiety that is exchanged with the environment only through the chemostatted species X_{σ_y} . Equation (95) shows that the energetic contribution of the broken components can be expressed as the Gibbs free energy carried by these specific moieties.

Example 12.—A simple implementation of this scenario is the thermodynamic description of CRNs at constant $p\text{H}$ (Ref. [23], Chap. 4), where the chemostatted species becomes the ion H^+ and M^{H^+} is the total amount of H^+ ions in the system. The transformed Gibbs potential thus becomes $G' = G - \mu_{\text{H}^+} M^{\text{H}^+}$ and the transformed chemical potentials can be written in our formalism as $\mu'_{\sigma_x} = \mu_{\sigma_x} - \mu_{\text{H}^+} \hat{\ell}_{\text{H}^+}^{\sigma_x} \ell_{\sigma}^{\text{H}^+}$, where $\ell_{\sigma}^{\text{H}^+}$ is the conservation law broken by chemostatting H^+ . \square

Example 13.—For the CRN in Fig. 2, whose conservation laws are given in example 4, the concentrations of the exchanged moieties are

$$\begin{aligned} M^1 &= Z^a + \frac{1}{2} Z^b, \\ M^2 &= Z^d + Z^e. \end{aligned} \quad (97)$$

For the specific implementation of that CRN, Fig. 3, the first term (second term) is the total number of moiety 2H (C) in the system, which can be exchanged with the environment only via the chemostatted species H_2O (CO). \square

We now turn to the new work relation. From the general work relation Eq. (78), using Eqs. (90) and (95), we find

$$T\dot{S}_i = \dot{W}_d - d_t \mathcal{G} \geq 0, \quad (98)$$

where the driving work due to the time-dependent driving of the chemostatted species is obtained using the chemical work rate Eq. (69) together with Eqs. (95) and (96):

$$\begin{aligned} \dot{W}_d &\equiv \dot{W}_c - d_t (f_{\lambda_b} L^{\lambda_b}) \\ &= \mu_{\sigma_y}^{\text{eq}} d_t M^{\sigma_y} - d_t (\mu_{\sigma_y}^{\text{eq}} M^{\sigma_y}) \\ &= -d_t \mu_{\sigma_y}^{\text{eq}} M^{\sigma_y}. \end{aligned} \quad (99)$$

Equivalently, the driving work rate Eq. (99) can be defined as the rate of change of the transformed Gibbs free energy Eq. (90) due to the time-dependent driving only; i.e.,

$$\dot{W}_d \equiv \frac{\partial \mathcal{G}}{\partial t} \equiv d_t \mu_{\sigma_y}^{\text{eq}} \frac{\partial \mathcal{G}}{\partial \mu_{\sigma_y}^{\text{eq}}}. \quad (100)$$

To relate this alternative definition to Eq. (99), all $\{Z^{\sigma_y}\}$ must be expressed in terms of $\{\mu_{\sigma_y}^{\text{eq}}\}$ using the definition of chemical potential Eq. (45).

The driving work rate \dot{W}_d vanishes in nondriven CRNs, where Eq. (98) reduces to Eq. (93). After demonstrating that the entropy production rate is always proportional to the difference between the chemical work rate and the change of nonequilibrium Gibbs free energy in Eq. (79), we show that, for unconditionally detailed-balanced CRNs, it is also proportional to the difference between the driving work rate and the change in transformed nonequilibrium Gibbs free energy, Eq. (98).

We end by formulating a nonequilibrium Landauer principle for the driving work instead of the chemical work done in Sec. III E 3. We consider a time-dependent transformation driving the unconditionally detailed-balanced CRN from $\{Z_i^{\sigma}\}$ to $\{Z_f^{\sigma}\}$. The distribution $\{Z_{\text{eq}_i}^{\sigma}\}$ ($\{Z_{\text{eq}_f}^{\sigma}\}$) denotes the equilibrium distribution obtained from $\{Z_i^{\sigma}\}$ ($\{Z_f^{\sigma}\}$) by stopping the time-dependent driving and letting the system relax towards the equilibrium, Fig. 4. Note that this reference equilibrium state is different from the one obtained by closing the network in Sec. III E 3. Integrating Eq. (98) over time and using Eq. (92), we get

$$W_d - \Delta \mathcal{G}_{\text{eq}} = RT\Delta \mathcal{L} + T\Delta_i S, \quad (101)$$

where

$$\Delta \mathcal{L} \equiv \mathcal{L}(\{Z_f^{\sigma_x}\}|\{Z_{\text{eqf}}^{\sigma_x}\}) - \mathcal{L}(\{Z_i^{\sigma_x}\}|\{Z_{\text{eqi}}^{\sigma_x}\}). \quad (102)$$

$\Delta \mathcal{G}_{\text{eq}}$ represents the reversible driving work, and the irreversible driving work satisfies the inequality

$$W_d^{\text{irr}} \equiv W_d - \Delta \mathcal{G}_{\text{eq}} \geq RT\Delta \mathcal{L}. \quad (103)$$

This central relation sets limits on the irreversible work spent to manipulate nonequilibrium distributions. It is a nonequilibrium Landauer principle for the driving work by the same reasons that inequality Eq. (83) is a nonequilibrium Landauer principle for the chemical work. The key difference is that the choice of the reference equilibrium state is different in the two cases. The above discussed inequality Eq. (103) holds only for unconditionally detailed-balanced CRNs, while Eq. (83) is valid for any CRNs.

VI. CONCLUSIONS AND PERSPECTIVES

Following a strategy reminiscent of stochastic thermodynamics, we systematically build a nonequilibrium thermodynamic description for open driven CRNs made of elementary reactions in homogeneous ideal dilute solutions. The dynamics is described by deterministic rate equations whose kinetics satisfies mass action law. Our framework is not restricted to steady states and allows us to treat transients as well as time-dependent drivings performed by externally controlled chemostatted concentrations. Our theory embeds the nonequilibrium thermodynamic framework of irreversible processes established by the Brussels School of Thermodynamics.

We now summarize our results. Starting from the expression for the entropy production rate, we establish a nonequilibrium formulation of the first and second law of thermodynamics for CRNs. The resulting expression for the system entropy is that of an ideal dilute solution. The clear separation between chemostatted and internal species allows us to identify the chemical work done by the chemostats on the CRN and to relate it to the nonequilibrium Gibbs potential. We are also able to express the minimal chemical work necessary to change the nonequilibrium distribution of species in the CRN as a difference of relative entropies for non-normalized distributions. The latter measure the distance of the initial and final concentration distributions from their corresponding equilibrium ones, obtained by closing the network. This result is reminiscent of the nonequilibrium Landauer principle derived in stochastic thermodynamics [43] and which prove very useful to study the energetic cost of information processing [45]. We also highlight the deep relationship between the topology of CRNs, their dynamics, and their

thermodynamics. Closed CRNs (nondriven open detailed-balanced networks) always relax to a unique equilibrium by minimizing their nonequilibrium Gibbs free energy (transformed nonequilibrium Gibbs free energy). This latter is given, up to a constant, by the relative entropy between the nonequilibrium and equilibrium concentration distribution. Nondriven complex-balanced networks relax to complex-balanced nonequilibrium steady states by minimizing the relative entropy between the nonequilibrium and steady-state concentration distribution. In all these cases, even in the presence of driving, we show how the rate of change of the relative entropy relates to the CRN dissipation. For complex-balanced networks, we also demonstrate that the entropy production rate can be decomposed, as in stochastic thermodynamics, in its adiabatic and nonadiabatic contributions quantifying, respectively, the dissipation of the steady state and of the transient dynamics.

Our framework could be used to shed new light on a broad range of problems. We mention only a few.

Stochastic thermodynamics has been successfully used to study the thermodynamics cost of information processing in various synthetic and biological systems [44,83–87]. However, most of these are modeled as few state systems or linear networks [8,9]—e.g., quantum dots [88], molecular motors [89,90], and single enzyme mechanisms [91,92]—while biochemical networks involve more-complex descriptions. The present work overcomes this limitation. It could be used to study biological information-handling processes, such as kinetic proofreading [93–99] or enzyme-assisted copolymerization [92,100–105], which have currently only been studied as single enzyme mechanisms.

Our theory could also be used to study metabolic networks. However, these require some care, since complex enzymatic reaction mechanisms are involved [106]. Nevertheless, our framework provides a basis to build effective coarse-graining procedures for enzymatic reactions [107]. For instance, proofreading mechanisms operating in metabolic processes could be considered [108]. We foresee an increasing use of thermodynamics to improve the modeling of metabolic networks, as recently shown in Refs. [30,32,33].

Since our framework accounts for time-dependent drivings and transient dynamics, it could be used to represent the transmission of signals through CRNs or their response to external modulations in the environment. These features become crucial when considering problems such as signal transduction and biochemical switches [24,109,110], biochemical oscillations [28,111], growth and self-organization in evolving biosystems [112,113], or sensory mechanisms [85,87,114–117]. Also, since transient signals in CRNs can be used for computation [118] and have been shown to display Turing universality [119–122], one should be able to study the thermodynamic cost of chemical computing [123].

Finally, one could use our framework to study any process that can be described as nucleation or reversible polymerization [124–129] (see also Ref. [130], Chaps. 5 and 6) since these processes can be described as CRNs [63].

As closing words, we believe that our results constitute an important contribution to the theoretical study of CRNs. It does for nonlinear chemical kinetics what stochastic thermodynamics has done for stochastic dynamics, namely, build a systematic nonequilibrium thermodynamics on top of the dynamics. It also opens many new perspectives and builds bridges between approaches from different communities studying CRNs: mathematicians who study CRNs as dynamical systems, physicists who study them as nonequilibrium complex systems, and biochemists as well as bioengineers who aim for accurate models of metabolic networks.

ACKNOWLEDGMENTS

The present project was supported by the Luxembourg National Research Fund (Project No. FNR/A11/02 and AFR Ph.D. Grant 2014-2, No. 9114110) as well as by the European Research Council (Project No. 681456).

APPENDIX A: THERMODYNAMICS OF IDEAL DILUTE SOLUTIONS

We show that the nonequilibrium Gibbs free energy Eq. (70) is the Gibbs free energy of an ideal dilute solution (Ref. [131], Chap. 7) (see also Ref. [51]). We also show that in open detailed-balanced networks in which the solvent reacts with the solutes, the expression of the transformed Gibbs free energy Eq. (92) is recovered by treating the solvent as a special chemostatted species.

The Gibbs free energy (density) of an ideal dilute mixture of chemical compounds kept at constant temperature and pressure reads

$$G = Z^\sigma \mu_\sigma + Z_0 \mu_0 \quad (\text{A1})$$

where the labels $\sigma \in \mathcal{S}$ refer to the solutes and 0 to the solvent. The chemical potentials of each species Eq. (45) read

$$\begin{aligned} \mu_\sigma &= \mu_\sigma^\circ + RT \ln \frac{Z_\sigma}{Z_{\text{tot}}}, \quad \forall \sigma \in \mathcal{S}, \\ \mu_0 &= \mu_0^\circ + RT \ln \frac{Z_0}{Z_{\text{tot}}}. \end{aligned} \quad (\text{A2})$$

Since the solution is dilute, $Z_{\text{tot}} = \sum_{\sigma \in \mathcal{S}} Z^\sigma + Z_0 \approx Z_0$ and the standard-state chemical potentials $\{\mu_\sigma^\circ\}$ depend on the nature of the solvent. Hence, the chemical potentials of the solutes read

$$\mu_\sigma \approx \mu_\sigma^\circ + RT \ln \frac{Z_\sigma}{Z_0}, \quad \forall \sigma \in \mathcal{S}, \quad (\text{A3})$$

while that of the solvent reads

$$\mu_0 \approx \mu_0^\circ - RT \frac{Z^\mathcal{S}}{Z_0}, \quad (\text{A4})$$

where $Z^\mathcal{S} \equiv \sum_{\sigma \in \mathcal{S}} Z^\sigma$. Therefore, the Gibbs free energy Eq. (A1) reads

$$G \approx Z^\sigma \mu_\sigma + Z^0 \mu_0^\circ - RT Z^\mathcal{S}, \quad (\text{A5})$$

which is Eq. (70), where G_0 is equal to $Z^0 \mu_0^\circ$ plus possibly the Gibbs free energy of solutes that do not react.

We now consider the case where the solvent reacts with the solutes. We assume that both the solutes and the solvent react according to the stoichiometric matrix

$$\nabla = \begin{pmatrix} \nabla^0 \\ \nabla^X \\ \nabla^Y \end{pmatrix}, \quad (\text{A6})$$

where the first row refers to the solvent, the second block of rows to the internal species, and the last one to the chemostatted species. The solvent is treated as a chemostatted species such that $d_t Z_0 = 0$.

In order to recover the expression for the transformed Gibbs free energy Eq. (92) in unconditionally detailed-balanced networks, we observe that, at equilibrium,

$$\nabla_\rho^\sigma \mu_\sigma^{\text{eq}} + \nabla_\rho^0 \mu_0^{\text{eq}} = 0. \quad (\text{A7})$$

Therefore, the equilibrium chemical potentials are a linear combination of the conservation laws of ∇ [Eq. (A6)],

$$\begin{aligned} \mu_\sigma^{\text{eq}} &= f_\lambda \mathcal{L}_\sigma^\lambda, \\ \mu_0^{\text{eq}} &= f_\lambda \mathcal{L}_0^\lambda. \end{aligned} \quad (\text{A8})$$

As mentioned Sec. II C, the chemostatting procedure breaks some conservation laws, which are labeled by λ_b . The unbroken ones are labeled by λ_u .

The transformed Gibbs free energy is defined as in Eq. (90), reported here for convenience,

$$\mathcal{G} \equiv G - f_{\lambda_b} L^{\lambda_b}, \quad (\text{A9})$$

where G reads as in Eq. (A1), $\{L^{\lambda_b}\}$ are the broken components, and $\{f_{\lambda_b}\}$ are here interpreted as the conjugated generalized forces. Adding and subtracting the term $Z^\sigma \mu_\sigma^{\text{eq}} + Z_0 \mu_0^{\text{eq}}$ from the last equation and using Eq. (A8), we obtain

$$G = \mathcal{G}_{\text{eq}} + Z^\sigma (\mu_\sigma - \mu_\sigma^{\text{eq}}) + Z^0 (\mu_0 - \mu_0^{\text{eq}}), \quad (\text{A10})$$

where

$$\mathcal{G}_{\text{eq}} = f_{\lambda_u} L^{\lambda_u}. \quad (\text{A11})$$

From Eqs. (A3) and (A4) and the fact that $Z^{\sigma_y} = Z_{\text{eq}}^{\sigma_y}$ and $Z_0 = Z_0^{\text{eq}}$, we obtain

$$\begin{aligned} \mathcal{G} &\simeq \mathcal{G}_{\text{eq}} + Z^{\sigma_x} (\mu_{\sigma_x} - \mu_{\sigma_x}^{\text{eq}}) - RT(Z^{\sigma_x} - Z_{\text{eq}}^{\sigma_x}) \\ &= \mathcal{G}_{\text{eq}} + Z^{\sigma_x} RT \ln \frac{Z^{\sigma_x}}{Z_{\text{eq}}^{\sigma_x}} - RT(Z^{\sigma_x} - Z_{\text{eq}}^{\sigma_x}) \\ &\equiv \mathcal{G}_{\text{eq}} + RT\mathcal{L}(\{Z^{\sigma_x}\}|\{Z_{\text{eq}}^{\sigma_x}\}), \end{aligned} \quad (\text{A12})$$

in agreement with the expression derived in the main text, Eq. (92).

APPENDIX B: ENTROPY OF CRNs

We show how the nonequilibrium entropy Eq. (63) can be obtained as a large particle limit of the stochastic entropy. We point out that very recently similar derivations for other thermodynamic quantities have been obtained in Refs. [51,132].

In the stochastic description of CRNs, the state is characterized by the population vector $\mathbf{n} = \{n^\sigma\}$. The probability to find the network is in state \mathbf{n} at time t is denoted $p_t(\mathbf{n})$. The stochastic entropy of that state reads [21,107], up to constants,

$$S(\mathbf{n}) = -k_B \ln p_t(\mathbf{n}) + s(\mathbf{n}). \quad (\text{B1})$$

The first term is a Shannon-like contribution, while the second term is the configurational entropy,

$$s(\mathbf{n}) \equiv n^\sigma \tilde{s}_\sigma^\circ - k_B \sum_\sigma \ln \frac{n^\sigma!}{n_0^{n^\sigma}}. \quad (\text{B2})$$

\tilde{s}_σ° is the standard entropy of one single X_σ molecule, and n_0 is the very large number of solvent molecules.

We now assume that the probability becomes very narrow in the large particle limit $n^\sigma \gg 1$ and behaves as a discrete delta function $p_t(\mathbf{n}) \simeq \delta[\mathbf{n} - \hat{\mathbf{n}}(t)]$. The vector $\hat{\mathbf{n}}(t) \equiv \{\hat{n}^\sigma\}$ denotes the most probable and macroscopic amount of chemical species, such that $Z^\sigma = \hat{n}^\sigma / (VN_A)$. Hence, the average entropy becomes

$$\langle S \rangle = \sum_{\mathbf{n}} p_t(\mathbf{n}) S(\mathbf{n}) \simeq s(\hat{\mathbf{n}}). \quad (\text{B3})$$

When using the Stirling approximation ($\ln m! \simeq m \ln m - m$ for $m \gg 1$), we obtain

$$\begin{aligned} s(\hat{\mathbf{n}}) &\simeq \hat{n}^\sigma \tilde{s}_\sigma^\circ - \hat{n}^\sigma k_B \ln \frac{\hat{n}^\sigma}{n_0} + k_B \sum_\sigma \hat{n}^\sigma \\ &= \hat{n}^\sigma \left(\tilde{s}_\sigma^\circ + k_B \ln \frac{n_0}{VN_A} \right) \\ &\quad + -\hat{n}^\sigma k_B \ln \frac{\hat{n}^\sigma}{VN_A} + k_B \sum_\sigma \hat{n}^\sigma \\ &\equiv \hat{n}^\sigma (\tilde{s}_\sigma^\circ + k_B \ln Z_0) \\ &\quad + -\hat{n}^\sigma k_B \ln Z^\sigma + k_B \sum_\sigma \hat{n}^\sigma. \end{aligned} \quad (\text{B4})$$

Dividing by V and using the relation $R = N_A k_B$, we finally get the macroscopic entropy density Eq. (63)

$$\langle S \rangle / V \simeq Z^\sigma s_\sigma^\circ - Z^\sigma R \ln Z_\sigma + R Z^\sigma S, \quad (\text{B5})$$

where the (molar) standard entropies of formation s_σ° reads

$$s_\sigma^\circ = N_A (\tilde{s}_\sigma^\circ + k_B \ln Z_0). \quad (\text{B6})$$

Mindful of the information-theoretical interpretation of the entropy [133], we note that the uncertainty due to the stochasticity of the state disappears [the first term on the rhs of Eq. (B1)]. However, the uncertainty due to the indistinguishability of the molecules of the same species—quantified by the configurational entropy Eq. (B2)—remains and contributes to the whole deterministic entropy function Eq. (63).

APPENDIX C: ADIABATIC-NONADIABATIC DECOMPOSITION

We prove the positivity of the adiabatic and nonadiabatic entropy production rates Eq. (84) using the theory of complex-balanced networks; see Sec. II E.

We first rewrite the mass action kinetics currents Eq. (5) as [53,81] $J^\rho = K_\gamma^\rho \psi^\gamma$, where $\psi^\gamma \equiv Z^{\sigma \Gamma_\sigma^\gamma}$ and $K = \{K_\gamma^\rho \equiv K_\gamma^{+\rho} - K_\gamma^{-\rho}\}$ is the rate constants matrix whose entries are defined by

$$K_\gamma^\rho = \begin{cases} k^{+\rho} & \text{if } \gamma \text{ is the reactant complex of } +\rho \\ -k^{-\rho} & \text{if } \gamma \text{ is the product complex of } +\rho \\ 0 & \text{otherwise.} \end{cases} \quad (\text{C1})$$

Hence, the definition of a complex-balanced network Eq. (37) reads

$$\sum_{\gamma \in \mathcal{C}_j} \mathcal{W}_\gamma^j \bar{\psi}^\gamma = 0, \quad \forall j, \quad (\text{C2})$$

where $\mathcal{W} \equiv \partial K = \{\partial_\rho^\gamma K_\gamma^\rho\} \equiv \{\mathcal{W}_\gamma^j\}$ is the so-called kinetic matrix [35], and $\bar{\psi}^\gamma \equiv \bar{Z}^{\sigma \Gamma_\sigma^\gamma}$.

The kinetic matrix \mathcal{W} is a Laplacian matrix [76,81]: any off-diagonal term is equal to the rate constant of the reaction having γ' as a reactant and γ as a product if the reaction exists, and it is zero otherwise. Also, it satisfies

$$\sum_{\gamma \in \mathcal{C}} \mathcal{W}_{\gamma'}^{\gamma} = 0, \quad (\text{C3})$$

which is a consequence of the fact that the diagonal terms are equal to minus the sum of the off-diagonal terms along the columns. The detailed balanced property Eq. (18) implies that

$$\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\text{eq}} = \mathcal{W}_{\gamma}^{\gamma'} \psi_{\gamma}^{\text{eq}}, \quad \forall \gamma, \gamma', \quad (\text{C4})$$

where $\psi_{\gamma}^{\text{eq}} \equiv Z_{\sigma}^{\text{eq}} \Gamma_{\gamma}^{\sigma}$.

In order to prove the non-negativity of the adiabatic term Eq. (84), we rewrite it as

$$\begin{aligned} \dot{S}_{\text{a}} &\equiv J^{\rho} \ln \frac{\bar{J}_{+\rho}}{\bar{J}_{-\rho}} = K_{\gamma'}^{\rho} \psi_{\gamma'}^{\rho} \ln \left(\frac{\bar{Z}_{\sigma}}{Z_{\sigma}^{\text{eq}}} \right)^{-\nabla_{\rho}^{\sigma}} \\ &= -\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \ln \frac{\bar{\psi}_{\gamma}^{\rho}}{\psi_{\gamma}^{\text{eq}}}. \end{aligned} \quad (\text{C5})$$

The detailed balance property is used in the first equality, and the decomposition of the stoichiometric matrix Eq. (30) in the second one. Also, the constant RT is taken equal to 1. Using Eq. (C3), Eq. (C5) can be rewritten as

$$\dot{S}_{\text{a}} = -\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \ln \frac{\bar{\psi}_{\gamma}^{\rho} \psi_{\gamma'}^{\text{eq}}}{\psi_{\gamma}^{\text{eq}} \bar{\psi}_{\gamma'}^{\rho}}. \quad (\text{C6})$$

From the log inequality $-\ln x \geq 1 - x$ and the detailed balance property Eq. (C4), we obtain

$$\begin{aligned} \dot{S}_{\text{a}} &\geq \mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \left(1 - \frac{\bar{\psi}_{\gamma}^{\rho} \psi_{\gamma'}^{\text{eq}}}{\psi_{\gamma}^{\text{eq}} \bar{\psi}_{\gamma'}^{\rho}} \right) \\ &= -\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\text{eq}} \frac{\bar{\psi}_{\gamma}^{\rho} \psi_{\gamma'}^{\rho}}{\psi_{\gamma}^{\text{eq}} \bar{\psi}_{\gamma'}^{\rho}} = -\mathcal{W}_{\gamma'}^{\gamma} \bar{\psi}_{\gamma}^{\rho} \frac{\psi_{\gamma'}^{\rho}}{\bar{\psi}_{\gamma'}^{\rho}} = 0. \end{aligned} \quad (\text{C7})$$

The last equality follows from the assumption of a complex-balanced steady state Eq. (C2), the properties of the groups of complexes $\{\mathcal{C}_j\}$ (Sec. II E), and the fact that $\{Z_{\sigma_y} = \bar{Z}_{\sigma_y}\}$. Indeed,

$$\begin{aligned} \mathcal{W}_{\gamma'}^{\gamma} \bar{\psi}_{\gamma}^{\rho} \frac{\psi_{\gamma'}^{\rho}}{\bar{\psi}_{\gamma'}^{\rho}} &= \sum_j \sum_{\rho' \in \mathcal{C}_j} \mathcal{W}_{\gamma'}^{\rho'} \bar{\psi}_{\gamma}^{\rho'} \left(\frac{Z_{\sigma_x}}{\bar{Z}_{\sigma_x}} \right)^{\Gamma_{\gamma'}^{\sigma_x}} \\ &= \sum_j \left(\frac{Z_{\sigma_x}}{\bar{Z}_{\sigma_x}} \right)^{\Gamma_j^{\sigma_x}} \sum_{\rho' \in \mathcal{C}_j} \mathcal{W}_{\gamma'}^{\rho'} \bar{\psi}_{\gamma}^{\rho'} = 0. \end{aligned} \quad (\text{C8})$$

Concerning the nonadiabatic term Eq. (84), using the rate equations (9) and the fact that $\{Z_{\sigma_y} = \bar{Z}_{\sigma_y}\}$, we can rewrite it as

$$\dot{S}_{\text{na}} \equiv -d_{\gamma} Z^{\sigma} \ln \frac{Z_{\sigma}}{Z_{\sigma}^{\text{eq}}} = -\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \ln \frac{\psi_{\gamma}^{\rho}}{\bar{\psi}_{\gamma}^{\rho}}. \quad (\text{C9})$$

Because of Eq. (C3), we further get that

$$\dot{S}_{\text{na}} = -\mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \ln \frac{\psi_{\gamma}^{\rho} \bar{\psi}_{\gamma'}^{\rho}}{\bar{\psi}_{\gamma}^{\rho} \psi_{\gamma'}^{\rho}}. \quad (\text{C10})$$

From the log inequality $-\ln x \geq 1 - x$ and from Eq. (C4),

$$\dot{S}_{\text{na}} \geq \mathcal{W}_{\gamma'}^{\gamma} \psi_{\gamma'}^{\rho} \left(1 - \frac{\psi_{\gamma}^{\rho} \bar{\psi}_{\gamma'}^{\rho}}{\bar{\psi}_{\gamma}^{\rho} \psi_{\gamma'}^{\rho}} \right) = -\mathcal{W}_{\gamma'}^{\gamma} \bar{\psi}_{\gamma}^{\rho} \frac{\psi_{\gamma}^{\rho}}{\bar{\psi}_{\gamma}^{\rho}} = 0. \quad (\text{C11})$$

The last equality again follows from the assumption of a complex-balance steady state Eq. (C2) as in Eq. (C8).

APPENDIX D: DEFICIENCY OF CRNs

The deficiency of an open CRN is defined as [22]

$$\delta = \dim \ker \nabla^{\text{X}} - \dim \ker \partial^{\mathcal{C}} \geq 0, \quad (\text{D1})$$

where $\partial^{\mathcal{C}} = \{\partial_{\rho}^j \equiv \sum_{\gamma \in \mathcal{C}_j} \partial_{\rho}^{\gamma}\}$. Other equivalent definitions can be found in Refs. [52,53]. The kernel of ∇^{X} identifies the set of cycles, Eqs. (19) and (20), while the kernel of the incidence matrix $\hat{\partial}$ identifies the set of cycles of the reaction graph. Hence, the deficiency measures the difference between the number of cyclic transformations on chemical species and how many of them can be represented as cycles on the reaction graph. Deficiency-zero networks are defined by $\delta = 0$; i.e., they exhibit a one-to-one correspondence between the two. This topological property has many dynamical consequences, the most important of which is that deficiency-zero networks are unconditionally complex balanced [36,37]. As shown in Ref. [22], deficiency also has implications on the stochastic thermodynamic description of networks: the stochastic entropy production of a deficiency-zero network converges to the deterministic entropy production in the long-time limit. Linear networks are the simplest class of deficiency-zero networks. Since only one internal species appears in each complex with a stoichiometric coefficient equal to 1, $\nabla^{\text{X}} \equiv \partial^{\mathcal{C}}$, and thus $\delta = 0$.

-
- [1] J. W. Gibbs, *The Scientific Papers of J. Willard Gibbs, Vol. 1: Thermodynamics* (Dover, New York, 1961).
 [2] T. de Donder, *L'affinité*, Mémoires de la Classe des Sciences (Gauthier-Villars, Paris, 1927), Vol. 1.

- [3] I. Prigogine, *Etude Thermodynamique des Phénomènes Irréversibles* (Desoer, Liège, 1947).
- [4] I. Prigogine, *Introduction to Thermodynamics of Irreversible Processes* (John Wiley & Sons, New York, 1967).
- [5] I. Prigogine and R. Defay, *Chemical Thermodynamics* (Longmans, Green & Co., New York, 1954).
- [6] G. F. Oster, A. S. Perelson, and A. Katchalsky, *Network Thermodynamics: Dynamic Modelling of Biophysical Systems*, *Q. Rev. Biophys.* **6**, 1 (1973).
- [7] M. Malek-Mansour and G. Nicolis, *A Master Equation Description of Local Fluctuations*, *J. Stat. Phys.* **13**, 197 (1975).
- [8] J. Schnakenberg, *Network Theory of Microscopic and Macroscopic Behavior of Master Equation Systems*, *Rev. Mod. Phys.* **48**, 571 (1976).
- [9] T. L. Hill, *Free Energy Transduction in Biology* (Academic Press, New York, 1977).
- [10] C. Y. Mou, J. Luo, and G. Nicolis, *Stochastic Thermodynamics of Nonequilibrium Steady States in Chemical Reaction Systems*, *J. Chem. Phys.* **84**, 7011 (1986).
- [11] J. Ross, *Thermodynamics and Fluctuations Far from Equilibrium* (Springer, New York, 2008).
- [12] D. A. McQuarrie, *Stochastic Approach to Chemical Kinetics*, *J. Appl. Probab.* **4**, 413 (1967).
- [13] D. T. Gillespie, *A Rigorous Derivation of the Chemical Master Equation*, *Physica A* **188**, 404 (1992).
- [14] K. Sekimoto, in *Stochastic Energetics*, 1st ed., Lecture Notes in Physics (Springer-Verlag, Berlin, 2010), Vol. 799.
- [15] C. Jarzynski, *Equalities and Inequalities: Irreversibility and the Second Law of Thermodynamics at the Nanoscale*, *Annu. Rev. Condens. Matter Phys.* **2**, 329 (2011).
- [16] U. Seifert, *Stochastic Thermodynamics, Fluctuation Theorems and Molecular Machines*, *Rep. Prog. Phys.* **75**, 126001 (2012).
- [17] C. Van den Broeck and M. Esposito, *Ensemble and Trajectory Thermodynamics: A Brief Introduction*, *Physica A* **418**, 6 (2015).
- [18] P. Gaspard, *Fluctuation Theorem for Nonequilibrium Reactions*, *J. Chem. Phys.* **120**, 8898 (2004).
- [19] D. Andrieux and P. Gaspard, *Fluctuation Theorem and Onsager Reciprocity Relations*, *J. Chem. Phys.* **121**, 6167 (2004).
- [20] T. Schmiedl, T. Speck, and U. Seifert, *Entropy Production for Mechanically or Chemically Driven Biomolecules*, *J. Stat. Phys.* **128**, 77 (2007).
- [21] T. Schmiedl and U. Seifert, *Stochastic Thermodynamics of Chemical Reaction Networks*, *J. Chem. Phys.* **126**, 044101 (2007).
- [22] M. Poletini, A. Wachtel, and M. Esposito, *Dissipation in Noisy Chemical Networks: The Role of Deficiency*, *J. Chem. Phys.* **143**, 184103 (2015).
- [23] R. A. Alberty, *Thermodynamics of Biochemical Reactions* (Wiley-Interscience, New York, 2003).
- [24] D. A. Beard and H. Qian, *Chemical Biophysics. Quantitative Analysis of Cellular Systems* (Cambridge University Press, Cambridge, England, 2008).
- [25] B. Ø. Palsson, *Systems Biology: Properties of Reconstructed Networks* (Cambridge University Press, Cambridge, England, 2006).
- [26] B. Ø. Palsson, *Systems Biology: Simulation of Dynamic Network States* (Cambridge University Press, Cambridge, England, 2011).
- [27] B. Ø. Palsson, *Systems Biology: Constraint-Based Network Reconstruction and Analysis* (Cambridge University Press, Cambridge, England, 2015).
- [28] A. Goldbeter, *Biochemical Oscillations and Cellular Rhythms* (Cambridge University Press, Cambridge, England, 1996).
- [29] I. R. Epstein and J. A. Pojman, in *An Introduction to Nonlinear Chemical Dynamics: Oscillations, Waves, Patterns, and Chaos*, Topics in Physical Chemistry (Oxford University Press, New York, 1998).
- [30] D. A. Beard, S.-d. Liang, and H. Qian, *Energy Balance for Analysis of Complex Metabolic Networks*, *Biophys. J.* **83**, 79 (2002).
- [31] H. Qian, D. A. Beard, and S.-d. Liang, *Stoichiometric Network Theory for Nonequilibrium Biochemical Systems*, *Eur. J. Biochem.* **270**, 415 (2003).
- [32] D. A. Beard, E. Babson, E. Curtis, and H. Qian, *Thermodynamic Constraints for Biochemical Networks*, *J. Theor. Biol.* **228**, 327 (2004).
- [33] A. Chakrabarti, L. Miskovic, K. C. Soh, and V. Hatzimanikatis, *Towards Kinetic Modeling of Genome-Scale Metabolic Networks without Sacrificing Stoichiometric, Thermodynamic and Physiological Constraints*, *Biotechnol. J.* **8**, 1043 (2013).
- [34] H. Qian and D. A. Beard, *Thermodynamics of Stoichiometric Biochemical Networks in Living Systems Far from Equilibrium*, *Biophys. Chem.* **114**, 213 (2005).
- [35] F. Horn and R. Jackson, *General Mass Action Kinetics*, *Arch. Ration. Mech. Anal.* **47**, 81 (1972).
- [36] M. Feinberg, *Complex Balancing in General Kinetic Systems*, *Arch. Ration. Mech. Anal.* **49**, 187 (1972).
- [37] F. Horn, *Necessary and Sufficient Conditions for Complex Balancing in Chemical Kinetics*, *Arch. Ration. Mech. Anal.* **49**, 172 (1972).
- [38] T. G. Kurtz, *The Relationship between Stochastic and Deterministic Models for Chemical Reactions*, *J. Chem. Phys.* **57**, 2976 (1972).
- [39] D. F. Anderson, G. Craciun, and T. G. Kurtz, *Product-Form Stationary Distributions for Deficiency Zero Chemical Reaction Networks*, *Bull. Math. Biol.* **72**, 1947 (2010).
- [40] D. F. Anderson, G. Craciun, M. Gopalkrishnan, and C. Wiuf, *Lyapunov Functions, Stationary Distributions, and Non-Equilibrium Potential for Reaction Networks*, *Bull. Math. Biol.* **77**, 1744 (2015).
- [41] D. Cappelletti and C. Wiuf, *Product-Form Poisson-like Distributions and Complex Balanced Reaction Systems*, *SIAM J. Appl. Math.* **76**, 411 (2016).
- [42] K. Takara, H.-H. Hasegawa, and D. J. Driebe, *Generalization of the Second Law for a Transition between Non-equilibrium States*, *Phys. Lett. A* **375**, 88 (2010).
- [43] M. Esposito and C. Van den Broeck, *Second Law and Landauer Principle Far from Equilibrium*, *Europhys. Lett.* **95**, 40004 (2011).
- [44] T. Sagawa, *Thermodynamics of Information Processing in Small Systems* (Springer, Japan, 2013).

- [45] J. M. R. Parrondo, J. M. Horowitz, and T. Sagawa, *Thermodynamics of Information*, *Nat. Phys.* **11**, 131 (2015).
- [46] M. Esposito, U. Harbola, and S. Mukamel, *Entropy Fluctuation Theorems in Driven Open Systems: Application to Electron Counting Statistics*, *Phys. Rev. E* **76**, 031132 (2007).
- [47] R. J. Harris and G. M. Schütz, *Fluctuation Theorems for Stochastic Dynamics*, *J. Stat. Mech.* (2007) P07020.
- [48] M. Esposito and C. Van den Broeck, *Three Faces of the Second Law. I. Master Equation Formulation*, *Phys. Rev. E* **82**, 011143 (2010).
- [49] C. Van den Broeck and M. Esposito, *Three Faces of the Second Law. II. Fokker-Planck Formulation*, *Phys. Rev. E* **82**, 011144 (2010).
- [50] H. Ge and H. Qian, *Physical Origins of Entropy Production, Free Energy Dissipation, and Their Mathematical Representations*, *Phys. Rev. E* **81**, 051133 (2010).
- [51] H. Ge and H. Qian, *Nonequilibrium Thermodynamic Formalism of Nonlinear Chemical Reaction Systems with Waage-Guldberg's Law of Mass Action*, *Chem. Phys.* **472**, 241 (2016).
- [52] M. Feinberg, "Lecture on Chemical Reaction Networks" (unpublished).
- [53] J. Gunawardena, "Chemical Reaction Network Theory for In Silico Biologists Contents" (unpublished).
- [54] S. R. de Groot and P. Mazur, *Non-Equilibrium Thermodynamics* (Dover, New York, 1984).
- [55] M. Pekař, *Thermodynamics and Foundations of Mass-Action Kinetics*, *Prog. React. Kinet. Mech.* **30**, 3 (2005).
- [56] M. Poletti and M. Esposito, *Irreversible Thermodynamics of Open Chemical Networks. I. Emergent Cycles and Broken Conservation Laws*, *J. Chem. Phys.* **141**, 024117 (2014).
- [57] D. Kondepudi and I. Prigogine, *Modern Thermodynamics: From Heat Engines to Dissipative Structures*, 2nd ed. (Wiley, New York, 2014).
- [58] A. D. McNaught and A. Wilkinson, *Compendium of Chemical Terminology*, 2nd ed. (Blackwell Science, 1997).
- [59] S. Schuster and R. Schuster, *A Generalization of Wegscheider's Condition. Implications for Properties of Steady States and for Quasi-Steady-State Approximation*, *J. Math. Chem.* **3**, 25 (1989).
- [60] J. Leiser and J. J. Blum, *On the Analysis of Substrate Cycles in Large Metabolic Systems*, *Cell Biophysics* **11**, 123 (1987).
- [61] S. Schuster and C. Hilgetag, *On Elementary Flux Modes in Biochemical Reaction Systems at Steady State*, *J. Biol. Syst.* **02**, 165 (1994).
- [62] S. Klamt and J. Stelling, *Two Approaches for Metabolic Pathway Analysis?*, *Trends Biotechnol.* **21**, 64 (2003).
- [63] R. Rao, D. Lacoste, and M. Esposito, *Glucans Monomer-Exchange Dynamics as an Open Chemical Network*, *J. Chem. Phys.* **143**, 244903 (2015).
- [64] F. Horn, *Stability and Complex Balancing in Mass-Action Systems with Three Short Complexes*, *Proc. R. Soc. A* **334**, 331 (1973).
- [65] S. Klamt, U.-U. Haus, and F. Theis, *Hypergraphs and Cellular Networks*, *PLoS Comput. Biol.* **5**, e1000385 (2009).
- [66] C. A. Petri and W. Reisig, *Petri Net*, *Scholarpedia* **3**, 6477 (2008), revision 91646.
- [67] J. C. Baez and B. Fong, *Quantum Techniques for Studying Equilibrium in Reaction Networks*, *J. Complex Netw.* **3**, 22 (2015).
- [68] G. Craciun, *Toric Differential Inclusions and a Proof of the Global Attractor Conjecture*, arXiv:1501.02860.
- [69] A. Dickenstein and M. P. Millán, *How Far is Complex Balancing from Detailed Balancing?*, *Bull. Math. Biol.* **73**, 811 (2011).
- [70] I. Prigogine, *Le Domaine de Validité de la Thermodynamique des Phénomènes Irréversibles*, *Physica A* **15**, 272 (1949).
- [71] E. R. Cohen, T. Cvitas, J. G. Frey, B. Holmström, K. Kuchitsu, R. Marquardt, I. Mills, F. Pavese, M. Quack, J. Stohner, H. L. Strauss, M. Takami, and A. J. Thor, *Quantities, Units and Symbols in Physical Chemistry*, 3rd ed. (IUPAC and RSC Publishing, 2008).
- [72] R. Holyst and A. Poniewierski, *Thermodynamics for Chemists, Physicists and Engineers* (Springer, New York, 2012).
- [73] D. Shear, *An Analog of the Boltzmann H-Theorem (a Liapunov Function) for Systems of Coupled Chemical Reactions*, *J. Theor. Biol.* **16**, 212 (1967).
- [74] J. Higgins, *Some Remarks on Shear's Liapunov Function for Systems of Chemical Reactions*, *J. Theor. Biol.* **21**, 293 (1968).
- [75] T. M. Cover and J. A. Thomas, *Elements of Information Theory*, 2nd ed. (Wiley-Interscience, New York, 2006).
- [76] A. van der Schaft, S. Rao, and B. Jayawardhana, *On the Mathematical Structure of Balanced Chemical Reaction Networks Governed by Mass Action Kinetics*, *SIAM J. Appl. Math.* **73**, 953 (2013).
- [77] I. Procaccia and R. D. Levine, *Potential Work: A Statistical-Mechanical Approach for Systems in Disequilibrium*, *J. Chem. Phys.* **65**, 3357 (1976).
- [78] S. Vaikuntanathan and C. Jarzynski, *Dissipation and Lag in Irreversible Processes*, *Europhys. Lett.* **87**, 60005 (2009).
- [79] H.-H. Hasegawa, J. Ishikawa, K. Takara, and D. J. Driebe, *Generalization of the Second Law for a Nonequilibrium Initial State*, *Phys. Lett. A* **374**, 1001 (2010).
- [80] M. Gopalkrishnan, *On the Lyapunov Function for Complex-Balanced Mass-Action Systems*, arXiv:1312.3043.
- [81] A. van der Schaft, S. Rao, and B. Jayawardhana, *Complex and Detailed Balancing of Chemical Reaction Networks Revisited*, *J. Math. Chem.* **53**, 1445 (2015).
- [82] The fact that the matrix whose entries are $\{\ell_{\sigma_i}^{\lambda_u}\}$ is nonsingular follows from $\ell^{\lambda_u} \in \ker \nabla$, $\forall \lambda_u$, and from the linear independence of $\{\ell^{\lambda}\}$.
- [83] M. Esposito and G. Schaller, *Stochastic Thermodynamics for "Maxwell Demon" Feedbacks*, *Europhys. Lett.* **99**, 30003 (2012).
- [84] G. Diana, G. B. Bageci, and M. Esposito, *Finite-Time Erasing of Information Stored in Fermionic Bits*, *Phys. Rev. E* **87**, 012111 (2013).
- [85] J. M. Horowitz and M. Esposito, *Thermodynamics with Continuous Information Flow*, *Phys. Rev. X* **4**, 031015 (2014).

- [86] A. C. Barato and U. Seifert, *Unifying Three Perspectives on Information Processing in Stochastic Thermodynamics*, *Phys. Rev. Lett.* **112**, 090601 (2014).
- [87] S. Bo, M. Del Giudice, and A. Celani, *Thermodynamic Limits to Information Harvesting by Sensory Systems*, *J. Stat. Mech.* (2015) P01014.
- [88] P. Strasberg, G. Schaller, T. Brandes, and M. Esposito, *Thermodynamics of a Physical Model Implementing a Maxwell Demon*, *Phys. Rev. Lett.* **110**, 040601 (2013).
- [89] N. Golubeva and A. Imparato, *Efficiency at Maximum Power of Interacting Molecular Machines*, *Phys. Rev. Lett.* **109**, 190602 (2012).
- [90] B. Altaner, A. Wachtel, and J. Vollmer, *Fluctuating Currents in Stochastic Thermodynamics. II. Energy Conversion and Nonequilibrium Response in Kinesin Models*, *Phys. Rev. E* **92**, 042133 (2015).
- [91] U. Seifert, *Stochastic Thermodynamics of Single Enzymes and Molecular Motors*, *Eur. Phys. J. E* **34**, 26 (2011).
- [92] R. Rao and L. Peliti, *Thermodynamics of Accuracy in Kinetic Proofreading: Dissipation and Efficiency Trade-Offs*, *J. Stat. Mech.* (2015) P06001.
- [93] J. J. Hopfield, *Kinetic Proofreading: A New Mechanism for Reducing Errors in Biosynthetic Processes Requiring High Specificity*, *Proc. Natl. Acad. Sci. U.S.A.* **71**, 4135 (1974).
- [94] J. Ninio, *Kinetic Amplification of Enzyme Discrimination*, *Biochimie* **57**, 587 (1975).
- [95] J. J. Hopfield, *The Energy Relay: A Proofreading Scheme Based on Dynamic Cooperativity and Lacking All Characteristic Symptoms of Kinetic Proofreading in DNA Replication and Protein Synthesis*, *Proc. Natl. Acad. Sci. U.S.A.* **77**, 5248 (1980).
- [96] E. D. Sontag, *Structure and Stability of Certain Chemical Networks and Applications to the Kinetic Proofreading Model of T-Cell Receptor Signal Transduction*, *IEEE Trans. Autom. Control* **46**, 1028 (2001).
- [97] H. Ge, M. Qian, and H. Qian, *Stochastic Theory of Nonequilibrium Steady States. Part II: Applications in Chemical Biophysics*, *Phys. Rep.* **510**, 87 (2012).
- [98] A. Murugan, D. H. Huse, and S. Leibler, *Speed, Dissipation, and Error in Kinetic Proofreading*, *Proc. Natl. Acad. Sci. U.S.A.* **109**, 12034 (2012).
- [99] A. Murugan, D. H. Huse, and S. Leibler, *Discriminatory Regimes in Non-Equilibrium Systems*, *Phys. Rev. X* **4**, 021016 (2014).
- [100] C. H. Bennett, *Dissipation-Error Tradeoff in Proofreading*, *BioSystems* **11**, 85 (1979).
- [101] D. Andrieux and P. Gaspard, *Nonequilibrium Generation of Information in Copolymerization Processes*, *Proc. Natl. Acad. Sci. U.S.A.* **105**, 9516 (2008).
- [102] D. Andrieux and P. Gaspard, *Molecular Information Processing in Nonequilibrium Copolymerizations*, *J. Chem. Phys.* **130**, 014901 (2009).
- [103] J. R. Arias-Gonzalez, *Entropy Involved in Fidelity of DNA Replication*, *PLoS One* **7**, e42272 (2012).
- [104] P. Sartori and S. Pigolotti, *Kinetic versus Energetic Discrimination in Biological Copying*, *Phys. Rev. Lett.* **110**, 188101 (2013).
- [105] P. Sartori and S. Pigolotti, *Thermodynamics of Error Correction*, *Phys. Rev. X* **5**, 041039 (2015).
- [106] A. Cornish-Bowden, *Fundamentals of Enzyme Kinetics*, 4th ed. (Wiley-Blackwell, 2012).
- [107] M. Esposito, *Stochastic Thermodynamics under Coarse Graining*, *Phys. Rev. E* **85**, 041125 (2012).
- [108] C. L. Linster, E. van Schaftingen, and A. D. Hanson, *Metabolite Damage and Its Repair or Pre-emption*, *Nat. Chem. Biol.* **9**, 72 (2013).
- [109] H. Qian and T. C. Reluga, *Nonequilibrium Thermodynamics and Nonlinear Kinetics in a Cellular Signaling Switch*, *Phys. Rev. Lett.* **94**, 028101 (2005).
- [110] H. Qian and S. Roy, *An Information Theoretical Analysis of Kinase Activated Phosphorylation Dephosphorylation Cycle*, *IEEE Trans. Nanobiosci.* **11**, 289 (2012).
- [111] J. M. G. Vilar, H. Y. Kueh, N. Barkai, and S. Leibler, *Mechanisms of Noise-Resistance in Genetic Oscillators*, *Proc. Natl. Acad. Sci. U.S.A.* **99**, 5988 (2002).
- [112] G. Nicolis and I. Prigogine, *Self-Organization in Nonequilibrium Systems: From Dissipative Structures to Order Through Fluctuations* (Wiley-Blackwell, 1977).
- [113] R. Feistel and W. Ebeling, *Physics of Self-Organization and Evolution* (Wiley VCH, 2011).
- [114] P. Mehta and D. J. Schwab, *Energetic Costs of Cellular Computation*, *Proc. Natl. Acad. Sci. U.S.A.* **109**, 17978 (2012).
- [115] P. Sartori, L. Granger, C. F. Lee, and J. M. Horowitz, *Thermodynamic Costs of Information Processing in Sensory Adaptation*, *PLoS Comput. Biol.* **10**, e1003974 (2014).
- [116] D. Hartich, A. C. Barato, and U. Seifert, *Nonequilibrium Sensing and Its Analogy to Kinetic Proofreading*, *New J. Phys.* **17**, 055026 (2015).
- [117] S.-W. Wang, Y. Lan, and L.-H. Tang, *Energy Dissipation in an Adaptive Molecular Circuit*, *J. Stat. Mech.* (2015) P07025.
- [118] *DNA Computing and Molecular Programming*, edited by Y. Sakakibara and Y. Y. Mi (Springer, Berlin, 2011); *DNA Computing and Molecular Programming*, edited by Luca Cardelli and W. Shih (Springer, Berlin, 2011); *DNA Computing and Molecular Programming*, edited by D. Stefanovic and A. Turberfields (Springer, Berlin, 2012); *DNA Computing and Molecular Programming*, edited by D. Soloveichik and B. Yurke (Springer International Publishing, 2013); *DNA Computing and Molecular Programming*, edited by S. Murata and S. Kobayashi (Springer International Publishing, 2014); *DNA Computing and Molecular Programming*, edited by A. Phillips and P. Yin (Springer International Publishing, 2015).
- [119] A. Hjelmfelt, E. D. Weinberger, and J. Ross, *Chemical Implementation of Neural Networks and Turing Machines*, *Proc. Natl. Acad. Sci. U.S.A.* **88**, 10983 (1991).
- [120] A. Hjelmfelt, E. D. Weinberger, and J. Ross, *Chemical Implementation of Finite-State Machines*, *Proc. Natl. Acad. Sci. U.S.A.* **89**, 383 (1992).
- [121] M. O. Magnasco, *Chemical Kinetics is Turing Universal*, *Phys. Rev. Lett.* **78**, 1190 (1997).
- [122] D. Soloveichik, M. Cook, E. Winfree, and J. Bruck, *Computation with Finite Stochastic Chemical Reaction Networks*, *Nat. Comput.* **7**, 615 (2008).
- [123] C. H. Bennett, *The Thermodynamics of Computation—A Review*, *Int. J. Theor. Phys.* **21**, 905 (1982).

- [124] T. P. J. Knowles, W. Shu, G. L. Devlin, S. Meehan, S. Auer, C. M. Dobson, and M. E. Welland, *Kinetics and Thermodynamics of Amyloid Formation from Direct Measurements of Fluctuations in Fibril Mass*, *Proc. Natl. Acad. Sci. U.S.A.* **104**, 10016 (2007).
- [125] T. P. J. Knowles, C. A. Waudby, G. L. Devlin, S. I. A. Cohen, A. Aguzzi, M. Vendruscolo, E. M. Terentjev, M. E. Welland, and C. M. Dobson, *An Analytical Solution to the Kinetics of Breakable Filament Assembly*, *Science* **326**, 1533 (2009).
- [126] S. I. A. Cohen, S. Linse, L. M. Luheshi, E. Hellstrand, D. A. White, L. Rajah, D. E. Otzen, M. Vendruscolo, C. M. Dobson, and T. P. J. Knowles, *Proliferation of Amyloid- β 42 Aggregates Occurs through a Secondary Nucleation Mechanism*, *Proc. Natl. Acad. Sci. U.S.A.* **110**, 9758 (2013).
- [127] Z. Budrikis, G. Costantini, C. A. M. La Porta, and S. Zapperi, *Protein Accumulation in the Endoplasmic Reticulum as a Non-Equilibrium Phase Transition*, *Nat. Commun.* **5**, 3620 (2014).
- [128] Ö. Kartal, S. Mahlow, A. Skupin, and O. Ebenhöf, *Carbohydrate-Active Enzymes Exemplify Entropic Principles in Metabolism*, *Mol. Syst. Biol.* **7**, 542 (2011).
- [129] S. Lahiri, Y. Wang, M. Esposito, and D. Lacoste, *Kinetics and Thermodynamics of Reversible Polymerization in Closed Systems*, *New J. Phys.* **17**, 085008 (2015).
- [130] P. Krapivsky, S. Redner, and E. Ben-Naim, *A Kinetic View of Statistical Physics* (Cambridge University Press, Cambridge, England, 2010).
- [131] Enrico Fermi, *Thermodynamics* (Dover, New York, 1956).
- [132] H. Ge and H. Qian, *Mesosopic Kinetic Basis of Macroscopic Chemical Thermodynamics: A Mathematical Theory*, *Phys. Rev. E* **94**, 052150 (2016).
- [133] E. T. Jaynes, *Information Theory and Statistical Mechanics*, *Phys. Rev.* **106**, 620 (1957).